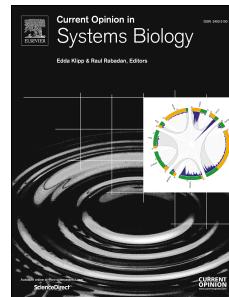


Journal Pre-proof

Systems Biology at the giga-scale: large multi-scale models of complex, heterogeneous multicellular systems

Arnau Montagud, Miguel Ponce de León, Alfonso Valencia



PII: S2452-3100(21)00079-2

DOI: <https://doi.org/10.1016/j.coisb.2021.100385>

Reference: COISB 100385

To appear in: *Current Opinion in Systems Biology*

Received Date: 21 May 2021

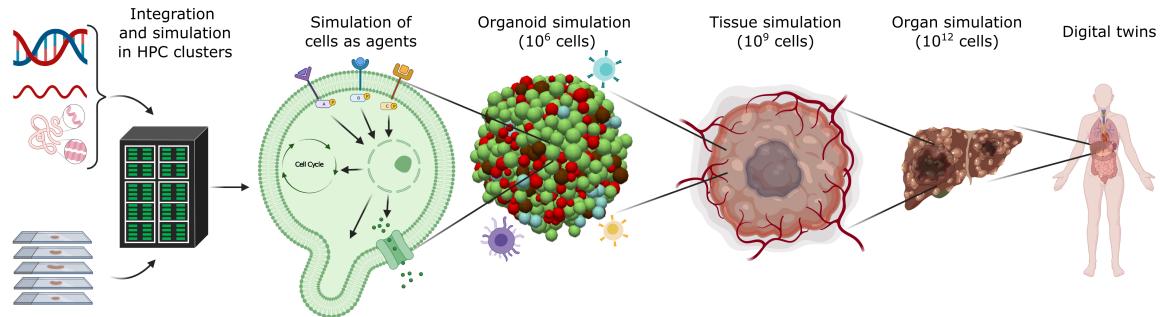
Revised Date: 16 August 2021

Accepted Date: 15 September 2021

Please cite this article as: Montagud A, Ponce de León M, Valencia A, Systems Biology at the giga-scale: large multi-scale models of complex, heterogeneous multicellular systems, *Current Opinion in Systems Biology*, <https://doi.org/10.1016/j.coisb.2021.100385>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2021 The Author(s). Published by Elsevier Ltd.



Journal Pre-proof

1 Systems Biology at the giga-scale: large multi-scale models of complex, heterogeneous
2 multicellular systems

3

4 Arnau Montagud^{1,*}, Miguel Ponce de León¹, Alfonso Valencia^{1,2,*}

5 1, Barcelona Supercomputing Center (BSC), Barcelona, Spain

6 2, Institució Catalana de Recerca i Estudis Avançats (ICREA), Barcelona, Spain

7 *: corresponding authors: arnau.montagud@bsc.es, alfonso.valencia@bsc.es

8

9 Abstract:

10 Agent-based modelling has proven its usefulness in several biomedical projects by
11 explaining and uncovering mechanisms in diseases. Nevertheless, the scenarios addressed
12 in these models usually consider a small number of cells, lack cell-specific characterisation
13 and dynamic interactions and have a simplistic environment description. Tools that enable
14 scalable, real-sized simulations of biological systems that require complex set-ups are
15 needed to have simulations closer to biomedical scenarios that can capture cell-to-cell
16 heterogeneity and system-wide emerging properties. To deliver simulations at the giga-scale
17 (10^9 cells), different tools have implemented technologies to run in high-performance
18 computing clusters. We hereby review these efforts and detail the main areas of
19 improvement the field needs to focus on to have simulations that are a step closer to having
20 digital twins.

21

22 Keywords:

23 Agent-based simulations, large-scale simulations, high-performance computing, model
24 exploration, cancer biology

25

26 Highlights:

- 27 ● Agent-based modelling aims to explain, model and predict mechanisms in diseases.
- 28 ● Large-scale agent-based modelling is needed to address real-sized, complex
29 scenarios such as simulating tumours.
- 30 ● The scaling-up of such models needs high-performance infrastructures and methods
31 to optimise runtime and distribute memory among computing nodes.
- 32 ● Such multi-scale models are complex and need tools such as model exploration
33 frameworks to help set-up parameters and provide new hypotheses.
- 34 ● Bringing together the systems biology and HPC communities is key for the
35 development of the field.

36 **Introduction:**

37 Modelling has helped researchers gain unprecedented insights into biological mechanisms
 38 [1]. With the rise of high-throughput technologies that led to the so-called omic revolution,
 39 the use of modelling in systems biology has become an integral part of the field [2,3].
 40 Bridging from intracellular mechanisms to tissue-level phenotype is still a problem to be
 41 solved, and multi-scale agent-based modelling is well suited to address it. Agent-based
 42 modelling is an approach to computing the potential system-level consequences of the
 43 behaviours of sets of individual agents [4]. In this framework, also known as individual-cell,
 44 individual-based or cell-based modelling, each agent has a set of behavioural rules that are
 45 relative to the environment and to other agents' behaviours. Agent-based models have been
 46 ubiquitous in a wide range of scientific disciplines, such as economics, engineering,
 47 epidemiology, urban transportation, ecology, and biology [5].

48 Agent-based models have a balanced combination that suits well systems biology modelling
 49 of tissues by, first, combining the study of cell populations' behaviours that have a direct
 50 correspondence to heterogeneous cellular systems encompassing a diversity of cell-cell
 51 interactions. Second, they have a cell-level granularity that allows studying genotypic and
 52 phenotypic variations at the single-cell level. Third, they can include intracellular models that
 53 can capture gene-level changes and, fourth, they explicitly describe the environment that
 54 can have a direct correspondence to laboratory experiments (Figure 1). Agent-based models
 55 have proven their usefulness in biomedicine by studying the selective pressure of the
 56 environment on tumour morphology [6], migrating cells [7–9] and their interaction with
 57 macrophages to suppress immune responses [10–12], cancer growth and how
 58 immunogenicity enable tumour cells at the outskirts to escape immune attack [13], or how
 59 dynamic regimes can counter the tumour cells' resistance to TNF [14]. Recently, some of
 60 these approaches have been directed at the study of COVID-19 [15].

61 **Agent-based includes different modelling frameworks with
 62 different focuses**

63 Different agent-based modelling approaches exist to model cell populations with different
 64 underlying assumptions and constraints. Still, they are usually grouped into two families:
 65 lattice-based (or on-lattice) and lattice-free (or off-lattice) (Figure 2A). For complete reviews,
 66 see references [16,17].

67 Lattice-based agent-based models are the ones where the environment is defined by a grid
 68 or lattice, which the agents will follow or be constrained by it. This type of modelling can be
 69 categorised by three different subtypes depending on their spatial resolution:

- 70 ● Cellular automata, in which there is one agent in each lattice site (e.g., NetLogo [18],
 71 Chaste [19], and others [20–22]).
- 72 ● Cellular Potts model, in which one agent can occupy several lattice sites [23] (e.g.,
 73 CompuCell3D [24], Morpheus [25], EPISIM [26], and others [27]).
- 74 ● A third type in which a lattice site can be occupied by more than one agent (e.g.,
 75 COMETS [28,29], and others [30,31]). Lattice-gas cellular automata is a special such
 76 automaton in which the velocity of the agents is also tracked (e.g., NetLogo).

77 Lattice-free agent-based models are the ones where the environment is not defined by a grid
 78 or lattice. This type of modelling can be categorised into two main families of models,
 79 depending on whether the model's focus is the cells' boundaries or the cells' volume:

- 80 ● Vertex models are boundary-tracking models where cells are modelled as polygons
 81 or polyhedra, and the focus of the model is to study the different forces applied to the
 82 vertices (e.g., Chaste, Tyssue [32], and reference [33]). They can also use Voronoi
 83 tessellation (e.g., Chaste and reference [34]).
- 84 ● Overlapping spheres or centre-based models (CBM) are models where the agents
 85 are represented by spheres that can overlap among them (e.g., Chaste, Biocellion
 86 [35], CellSys [36], PhysiCell [37], Timothy [38]).

87 When modelling in the context of biomedical research, one of the goals is to find molecular
 88 mechanisms and interactions that may explain the observed disease phenotypes. Agent-
 89 based modelling is general and flexible enough to be the basis for constructing multi-scale
 90 models of cell populations that include intracellular models of different processes updated at
 91 different time scales when reacting to environmental conditions. This intracellular model can
 92 be a system of differential equations (as in Chaste, CompuCell3D, and EPISIM), neural
 93 networks [20], metabolic models of bacteria using dynamic FBA in BacArena [22] or
 94 COMETS [29] or Boolean models simulated stochastically using PhysiBoSS [14] (Figure
 95 2B).

96 These CBM frameworks consider different time scales for simulating different processes
 97 such as molecules' diffusion, cell movement and mechanical interactions as well as cellular
 98 phenotypes (e.g., proliferation, apoptosis, migration, attachment) (Figure 2C) following a
 99 type 3 multi-scale model taxonomy from reference [39]. In addition, they consider different
 100 types of cells (e.g., immune cells, blood vessels and cancer tissue) and dynamic
 101 environments. Still, they usually take into account up to several million cells, which is far
 102 from the goal of having large multi-scale simulations of complex, heterogeneous tumours.

103 There are several alternatives to agent-based modelling. One of them, rule-based modelling,
 104 is a flexible space-free approach used in a wide range of fields, including biology, physics,
 105 chemistry, and computer science [40]. Notably, it has been successful in biochemistry and
 106 cell biology projects by simulating agents not as cells but as proteins and molecular
 107 complexes in well-mixed spatial compartments [41]. This approach has been combined with
 108 detailed spatial representations (e.g., the integration of BioNetGen [42] with Smoldyn [43] or
 109 VCell [44]) and is well suited to simulate a large number of biochemical reactions [45].
 110 Interestingly, there have been efforts to use rule-based modelling to simulate cells and
 111 tissues [46].

112 Another alternative to agent-based modelling, hybrid discrete-continuum modelling, is useful
 113 when aiming to model large multi-cellular systems as continuum models without explicitly
 114 defining individual cells, their environment or their intrinsic scales [47]. Even though
 115 continuum models generally have fewer parameters and are less explicit than CBM, they
 116 can be useful to uncover insights into biological mechanisms [48].

117 These methods represent different approaches to biomedical problems and correspond to
 118 different assumptions and modelling decisions. The rest of this review will focus on CBM, a
 119 type of agent-based modelling, as we consider that they have a balanced combination of
 120 variables and assumptions that suits well tissue and cancer modelling scenarios making it

121 possible to model genes, pathways, cells and environment explicitly. In this case, as the
 122 models include genes and proteins, we can directly integrate available omics data on them
 123 and, as the cells are simulated individually, they can deliver biologically realistic simulations
 124 that have a direct correlation to clinical images.

125 Motivation for a distributed simulation at the giga-scale

126 Historically, there have been a number of efforts to build realistic surrogates of human tissue
 127 with increasing importance in personalised medicine [49]. The development of humanised
 128 patient-derived xenografts [50] and digital twins [51] are just two recent examples in
 129 biomedicine. To bring these models closer to clinical scenarios, e.g., capable of simulating
 130 the evolution of clinically detectable tumours, we still need more powerful large-scale
 131 modelling tools able to produce more realistic, real-sized simulations [52].

132 Giga-scale simulations should be able to address questions that current agent-based, as
 133 well as other multi-scale models hardly, can, such as those related to the interaction
 134 between processes and mechanisms modelled at different time and spatial scales
 135 [14,53,54]. A prototypical example of a very complex giga-scale simulation would be a
 136 realistic cancer response to different chemo- and immune therapies, able to explore the
 137 influence of tumour shape and environment interactions. This type of model would provide
 138 "virtual cuts" images comparable to digital pathology images of real tumours. The set-up
 139 needed to perform these simulations requires the knowledge of signalling pathways'
 140 activities, population-level biophysics, as well as the effect of the extracellular matrix and
 141 immune system on the diffusive properties of different molecules. Most of these mechanisms
 142 have been studied separately, their mechanisms have been documented in different
 143 situations, and a number of useful models build by experts are available [9,11,55,56], but the
 144 explanations of system-wide emerging properties require addressing the challenging
 145 problem of putting all of them together. The TNF dynamic regimes and cells' resistance [14]
 146 represents an initial example in this direction.

147 With the current state-of-the-art methods, we are unable to study the aforementioned
 148 prototypical example: a real-sized invasive tumour of an irregular shape that is being
 149 targeted by drugs arriving from a distant, single blood vessel surrounded by a complex
 150 extracellular matrix with patches of different densities that affect drug diffusion and
 151 movement of the cells. A new complex set-up will be required to test the shielding effect that
 152 the shape of the tumour might have against drugs and immune cells. Also, in regards to the
 153 invasive potential of the cells, current simulations are insufficient to describe the
 154 heterogeneous biological scenarios [57]. With this simulation, it will be possible to test the
 155 effect of an extracellular matrix with different densities together with the cellular
 156 heterogeneity on the different invasion phenotypes as described in clinical images. In
 157 addition, models that include studies of tumour evolution and genetic drifts might be able to
 158 capture behaviours related to the appearance of heterogeneity in otherwise clonal
 159 populations that might be missed in smaller simulations [58].

160 Another system-wide emerging property that could be studied with these giga-scale
 161 simulations is drug resistance against cancer therapies. It has been described that
 162 population-level dynamics such as competition and cell-cell variability can play a key role in
 163 the evolution of drug resistance [59]. In addition, the environment architecture has been
 164 described to affect the cells' response to drugs: conventional 2D-cultured cell line screens

165 fail in clinical studies [60] as traditional cell cultures do not recapitulate the heterogeneity and
 166 intrinsic drug sensitivity of the original tumour [61].

167 In our opinion, there are three main bottlenecks to achieve such a real-sized tumour
 168 simulation: biological knowledge, data availability and technical performance of the tools.
 169 Our knowledge of biological processes of different time and spatial scales and, importantly,
 170 their interactions is far from perfect. In fact, one of the goals of modelling is to help to close
 171 this knowledge gap by mapping what is known and providing hypotheses to explore the
 172 more relevant elements of what is still unknown.

173 In addition, the new technologies provide a unique opportunity to extend and improve these
 174 comprehensive models. In this sense, the new developments in the field of single-cell omics
 175 provide a unique opportunity to fill models with real data [62], connecting cell heterogeneity
 176 with population dynamics [63].

177 Finally, modelling these complex scenarios needs substantial computing resources that far
 178 exceed current desktop and university clusters. Particularly important are the limitations on
 179 the use and management of the memory for the data structures [38,64]. Thus, the scale-up
 180 of the simulations is only achievable by optimal, full-scale use of parallel systems in HPC
 181 systems [38,54]. While the access to additional biological knowledge and heterogeneous
 182 data is continuously evolving, the HPC remains the key enabling technology for the giga-
 183 scale simulations.

184 Paving the way for a distributed simulation at the giga-scale

185 In the past years, efforts have been directed at having large-scale CBM simulations by
 186 distributing the computation following a hybrid model combining MPI and OpenMP. We
 187 hereby present several centre-based agent-based modelling frameworks that have the
 188 potential to reach the billion cells' milestone.

189 Timothy [38,65] is an open-source, MPI-based tool that has proven to be able to simulate
 190 biological models of up to 0.3 billion cells in cancer projects [66] as well as large-scale
 191 models with nuclear-cytoplasmic oscillations of NF-κB [53].

192 Biocellion [35] is a flexible, discrete agent-based simulation framework that has been used to
 193 model a wide range of multi-cellular biological models, such as cell sorting, microbial
 194 patterning and a bacterial system in soil aggregate of over 26 million cells. Even though it's
 195 freely available for academic use, Biocellion's closed source can deter potential users.

196 Chaste [19] is an open-source, general-purpose simulation package for modelling soft
 197 tissues and discrete cell populations that can be used with MPI. This tool allows using
 198 different modelling frameworks on a given problem, enabling users to select the most
 199 appropriate one for their research and to better understand the limitations of each one of
 200 them. Chaste has been used for a wide range of projects, such as intestinal [67] or colonic
 201 crypt [68] studies.

202 FLAME [69] is an open-source, generic formal framework for agent-based modelling that
 203 allows parallelisation using MPI, and developers can use it to create models and tools.
 204 Implementing a Flame simulation is done by using finite-state automata with memory [70].
 205 This tool has been adapted to be used with distributed GPUs using the OpenCL standard

206 [71]. Examples of uses of FLAME range from bioreactor studies [72] to immunogenic studies
 207 [73] or also epidermis modelling [74].
 208 To study biofilm-scale bacterial population, CellModeller [75] was developed as a rigid-body
 209 method that includes models of biophysics, growth of rod-shaped cells and intra- and inter-
 210 cellular signalling using ODEs. CellModeller uses OpenCL cross-platform framework that
 211 enables the implementation of parallel software on both GPU and CPU architectures.
 212 BioDynaMo [76] is an open-source simulation tool fully parallelised, able to offload
 213 computation to hardware accelerators and load-balance agents and their environment on
 214 available nodes. Being general-purpose, it allows simulating models from various fields by
 215 being extensible and modular, showcasing its use in neurite growth, tumour spheroid and
 216 epidemiology [76].
 217 PhysiCell-X (Saxena et al., in preparation) is our current effort to include MPI in PhysiCell,
 218 an open-source, flexible, and lattice-free agent-based framework for multi-scale simulation of
 219 multi-cellular systems that currently only support shared-memory parallelisation. The main
 220 advantage of PhysiCell is its lightweight, very efficient and self-contained framework.
 221 Additionally, PhysiCell can be expanded using add-ons, such as PhysiBoSS [14], allowing
 222 the integration of individual Boolean models for the signalling networks embedded into each
 223 agent. PhysiCell-X expands our efforts to include distributed-memory in the solver that
 224 manages the chemicals' diffusion of PhysiCell: BioFVM-X [64].
 225 We have hereby focused on examples of distributed CBM software. Still, other agent-based
 226 tools use distributed memory and have the potential of enabling giga-scale simulations that
 227 are not CBM [21,25,26,77–80].

228 Wish list for large-scale multi-scale modelling tools

229 Many problems addressed by modelling in biology can consider populations of cells ranging
 230 from 10^6 to 10^{11} [81], and CBM tools from the last section allow to reach only the lower
 231 bound of this range. To have simulations at the giga-scale (10^9 cells), we need novel tools to
 232 scale up efficiently by incorporating technologies from biology and computer science (Figure
 233 3).
 234 Nevertheless, having CBM simulations with large numbers of cells is not the only problem,
 235 as the underlying models need to be more realistic and complex. We need novel tools that
 236 can simulate real-size tumours of billions of cells, where there are explicit descriptions of
 237 intracellular mechanisms (i.e., signalling pathways, metabolism, cell cycle, cell division, etc.),
 238 each cell has its idiosyncratic properties without systematic regularity, where this
 239 idiosyncrasy changes in time, e.g., by the effect of selective pressure, and where we can
 240 study the clonal heterogeneities of cells and thus the evolution of cell strains. Such a tool
 241 should include, among others, an explicit simulation of the interactions among the different
 242 cells types, different cell shapes that better capture the tensions around a cell, and a
 243 complete description of a complex 3D environment with blood vessels (ideally with clinical-
 244 image-level detail) and cocktails of chemicals (e.g., hormones, metabolites, oxygen, etc.)
 245 that allows for the cells to modify it and be modified by it.
 246 These complex simulations should be facilitated by enabling technologies in high-
 247 performance computing (HPC) clusters that allow having distributed memory and

248 parallelisation, analysis and integration of high-throughput personal molecular data in the
 249 models and, model exploration and optimisation of the models' parameters.

250 There are already tools that perform some of the goals described in this wish list, such as
 251 those in the previous section, but no one can do it all, let alone at a large scale.

252 Model exploration is needed to fit and to find optimised 253 solutions

254 Agent-based models such as CBM are complex objects that can exhibit nonlinear dynamics
 255 and emergent behaviours [13]. Thus, the effect of the input parameters and the model
 256 dynamics can only be addressed by running simulations where stochasticity is an intrinsic
 257 component of the model. Furthermore, complex multi-scale CBM usually have a large
 258 number of parameters that should be specified. In the case of multi-cellular models, such
 259 parameters may involve, among others, rates of cell processes (e.g., division, death), kinetic
 260 constants of the biochemical processes (e.g., reactions, transports), mechanical constants
 261 (e.g., cell-cell interactions, migration velocities) as well as physicochemical properties of the
 262 environment (e.g., diffusion constants, extracellular matrix properties). In many cases, a
 263 model can have so many parameters that the data required for its calibration and validation
 264 is currently not available [82,83].

265 The analysis of complex CBM requires running large numbers of simulations applying
 266 efficient strategies to explore the parameter spaces [84,85]. This general problem is
 267 commonly referred to as model exploration (ME), a multifaceted iterative process that
 268 enables an adaptive exploration of the model's parameter space. ME strategies are
 269 commonly applied to calibration, optimisation and exploration of models and usually rely on
 270 metaheuristics (e.g., evolutionary algorithm, simulated annealing) to optimise "black box"
 271 functions or machine learning approaches to characterise the parameter space [86,87].

272 Model calibration is the task of estimating the unknown parameters using experimental data
 273 and was successfully used in a tumoural CBM by comparing simulated growth dynamics and
 274 spatio-temporal data extracted from experimental image analysis [88]. In model optimisation,
 275 also called simulation-based optimisation or optimisation via simulation [89], simulations are
 276 run to predict the values of the parameters that allow achieving the desired goal. For
 277 instance, model optimisation was used with a genetic algorithm to identify the set of
 278 biomedical interventions needed to minimise the number of cancer cells in a CBM of
 279 tumoural immunosurveillance [90] or to predict the drug dose schedule that minimises the
 280 tumour size while avoiding the emergence of resistant cells [91].

281 As discussed before, the complexity in CBM can come from a large number of cells or by
 282 having a more complex environment. As the complexity of multi-scale CBM increases, the
 283 simulations become computationally-intensive tasks, and thus ME can become challenging.
 284 In such scenarios, ME strategies should be implemented as distributed workflows to be
 285 executed in HPC clusters [13,85] in a "many-task computing" paradigm [92]. There are
 286 several frameworks for distributed ME on HPC clusters [93], including EMEWS [94],
 287 OpenMOLE [95] and ECJ [96]. Nonetheless, besides the great improvement in having HPC-
 288 ready ME strategies, the calibration and estimation of many model parameters will require
 289 novel high-throughput experimental data at the single-cell level [62,97].

290 Alternatively, there have been efforts to leverage the trove of molecular omics data available
 291 to parametrise multi-scale models by using machine learning methodologies. To accomplish
 292 this, machine learning and multi-scale modelling can complement each other on the
 293 parameter level, e.g. by identifying parameter values [98], and on the system level, e.g. by
 294 identifying system dynamics [99]. Machine learning can be useful by providing tools towards
 295 preventing overfitting or quantifying uncertainty while exploring huge parameter spaces (for a
 296 review, see reference [100]).

297 Perspectives

298 Modelling diseases has the potential of being an enabling technology in helping deliver truly
 299 personalised and precise medicine to the patients. For instance, leveraging single-cell
 300 resolution clinical images [101,102] and using personalised intracellular models with
 301 patients' data are initial steps in achieving patient-tailored drug regimes [91,103,104].
 302 Nevertheless, researchers need to provide tools that allow for more realistic and complex
 303 simulations that include, among others, dynamically responsive environments and
 304 simulations' scales of clinically detectable tumour sizes, i.e. from the giga-scale up.
 305 We have reviewed efforts directed at having such CBM tools that aim to model biological
 306 phenotypes in HPC clusters. Previously, we have described two improvements that would
 307 increase the relevance of these giga-scale simulations: the biological knowledge on the
 308 interaction of mechanisms at different time and spatial scales and the experimental data that
 309 could provide single-cell heterogeneity [55]. Nevertheless, some additional challenges need
 310 to be solved to have simulations that bring the field closer to having digital twins [51] (Figure
 311 1 and 3).
 312 One of them is to extend the cell-level CBM simulations to reach molecular dynamics and
 313 tissue simulations. There have been efforts in using CBM to reach down to molecular
 314 dynamics [54], to study the effect of molecular crowding on a virtual cytoplasm [105] or to
 315 focus on mRNA export mechanisms [106]. Likewise, CBM have been used to reach up to
 316 tissue modelling to study muscle regeneration [107] or combining finite elements with agents
 317 to study glioma invasion and vascularity [108]. These extensions that integrate models with
 318 very different granularities have their limits and challenges, and to address them, the field
 319 could learn from lessons from the whole-cell modelling community [83,97,109–111].
 320 Another challenge is to use tools that deal with the complexity of the model set-up, identified
 321 as one of the main obstacles for agent-based modelling use [54]. In this line, ME methods
 322 have been useful to set-up, fit and explore models [90]. In addition, the use of common
 323 languages and standards formats, such as SBML [112], MultiCellIDS [113], or OpenABL
 324 [114], could increase the complexity of the models by encapsulating its minor details and
 325 ease the comparison, reproducibility and benchmarking of models [115].
 326 Finally, all these efforts need to be scalable and efficient to take full advantage of HPC
 327 clusters; thus, we need to bring closer the systems biology and HPC communities, following
 328 recent efforts such as the PerMEDCoE centre of excellence (<https://permedcoe.eu/>).

329 Acknowledgements:

330 The authors acknowledge the reviewers for their comments and suggestions that helped
 331 improve and clarify this manuscript.

332 This work has been partially supported by the European Commission under the INFORE
 333 project (H2020-ICT-825070) and the PerMedCoE project (H2020-ICT-951773).

334 **References' comments:**

335 Papers of particular interest, published within the period of review, have been highlighted as:

336 * of special interest

337 ** of outstanding interest

338

339 Macal & North, 2010, [5] *: Fundamental primer on agent-based modelling that presents the
 340 main concepts and some applications across disciplines.

341 Spatarelu et al., 2019, [9] *: Review on experimental and computational methods collective
 342 cell migration in cancer and how agent-based methods have helped gain insights into
 343 mechanisms.

344 Reticker-Flynn et al., 2020, [11] *: Review on the field of cancer systems immunology where
 345 the authors discuss the advances done by agent-based modelling and address the future
 346 avenues of study.

347 Norton et al., 2019, [12] *: Review on agent-based modelling of the immune
 348 microenvironment surrounding tumours and how different modelling approaches are best
 349 suited for different spatial scales.

350 Metzcar et al., 2019, [16] **: Comprehensive review on agent-based modelling, their different
 351 types and examples of its use in cancer biology.

352 Cooper et al., 2020, [19] *: Chaste is a simulation platform that allows users to test their
 353 models in three different agent-based modelling approaches (e.g., cellular automata, vertex-
 354 based, CBM).

355 Cytowski et al., 2014, [38] *: Timothy allows scaling up simulations using dynamic and
 356 asymmetric domain decomposition, based on Peano-Hilbert space-filling curves, with the
 357 potential to reach the giga-scale.

358 Topol 2019, [49] *: Overview of the potential and limitations of different technologies, such as
 359 AI, Big Data and HPC, that have recently started to be used in biomedicine.

360 Björnsson et al., [51] *: The authors introduce the concept of the digital twin applied to
 361 personalised medicine and the tools and data that needs to be included to achieve it.

362 Nguyen et al., 2021, [63] *: Complete benchmark of different network inference methods
 363 developed for single-cell data. These single-cell networks could then be converted to
 364 individual cells' models to be used in multi-scale simulations.

365 Ozik et al., 2018, [94] **: Presentation of a model exploration framework compatible with
 366 HPC clusters that helps in setting-up, fitting and exploring multi-scale models.

367

368

References:

- 369 1. Jacob F, Monod J: **Genetic regulatory mechanisms in the synthesis of proteins.** *J
370 Mol Biol* 1961, **3**:318–356.
- 371 2. Ideker T, Galitski T, Hood L: **A new approach to decoding life: systems biology.**
372 *Annu Rev Genomics Hum Genet* 2001, **2**:343–372.
- 373 3. Kitano H: **Computational systems biology.** *Nature* 2002, **420**:206–10.
- 374 4. North MJ, Macal CM: *Managing Business Complexity: Discovering Strategic Solutions
375 with Agent-Based Modeling and Simulation.* Oxford University Press; 2007.
- 376 5. Macal CM, North MJ: **Tutorial on agent-based modelling and simulation.** *J Simul*
377 2010, **4**:151–162.
- 378 6. Anderson ARA, Weaver AM, Cummings PT, Quaranta V: **Tumor Morphology and
379 Phenotypic Evolution Driven by Selective Pressure from the Microenvironment.**
380 *Cell* 2006, **127**:905–915.
- 381 7. Wise SM, Lowengrub JS, Frieboes HB, Cristini V: **Three-dimensional multispecies
382 nonlinear tumor growth—I: Model and numerical method.** *J Theor Biol* 2008,
383 **253**:524–543.
- 384 8. Frieboes HB, Jin F, Chuang Y-L, Wise SM, Lowengrub JS, Cristini V: **Three-
385 dimensional multispecies nonlinear tumor growth—II: Tumor invasion and
386 angiogenesis.** *J Theor Biol* 2010, **264**:1254–1278.
- 387 9. Spatarello C-P, Zhang H, Nguyen DT, Han X, Liu R, Guo Q, Notbohm J, Fan J, Liu L,
388 Chen Z: **Biomechanics of Collective Cell Migration in Cancer Progression:
389 Experimental and Computational Methods.** *ACS Biomater Sci Eng* 2019, **5**:3766–
390 3787.
- 391 10. Gatenbee C, West J, Baker AM, Guljar N, Jones L, Graham TA, Robertson-Tessi M,
392 Anderson ARA: **Macrophage-mediated immunoediting drives ductal carcinoma
393 evolution: Space is the game changer.** *bioRxiv* 2019, doi:10.1101/594598.
- 394 11. Reticker-Flynn NE, Engleman EG: **Cancer systems immunology.** *eLife* 2020,
395 **9**:e53839.
- 396 12. Norton K-A, Gong C, Jamalian S, Popel AS: **Multiscale Agent-Based and Hybrid
397 Modeling of the Tumor Immune Microenvironment.** *Processes* 2019, **7**:37.
- 398 13. Ozik J, Collier N, Wozniak JM, Macal C, Cockrell C, Friedman SH, Ghaffarizadeh A,
399 Heiland R, An G, Macklin P: **High-throughput cancer hypothesis testing with an
400 integrated PhysiCell-EMEWS workflow.** *BMC Bioinformatics* 2018, **19**:483.
- 401 14. Letort G, Montagud A, Stoll G, Heiland R, Barillot E, Macklin P, Zinovyev A, Calzone
402 L: **PhysiBoSS: a multi-scale agent-based modelling framework integrating
403 physical dimension and cell signalling.** *Bioinformatics* 2019,
404 doi:10.1093/bioinformatics/bty766.
- 405 15. Getz M, Wang Y, An G, Becker A, Cockrell C, Collier N, Craig M, Davis CL, Faeder J,
406 Versypt ANF, et al.: **Rapid community-driven development of a SARS-CoV-2
407 tissue simulator.** *bioRxiv* 2020, doi:10.1101/2020.04.02.019075.
- 408 16. Metzcar J, Wang Y, Heiland R, Macklin P: **A Review of Cell-Based Computational
409 Modeling in Cancer Biology.** *JCO Clin Cancer Inform* 2019,
410 doi:10.1200/CCI.18.00069.
- 411 17. Osborne JM, Fletcher AG, Pitt-Francis JM, Maini PK, Gavaghan DJ: **Comparing
412 individual-based approaches to modelling the self-organization of multicellular
413 tissues.** *PLOS Comput Biol* 2017, **13**:e1005387.
- 414 18. Tisue S, Wilensky U: **NetLogo: A simple environment for modeling complexity.** In
415 *International conference on complex systems.* . Boston, MA; 2004:16–21.
- 416 19. Cooper F, Baker R, Bernabeu M, Bordas R, Bowler L, Bueno-Orovio A, Byrne H,
417 Carapella V, Cardone-Noott L, Cooper J, et al.: **Chaste: Cancer, Heart and Soft
418 Tissue Environment.** *J Open Source Softw* 2020, **5**:1848.
- 419 20. Gerlee P, Anderson ARA: **A hybrid cellular automaton model of clonal evolution
420 in cancer: The emergence of the glycolytic phenotype.** *J Theor Biol* 2008,
421 **250**:705–722.

- 422 21. Salguero AG, Capel MI, Tomeu AJ: **Parallel Cellular Automaton Tumor Growth**
 423 **Model.** In *Practical Applications of Computational Biology and Bioinformatics, 12th*
 424 *International Conference.* Edited by Fdez-Riverola F, Mohamad MS, Rocha M, De
 425 Paz JF, González P. Springer International Publishing; 2019:175–182.
- 426 22. Bauer E, Zimmermann J, Baldini F, Thiele I, Kaleta C: **BacArena: Individual-based**
 427 **metabolic modeling of heterogeneous microbes in complex communities.** *PLOS*
 428 *Comput Biol* 2017, **13**:e1005544.
- 429 23. Anderson A, Chaplain M, Rejniak K: *Single-cell-based models in biology and*
 430 *medicine.* Springer Science & Business Media; 2007.
- 431 24. Swat MH, Thomas GL, Belmonte JM, Shirinifard A, Hmeljak D, Glazier JA: **Multi-**
 432 **Scale Modeling of Tissues Using CompuCell3D.** In *Methods in Cell Biology.* .
 433 Elsevier; 2012:325–366.
- 434 25. Starruß J, Back W de, Brusch L, Deutsch A: **Morpheus: a user-friendly modeling**
 435 **environment for multiscale and multicellular systems biology.** *Bioinformatics*
 436 2014, **30**:1331–1332.
- 437 26. Sütterlin T, Kolb C, Dickhaus H, Jäger D, Grabe N: **Bridging the scales: semantic**
 438 **integration of quantitative SBML in graphical multi-cellular models and**
 439 **simulations with EPISIM and COPASI.** *Bioinformatics* 2013, **29**:223–229.
- 440 27. Norton K-A, Wallace T, Pandey NB, Popel AS: **An agent-based model of triple-**
 441 **negative breast cancer: the interplay between chemokine receptor CCR5**
 442 **expression, cancer stem cells, and hypoxia.** *BMC Syst Biol* 2017, **11**:68.
- 443 28. Harcombe WR, Riehl WJ, Dukovski I, Granger BR, Betts A, Lang AH, Bonilla G, Kar
 444 A, Leiby N, Mehta P, et al.: **Metabolic Resource Allocation in Individual Microbes**
 445 **Determines Ecosystem Interactions and Spatial Dynamics.** *Cell Rep* 2014,
 446 **7**:1104–1115.
- 447 29. Dukovski I, Bajić D, Chacón JM, Quintin M, Vila JC, Sulheim S, Pacheco AR,
 448 Bernstein DB, Rieh WJ, Korolev KS, et al.: **Computation Of Microbial Ecosystems**
 449 **in Time and Space (COMETS): An open source collaborative platform for**
 450 **modeling ecosystems metabolism.** *ArXiv200901734 Q-Bio* 2020,
- 451 30. Ruan X, Zhou J, Tu H, Jin Z, Shi X: **An improved cellular automaton with axis**
 452 **information for microscopic traffic simulation.** *Transp Res Part C Emerg Technol*
 453 2017, **78**:63–77.
- 454 31. Radzsuweit M, Block M, Hengstler JG, Schöll E, Drasdo D: **Comparing the growth**
 455 **kinetics of cell populations in two and three dimensions.** *Phys Rev E* 2009,
 456 **79**:051907.
- 457 32. Theis S, Suzanne M, Gay G: **Tyssue: an epithelium simulation library.** *J Open*
 458 *Source Softw* 2021, **6**:2973.
- 459 33. Fletcher AG, Osterfield M, Baker RE, Shvartsman SY: **Vertex Models of Epithelial**
 460 **Morphogenesis.** *Biophys J* 2014, **106**:2291–2304.
- 461 34. González- Valverde I, García- Aznar JM: **A hybrid computational model to explore**
 462 **the topological characteristics of epithelial tissues.** *Int J Numer Methods Biomed*
 463 *Eng* 2017, **33**:e2877.
- 464 35. Kang S, Kahan S, McDermott J, Flann N, Shmulevich I: **Biocellion: accelerating**
 465 **computer simulation of multicellular biological system models.** *Bioinforma Oxf*
 466 *Engl* 2014, **30**:3101–3108.
- 467 36. Hoehme S, Drasdo D: **A cell-based simulation software for multi-cellular**
 468 **systems.** *Bioinformatics* 2010, **26**:2641–2642.
- 469 37. Ghaffarizadeh A, Heiland R, Friedman SH, Mumenthaler SM, Macklin P: **PhysiCell:**
 470 **An open source physics-based cell simulator for 3-D multicellular systems.**
 471 *PLOS Comput Biol* 2018, **14**:e1005991.
- 472 38. Cytowski M, Szymanska Z: **Large-Scale Parallel Simulations of 3D Cell Colony**
 473 **Dynamics.** *Comput Sci Eng* 2014, **16**:86–95.
- 474 39. Walpole J, Papin JA, Peirce SM: **Multiscale computational models of complex**
 475 **biological systems.** *Annu Rev Biomed Eng* 2013, **15**:137–154.
- 476 40. Chylek LA, Harris LA, Faeder JR, Hlavacek WS: **Modeling for (physical) biologists:**

- 477 **an introduction to the rule-based approach.** *Phys Biol* 2015, **12**:045007.
- 478 41. Boutillier P, Maasha M, Li X, Medina-Abarca HF, Krivine J, Feret J, Cristescu I,
479 Forbes AG, Fontana W: **The Kappa platform for rule-based modeling.**
480 *Bioinformatics* 2018, **34**:i583–i592.
- 481 42. Harris LA, Hogg JS, Tapia J-J, Sekar JAP, Gupta S, Korsunsky I, Arora A, Barua D,
482 Sheehan RP, Faeder JR: **BioNetGen 2.2: advances in rule-based modeling.**
483 *Bioinformatics* 2016, **32**:3366–3368.
- 484 43. Andrews SS: **Smoldyn: particle-based simulation with rule-based modeling,
485 improved molecular interaction and a library interface.** *Bioinformatics* 2017,
486 **33**:710–717.
- 487 44. Blinov ML, Schaff JC, Vasilescu D, Moraru II, Bloom JE, Loew LM: **Compartmental
488 and Spatial Rule-Based Modeling with Virtual Cell.** *Biophys J* 2017, **113**:1365–
489 1372.
- 490 45. Santibáñez R, Garrido D, Martin AJM: **Atlas: automatic modeling of regulation of
491 bacterial gene expression and metabolism using rule-based languages.**
492 *Bioinformatics* 2020, **36**:5473–5480.
- 493 46. Maus C, Rybacki S, Uhrmacher AM: **Rule-based multi-level modeling of cell
494 biological systems.** *BMC Syst Biol* 2011, **5**:166.
- 495 47. Lowengrub JS, Frieboes HB, Jin F, Chuang Y-L, Li X, Macklin P, Wise SM, Cristini V:
496 **Nonlinear modelling of cancer: bridging the gap between cells and tumours.**
497 *Nonlinearity* 2010, **23**:R1–R9.
- 498 48. Sciumè G, Santagiuliana R, Ferrari M, Decuzzi P, Schrefler BA: **A tumor growth
499 model with deformable ECM.** *Phys Biol* 2014, **11**:065004.
- 500 49. Topol EJ: **High-performance medicine: the convergence of human and artificial
501 intelligence.** *Nat Med* 2019, **25**:44.
- 502 50. Zhao Y, Shuen TWH, Toh TB, Chan XY, Liu M, Tan SY, Fan Y, Yang H, Lyer SG,
503 Bonney GK, et al.: **Development of a new patient-derived xenograft humanised
504 mouse model to study human-specific tumour microenvironment and
505 immunotherapy.** *Gut* 2018, **67**:1845–1854.
- 506 51. Björnsson B, Borrebaeck C, Elander N, Gasslander T, Gawel DR, Gustafsson M,
507 Jörnsten R, Lee EJ, Li X, Lilja S, et al.: **Digital twins to personalize medicine.**
508 *Genome Med* 2019, **12**:4.
- 509 52. Jiao Y, Torquato S: **Emergent Behaviors from a Cellular Automaton Model for
510 Invasive Tumor Growth in Heterogeneous Microenvironments.** *PLOS Comput
511 Biol* 2011, **7**:e1002314.
- 512 53. Szymańska Z, Cytowski M, Mitchell E, Macnamara CK, Chaplain MAJ:
513 **Computational Modelling of Cancer Development and Growth: Modelling at
514 Multiple Scales and Multiscale Modelling.** *Bull Math Biol* 2018, **80**:1366–1403.
- 515 54. Soheilypour M, Mofrad MRK: **Agent-Based Modeling in Molecular Systems
516 Biology.** *BioEssays* 2018, **40**:1800020.
- 517 55. Cohen DPA, Martignetti L, Robine S, Barillot E, Zinovyev A, Calzone L: **Mathematical
518 Modelling of Molecular Pathways Enabling Tumour Cell Invasion and Migration.**
519 *PLoS Comput Biol* 2015, **11**:e1004571.
- 520 56. Calzone L, Tournier L, Fourquet S, Thieffry D, Zhivotovsky B, Barillot E, Zinovyev A:
521 **Mathematical modelling of cell-fate decision in response to death receptor
522 engagement.** *PLoS Comput Biol* 2010, **6**:e1000702.
- 523 57. Friedl P, Alexander S: **Cancer Invasion and the Microenvironment: Plasticity and
524 Reciprocity.** *Cell* 2011, **147**:992–1009.
- 525 58. Venkatesan S, Swanton C: **Tumor Evolutionary Principles: How Intratumor
526 Heterogeneity Influences Cancer Treatment and Outcome.** *Am Soc Clin Oncol
527 Educ Book* 2016, doi:10.1200/EDBK_158930.
- 528 59. Kim E, Kim J-Y, Smith MA, Haura EB, Anderson ARA: **Cell signaling heterogeneity
529 is modulated by both cell-intrinsic and -extrinsic mechanisms: An integrated
530 approach to understanding targeted therapy.** *PLoS Biol* 2018, **16**:e2002930.
- 531 60. Horvath P, Aulner N, Bickle M, Davies AM, Nery ED, Ebner D, Montoya MC, Östling

- 532 P, Pietiäinen V, Price LS, et al.: **Screening out irrelevant cell-based models of**
 533 **disease.** *Nat Rev Drug Discov* 2016, **15**:751–769.
- 534 61. Jabs J, Zickgraf FM, Park J, Wagner S, Jiang X, Jechow K, Kleinheinz K, Toprak UH,
 535 Schneider MA, Meister M, et al.: **Screening drug effects in patient-derived cancer**
 536 **cells links organoid responses to genome alterations.** *Mol Syst Biol* 2017, **13**:955.
- 537 62. Kulkarni A, Anderson AG, Merullo DP, Konopka G: **Beyond bulk: a review of single**
 538 **cell transcriptomics methodologies and applications.** *Curr Opin Biotechnol* 2019,
 539 **58**:129–136.
- 540 63. Nguyen H, Tran D, Tran B, Pehlivan B, Nguyen T: **A comprehensive survey of**
 541 **regulatory network inference methods using single cell RNA sequencing data.**
 542 *Brief Bioinform* 2021, **22**.
- 543 64. Saxena G, Ponce De Leon M, Montagud A, Vicente Dorca D, Valencia A: **BioFVM-X:**
 544 **An MPI+OpenMP 3-D Simulator for Biological Systems.** In *Computational Methods*
 545 *in Systems Biology*. . Springer International Publishing; 2021:5–18. In press.
- 546 65. Cytowski M, Szymanska Z: **Large-Scale Parallel Simulations of 3D Cell Colony**
 547 **Dynamics: The Cellular Environment.** *Comput Sci Eng* 2015, **17**:44–48.
- 548 66. Cytowski M, Szymańska Z, Umiński P, Andrejczuk G, Raszkowski K: **Implementation**
 549 **of an Agent-Based Parallel Tissue Modelling Framework for the Intel MIC**
 550 **Architecture.** *Sci Program* 2017, **2017**:e8721612.
- 551 67. Dunn S-J, Nähkne IS, Osborne JM: **Computational Models Reveal a Passive**
 552 **Mechanism for Cell Migration in the Crypt.** *PLOS ONE* 2013, **8**:e80516.
- 553 68. Dunn S-J, Fletcher AG, Chapman SJ, Gavaghan DJ, Osborne JM: **Modelling the role**
 554 **of the basement membrane beneath a growing epithelial monolayer.** *J Theor Biol*
 555 **2012**, **298**:82–91.
- 556 69. Coakley S, Gheorghe M, Holcombe M, Chin S, Worth D, Greenough C: **Exploitation**
 557 **of High Performance Computing in the FLAME Agent-Based Simulation**
 558 **Framework.** In *2012 IEEE 14th International Conference on High Performance*
 559 *Computing and Communication 2012 IEEE 9th International Conference on*
 560 *Embedded Software and Systems*. . 2012:538–545.
- 561 70. Coakley S, Smallwood R, Holcombe M: **Using x-machines as a formal basis for**
 562 **describing agents in agent-based modelling.** *Simul Ser* 2006, **38**:33.
- 563 71. Richmond P, Walker D, Coakley S, Romano D: **High performance cellular level**
 564 **agent-based simulation with FLAME for the GPU.** *Brief Bioinform* 2010, **11**:334–
 565 347.
- 566 72. Kaul H, Cui Z, Ventikos Y: **A Multi-Paradigm Modeling Framework to Simulate**
 567 **Dynamic Reciprocity in a Bioreactor.** *PLOS ONE* 2013, **8**:e59671.
- 568 73. Kabiri Chimeh M, Heywood P, Pennisi M, Pappalardo F, Richmond P: **Parallelisation**
 569 **strategies for agent based simulation of immune systems.** *BMC Bioinformatics*
 570 **2019**, **20**:579.
- 571 74. Li X, Upadhyay AK, Bullock AJ, Dicolandrea T, Xu J, Binder RL, Robinson MK, Finlay
 572 DR, Mills KJ, Bascom CC, et al.: **Skin Stem Cell Hypotheses and Long Term Clone**
 573 **Survival – Explored Using Agent-based Modelling.** *Sci Rep* 2013, **3**:1904.
- 574 75. Rudge TJ, Steiner PJ, Phillips A, Haseloff J: **Computational Modeling of Synthetic**
 575 **Microbial Biofilms.** *ACS Synth Biol* 2012, **1**:345–352.
- 576 76. Breitwieser L, Hesam A, Montigny J de, Vavourakis V, Iosif A, Jennings J, Kaiser M,
 577 Manca M, Meglio AD, Al-Ars Z, et al.: **BioDynaMo: a general platform for scalable**
 578 **agent-based simulation.** *bioRxiv* 2021, doi:10.1101/2020.06.08.139949.
- 579 77. Berghoff M, Rosenbauer J, Hoffmann F, Schug A: **Cells in Silico – introducing a**
 580 **high-performance framework for large-scale tissue modeling.** *BMC Bioinformatics*
 581 **2020**, **21**:436.
- 582 78. Collier N, North M: **Repast HPC: A Platform for Large-Scale Agent-Based**
 583 **Modeling.** In *Large-Scale Computing*. . John Wiley & Sons, Ltd; 2011:81–109.
- 584 79. Rousset A, Herrmann B, Lang C, Philippe L: **A Survey on Parallel and Distributed**
 585 **Multi-Agent Systems.** In *Euro-Par 2014: Parallel Processing Workshops*. Edited by
 586 Lopes L, Žilinskas J, Costan A, Casella RG, Kecskemeti G, Jeannot E, Cannataro M,

- 587 Ricci L, Benkner S, Petit S, et al. Springer International Publishing; 2014:371–382.
- 588 80. Cordasco G, De Chiara R, Mancuso A, Mazzeo D, Scarano V, Spagnuolo C: **Bringing together efficiency and effectiveness in distributed simulations: The experience with D-Mason.** *SIMULATION* 2013, **89**:1236–1253.
- 589 81. Byrne H, Drasdo D: **Individual-based and continuum models of growing cell populations: a comparison.** *J Math Biol* 2008, **58**:657.
- 590 82. Aguilar B, Gibbs DL, Reiss DJ, McConnell M, Danziger SA, Dervan A, Trotter M, Bassett D, Hershberg R, Ratushny AV, et al.: **A generalizable data-driven multicellular model of pancreatic ductal adenocarcinoma.** *GigaScience* 2020, **9**.
- 591 83. Szigeti B, Roth YD, Sekar JAP, Goldberg AP, Pochiraju SC, Karr JR: **A blueprint for human whole-cell modeling.** *Curr Opin Syst Biol* 2018, **7**:8–15.
- 592 84. Ozik J, Collier NT, Wozniak JM, Spagnuolo C: **From desktop to large-scale model exploration with SWIFT/T.** *Proc Winter Simul Conf Winter Simul Conf* 2016, **2016**:206–220.
- 593 85. Jagiella N, Rickert D, Theis FJ, Hasenauer J: **Parallelization and High-Performance Computing Enables Automated Statistical Inference of Multi-scale Models.** *Cell Syst* 2017, **4**:194–206.e9.
- 594 86. Rodriguez-Fernandez M, Egea JA, Banga JR: **Novel metaheuristic for parameter estimation in nonlinear dynamic biological systems.** *BMC Bioinformatics* 2006, **7**:483.
- 595 87. Audet C: **A Survey on Direct Search Methods for Blackbox Optimization and Their Applications.** In *Mathematics Without Boundaries*. Edited by Pardalos PM, Rassias TM. Springer New York; 2014:31–56.
- 596 88. Jagiella N, Müller B, Müller M, Vignon-Clementel IE, Drasdo D: **Inferring Growth Control Mechanisms in Growing Multi-cellular Spheroids of NSCLC Cells from Spatial-Temporal Image Data.** *PLOS Comput Biol* 2016, **12**:e1004412.
- 597 89. Tekin E, Sabuncuoglu I: **Simulation optimization: A comprehensive review on theory and applications.** *IIE Trans* 2004, **36**:1067–1081.
- 598 90. Ozik J, Collier N, Heiland R, An G, Macklin P: **Learning-accelerated discovery of immune-tumour interactions.** *Mol Syst Des Eng* 2019, **4**:747–760.
- 599 91. Akasiadis C, Ponce-de-Leon M, Montagud A, Michelioudakis E, Atsidakou A, Alevizos E, Artikis A, Valencia A, Paliouras G: **Parallel Model Exploration for Tumor Treatment Simulations.** *ArXiv210314132 Cs Q-Bio* 2021,
- 600 92. Raicu I, Foster IT, Yong Zhao: **Many-task computing for grids and supercomputers.** In *2008 Workshop on Many-Task Computing on Grids and Supercomputers*. . 2008:1–11.
- 601 93. Carillo M, Cordasco G, Serrapica F, Scarano V, Spagnuolo C, Szufel P: **Distributed simulation optimization and parameter exploration framework for the cloud.** *Simul Model Pract Theory* 2018, **83**:108–123.
- 602 94. Ozik J, Collier NT, Wozniak JM, Macal CM, An G: **Extreme-Scale Dynamic Exploration of a Distributed Agent-Based Model With the EMEWS Framework.** *IEEE Trans Comput Soc Syst* 2018, **5**:884–895.
- 603 95. Reuillon R, Leclaire M, Rey-Coyrehourcq S: **OpenMOLE, a workflow engine specifically tailored for the distributed exploration of simulation models.** *Future Gener Comput Syst* 2013, **29**:1981–1990.
- 604 96. Scott EO, Luke S: **ECJ at 20: toward a general metaheuristics toolkit.** In *Proceedings of the Genetic and Evolutionary Computation Conference Companion*. . ACM; 2019:1391–1398.
- 605 97. Babtie AC, Stumpf MPH: **How to deal with parameters for whole-cell modelling.** *J R Soc Interface* 2017, **14**:20170237.
- 606 98. Perdikaris P, Karniadakis GE: **Model inversion via multi-fidelity Bayesian optimization: a new paradigm for parameter estimation in haemodynamics, and beyond.** *J R Soc Interface* 2016, **13**:20151107.
- 607 99. Sahli Costabal F, Perdikaris P, Kuhl E, Hurtado DE: **Multi-fidelity classification using Gaussian processes: Accelerating the prediction of large-scale**

- 642 **computational models.** *Comput Methods Appl Mech Eng* 2019, **357**:112602.
- 643 100. Alber M, Tepole AB, Cannon WR, De S, Dura-Bernal S, Garikipati K, Karniadakis G,
644 Lytton WW, Perdikaris P, Petzold L, et al.: **Integrating machine learning and**
645 **multiscale modeling—perspectives, challenges, and opportunities in the**
646 **biological, biomedical, and behavioral sciences.** *Npj Digit Med* 2019, **2**:1–11.
- 647 101. Kiemen A, Braxton AM, Grahn MP, Han KS, Babu JM, Reichel R, Amoa F, Hong S-M,
648 Cornish TC, Thompson ED, et al.: **In situ characterization of the 3D microanatomy**
649 **of the pancreas and pancreatic cancer at single cell resolution.** *bioRxiv* 2020,
650 doi:10.1101/2020.12.08.416909.
- 651 102. Lomakin A, Svedlund J, Strell C, Gataric M, Shmatko A, Park JS, Ju YS, Dentro S,
652 Kleshchevnikov V, Vaskivskyi V, et al.: **Spatial genomics maps the structure,**
653 **character and evolution of cancer clones.** *bioRxiv* 2021,
654 doi:10.1101/2021.04.16.439912.
- 655 103. Béal J, Montagud A, Traynard P, Barillot E, Calzone L: **Personalization of logical**
656 **models with multi-omics data allows clinical stratification of patients.** *Front*
657 *Physiol* 2019, **9**:1965.
- 658 104. Liu A, Trairatphisan P, Gjerga E, Didangelos A, Barratt J, Saez-Rodriguez J: **From**
659 **expression footprints to causal pathways: contextualizing large signaling**
660 **networks with CARNIVAL.** *Npj Syst Biol Appl* 2019, **5**:1–10.
- 661 105. Ridgway D, Broderick G, Lopez-Campistrous A, Ru'aini M, Winter P, Hamilton M,
662 Boulanger P, Kovalenko A, Ellison MJ: **Coarse-grained molecular simulation of**
663 **diffusion and reaction kinetics in a crowded virtual cytoplasm.** *Biophys J* 2008,
664 **94**:3748–3759.
- 665 106. Azimi M, Bulat E, Weis K, Mofrad MRK: **An agent-based model for mRNA export**
666 **through the nuclear pore complex.** *Mol Biol Cell* 2014, **25**:3643–3653.
- 667 107. Westman AM, Peirce SM, Christ GJ, Blemker SS: **Agent-based model provides**
668 **insight into the mechanisms behind failed regeneration following volumetric**
669 **muscle loss injury.** *PLOS Comput Biol* 2021, **17**:e1008937.
- 670 108. de Montigny J, Iosif A, Breitwieser L, Manca M, Bauer R, Vavourakis V: **An in silico**
671 **hybrid continuum-/agent-based procedure to modelling cancer development:**
672 **Interrogating the interplay amongst glioma invasion, vascularity and necrosis.**
673 *Methods* 2021, **185**:94–104.
- 674 109. Goldberg AP, Szigeti B, Chew YH, Sekar JA, Roth YD, Karr JR: **Emerging whole-cell**
675 **modeling principles and methods.** *Curr Opin Biotechnol* 2018, **51**:97–102.
- 676 110. Stumpf MPH: **Statistical and computational challenges for whole cell modelling.**
677 *Curr Opin Syst Biol* 2021, doi:10.1016/j.coisb.2021.04.005.
- 678 111. Karr JR, Takahashi K, Funahashi A: **The principles of whole-cell modeling.** *Curr*
679 *Opin Microbiol* 2015, **27**:18–24.
- 680 112. Keating SM, Waltemath D, König M, Zhang F, Dräger A, Chaouiya C, Bergmann FT,
681 Finney A, Gillespie CS, Helikar T, et al.: **SBML Level 3: an extensible format for the**
682 **exchange and reuse of biological models.** *Mol Syst Biol* 2020, **16**:e9110.
- 683 113. Friedman SH, Anderson ARA, Bortz DM, Fletcher AG, Frieboes HB, Ghaffarizadeh A,
684 Grimes DR, Hawkins-Daarud A, Hoehme S, Juarez EF, et al.: **MultiCellIDS: a**
685 **community-developed standard for curating microenvironment-dependent**
686 **multicellular data.** *bioRxiv* 2016, doi:10.1101/090456.
- 687 114. Cosenza B, Popov N, Juurlink B, Richmond P, Chimeh MK, Spagnuolo C, Cordasco
688 G, Scarano V: **OpenABL: A Domain-Specific Language for Parallel and**
689 **Distributed Agent-Based Simulations.** In *Euro-Par 2018: Parallel Processing*.
690 Edited by Aldinucci M, Padovani L, Torquati M. Springer International Publishing;
691 2018:505–518.
- 692 115. Cosenza B, Popov N, Juurlink B, Richmond P, Chimeh MK, Spagnuolo C, Cordasco
693 G, Scarano V: **Easy and efficient agent-based simulations with the OpenABL**
694 **language and compiler.** *Future Gener Comput Syst* 2021, **116**:61–75.
- 695

696 **Figure legends:**

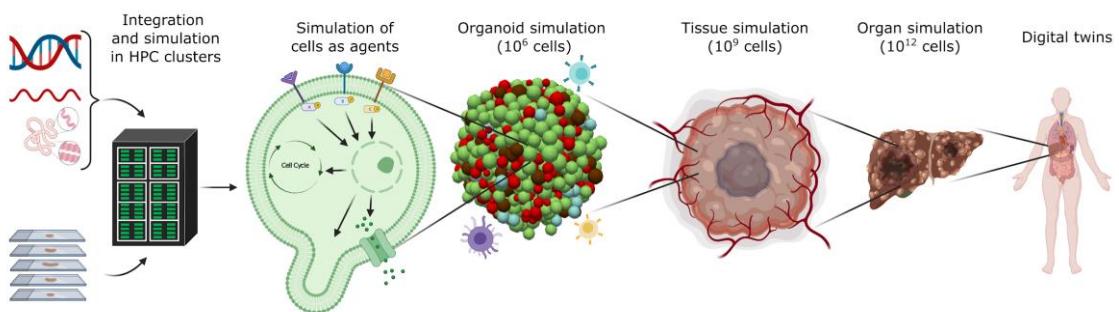
697 **Figure 1: Overview of the motivation to build large-scale CBM in biomedicine.** We
 698 make use of HPC clusters to integrate different types of omics and clinical data in agent-
 699 based models that are simulated with a focus on the cells' behaviours. These agents are
 700 studied as cells' populations where their emerging properties can be mechanistically
 701 inspected. The scale of these simulations currently ranges on the 10^6 cells but needs to
 702 reach 10^{12} cells and have complex environments if they are to simulate realistic, real-size
 703 tumours. These giga-scale simulations need to be enabled by HPC-optimised tools. Created
 704 with BioRender.com.

705 **Figure 2: Multi-scale agent-based models in systems biology.** A) Different agent-based
 706 approaches can be used for the modelling and simulation of cell populations, including, from
 707 top left to bottom right, lattice-based cellular automata, cellular Potts model, overlapping
 708 spheres and vertex models (see [17] for a review). B) Different models of intracellular
 709 models (e.g., metabolism, cell signalling) can be embedded into individual cell agents
 710 allowing to describe the problem at several different spatial scales and to consider
 711 heterogeneous populations. C) Schema of the multi-scale nature of the simulation with time
 712 intervals at which different processes are updated during a simulation. These are the
 713 diffusion of molecules and biochemical reactions (queried every 0.01 minute in [37]), the
 714 mechanical interactions (queried every 0.1 minute) and different cellular processes like
 715 commitment to cell division or apoptosis (queried every 6 minutes).

716 **Figure 3: Technologies and properties that will enable having large-scale agent-based
 717 simulations.** The field needs advances in four different areas to have simulations at the
 718 giga- scale: personalised intracellular mechanisms need to be included in the CBM (purple),
 719 these simulations need to allow for the description of a complex 3D environment (grey), the
 720 agents and the environment need to have dynamical biophysical properties (green), and all
 721 these tools need to be optimised for HPC clusters (blue). Created with BioRender.com.

722

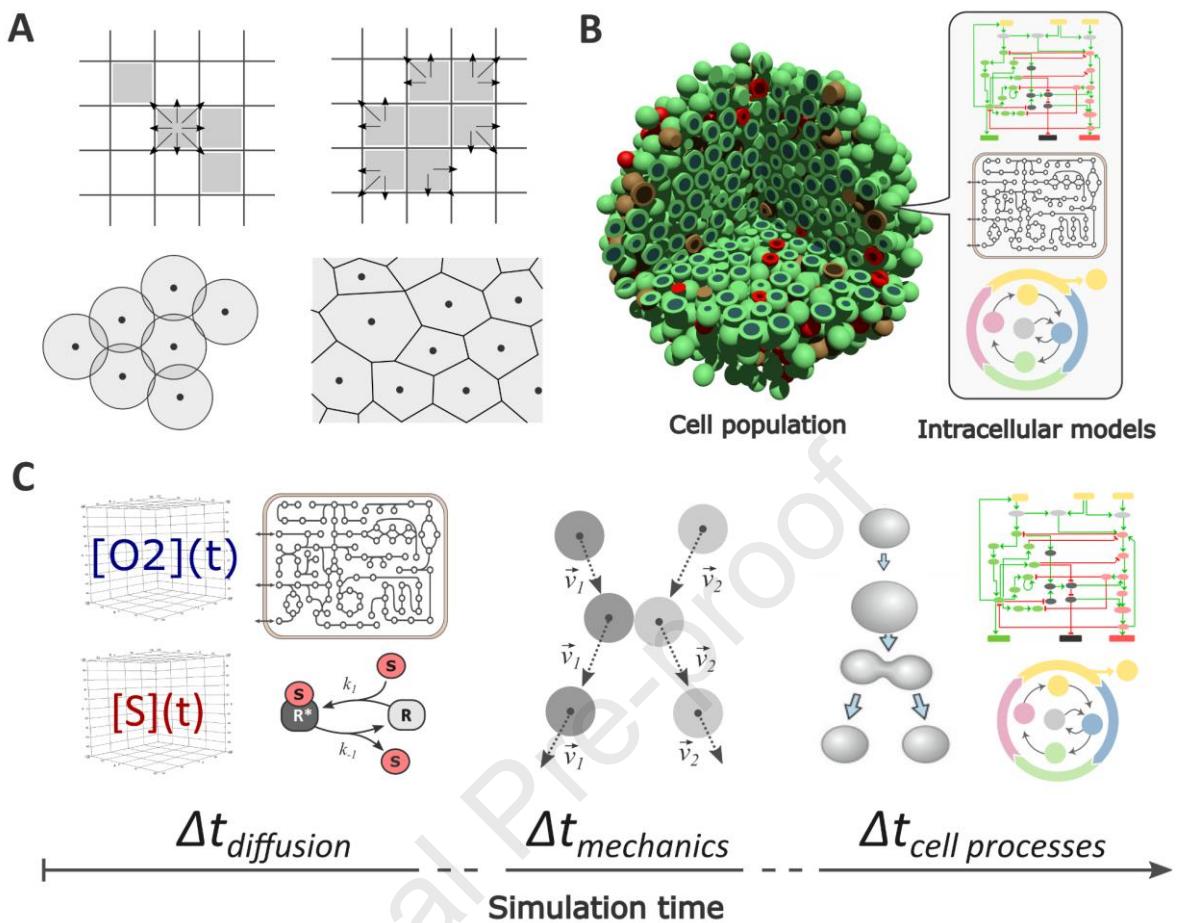
723

Figure 1:

724

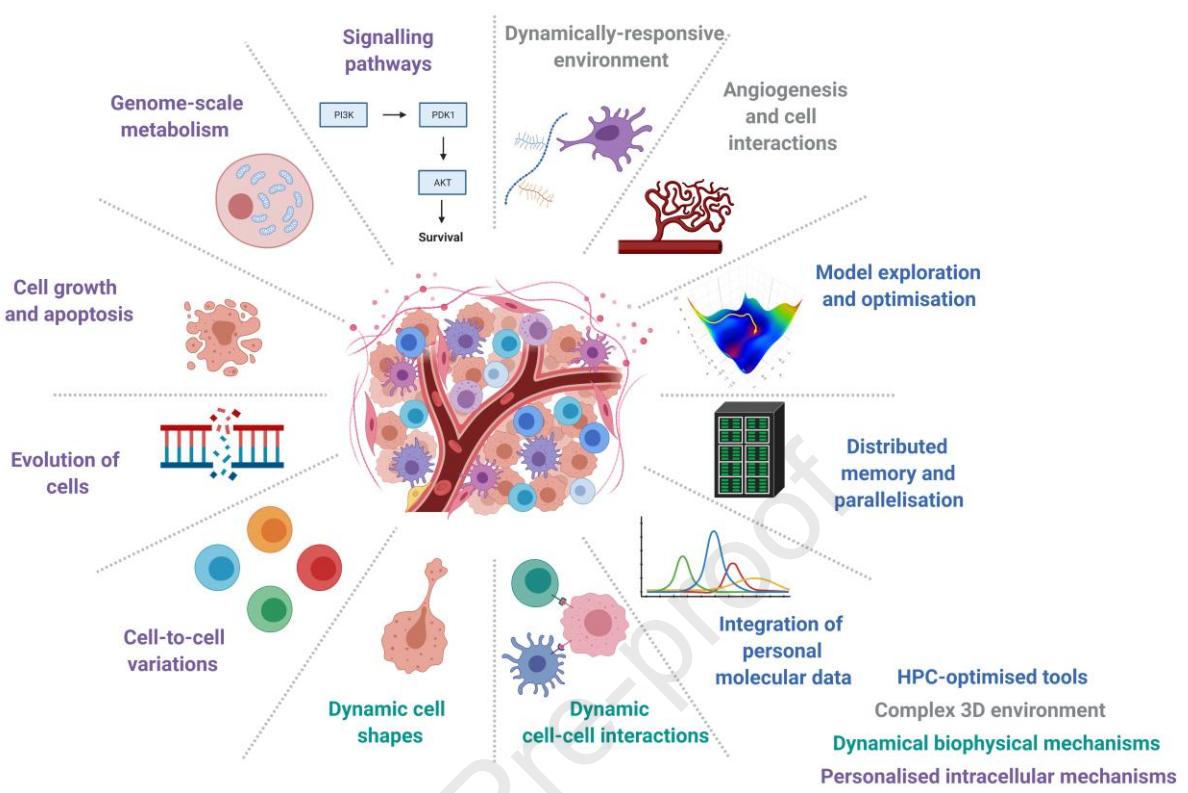
725

Figure 2:

726
727

728

Figure 3:



729

References' comments:

Papers of particular interest, published within the period of review, have been highlighted as:

* of special interest

** of outstanding interest

Macal & North, 2010, [5] *: Fundamental primer on agent-based modelling that presents the main concepts and some applications across disciplines.

Spatarelu et al., 2019, [9] *: Review on experimental and computational methods collective cell migration in cancer and how agent-based methods have helped gain insights into mechanisms.

Reticker-Flynn et al., 2020, [11] *: Review on the field of cancer systems immunology where the authors discuss the advances done by agent-based modelling and address the future avenues of study.

Norton et al., 2019, [12] *: Review on agent-based modelling of the immune microenvironment surrounding tumours and how different modelling approaches are best suited for different spatial scales.

Metzcar et al., 2019, [16] **: Comprehensive review on agent-based modelling, their different types and examples of its use in cancer biology.

Cooper et al., 2020, [19] *: Chaste is a simulation platform that allows users to test their models in three different agent-based modelling approaches (e.g., cellular automata, vertex-based, CBM).

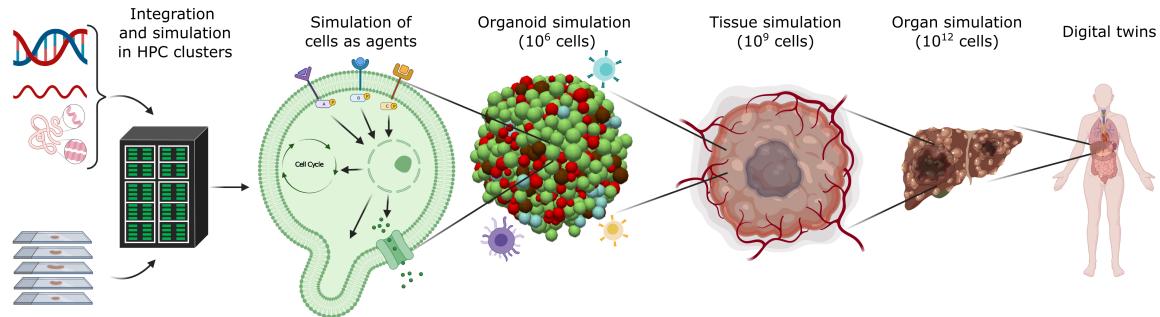
Cytowski et al., 2014, [38] *: Timothy allows scaling up simulations using dynamic and asymmetric domain decomposition, based on Peano-Hilbert space-filling curves, with the potential to reach the giga-scale.

Topol 2019, [49] *: Overview of the potential and limitations of different technologies, such as AI, Big Data and HPC, that have recently started to be used in biomedicine.

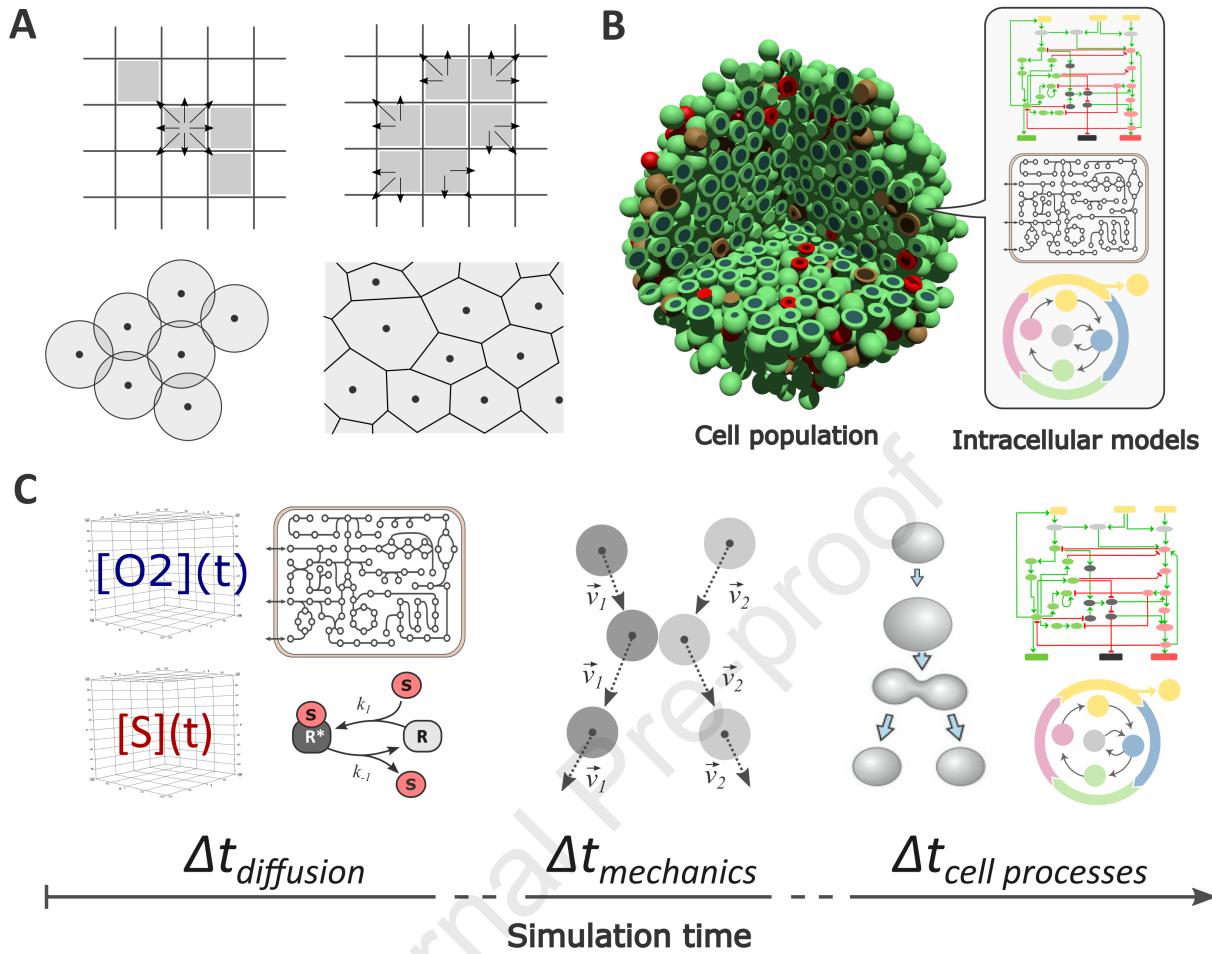
Björnsson et al., [51] *: The authors introduce the concept of the digital twin applied to personalised medicine and the tools and data that needs to be included to achieve it.

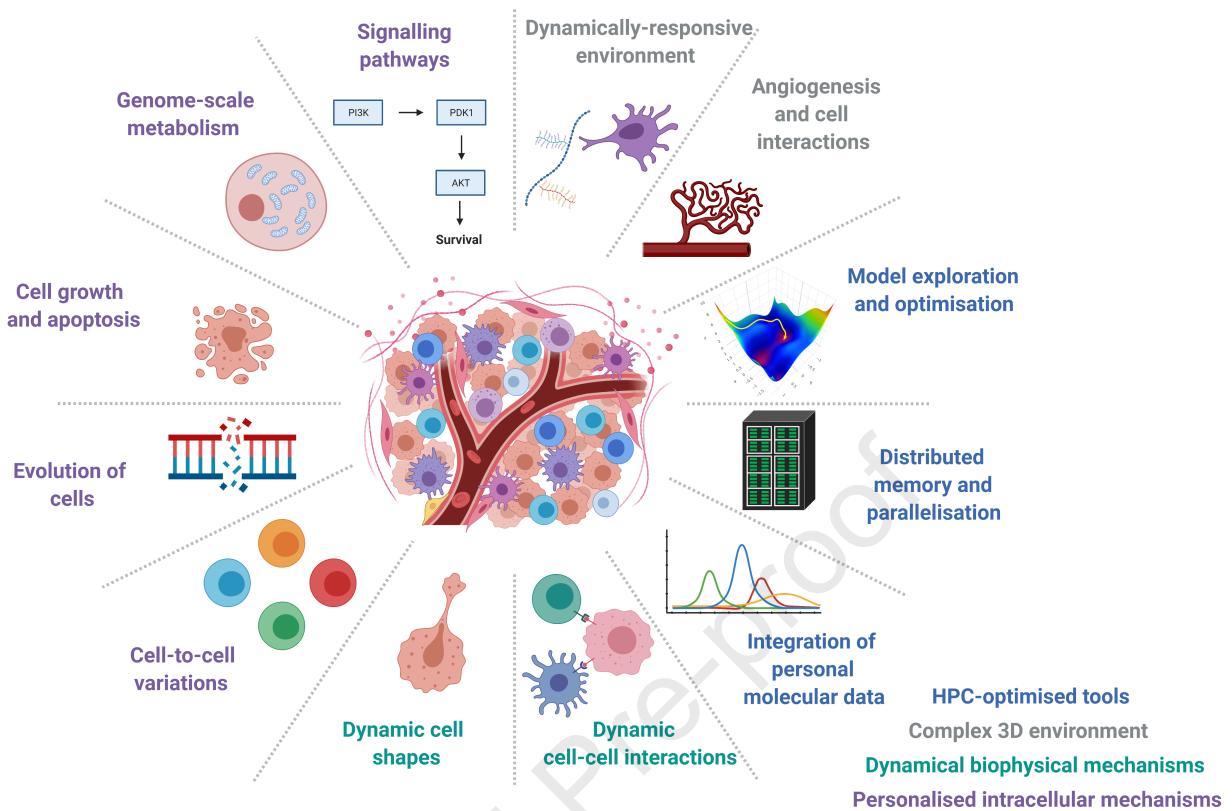
Nguyen et al., 2021, [63] *: Complete benchmark of different network inference methods developed for single-cell data. These single-cell networks could then be converted to individual cells' models to be used in multi-scale simulations.

Ozik et al., 2018, [94] **: Presentation of a model exploration framework compatible with HPC clusters that helps in setting-up, fitting and exploring multi-scale models.



Journal Pre-proof





Highlights:

- Agent-based modelling aims to explain, model and predict mechanisms in diseases.
- Large-scale agent-based modelling is needed to address real-sized, complex scenarios such as simulating tumours.
- The scaling-up of such models needs high-performance infrastructures and methods to optimise runtime and distribute memory among computing nodes.
- Such multi-scale models are complex and need tools such as model exploration frameworks to help set-up parameters and provide new hypotheses.
- Bringing together the systems biology and HPC communities is key for the development of the field.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

