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Multi-stage block freeze-concentration of green tea (*Camellia sinensis*) extract

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Abstract

Tea is the third most consumed beverage in the world. It has bioactive compounds that provide health benefits; for that reason, concentrating this product is a good alternative for different industries. Block freeze-concentration is a technology with the potential to be used to concentrate food solutions, preserving functional compounds. The aim of this work was to study the effect of block freeze-concentration on the total solids content, catechin content, polyphenol content, antioxidant activity and sensorial profile of green tea. Green tea extract at a 4 % (w/w) solid mass fraction was freeze-concentrated in three successive stages at -25°C. The effects of some parameters were evaluated, such as the concentration index, solid recovery yield, and concentration efficiency. After three stages, the solution was concentrated from 4% (w/w) to 14.1 % (w/w) with a distribution coefficient of 0.43 and a total concentration efficiency of 81.6%. The efficiency of concentration decreased with the number of stages. The levels of catechins and polyphenols were increased by 4.5- and 3.4-fold in the final liquid fraction, respectively. The antioxidant activity was preserved after three stages of freeze concentration. The physicochemical parameters were maintained: of the 27 sensory attributes evaluated, 18 of them were retained. The results demonstrated that block freeze-concentration is a useful technique for preserving and enhancing the bioactive compound content and the functional properties of green tea extracts.

1. Introduction

39 Some liquid products need to be concentrated to reduce transportation costs, reduce the
40 amount of water contained and increase its useful life as the number of microorganisms
41 that can grow is lower (Bozkir and Baysal, 2017). Some liquid products have bioactive
42 compounds with great benefits, but their concentrations are deficient within the matrix, as
43 is the case with green tea (Reygaert, 2017).

44 Tea is one of the most consumed beverages worldwide (Unno and Osakabe, 2018; Viljoen
45 et al., 2016). There are different types of tea depending on the production method, and
46 green tea is the only tea that is not fermented (Ho et al., 2015). Recently, interest in the
47 consumption of natural beverages has increased (Conger and Singg, 2019) due to their
48 different benefits. In the case of green tea the benefits include the prevention of heart
49 disease, insulin resistance, autoimmune disorders, and cancer prevention (Daglia et al.,
50 2017). There are many studies about its antioxidant activity, where the capacity to capture
51 free radicals is attributed to green tea (Ortiz-López et al., 2016). These benefits are
52 attributed to compounds known as catechins, which represent 70% of the bioactive
53 compounds in green tea extract. The catechin that is present in 80% of the total catechins,
54 the highest quantity, is 9-epigallocatechin-3-gallate (EGCG) (Yu et al., 2014). However,
55 catechin stability is affected when subjected to high temperatures, generating
56 epimerization of the molecules (Bhushani et al., 2017); thus, if necessary, it is more
57 advantageous to use non-thermal technology to concentrate green tea than traditional
58 evaporation.

59 Non-thermal technologies are widely used in the food industry for their ability to preserve
60 product quality by preventing the thermal degradation of temperature-sensitive compounds
61 (Chemat et al., 2017). These technologies can help reduce microbial load, avoid food
62 degradation, or increase food concentration (Pereira and Vicente, 2010). Freeze-
63 concentration or cryoconcentration is a non-thermal technology that allows the
64 concentration of different solutions (Aider and de Halleux, 2009); such technology
65 increases the concentration by separating the water from the solution, obtaining a very
66 diluted phase and a concentrated phase (Correa et al., 2017). There are three different
67 techniques of freeze concentration: the best-known at an industrial level is suspension
68 (Miyawaki et al., 2012). This technology is efficient in the separation process (Ding et al.,
69 2019), but at the operational level it has removable parts that require higher cost (Sanchez
70 et al., 2009); the other technologies are progressive freeze-concentration and block
71 freeze-concentration.

72 Block freeze-concentration (BFC) allows the solution to be completely frozen and then
73 partially thawed (Moreno et al., 2014a). The technique is also known as freeze-thaw
74 concentration. The first recovered fractions at the thawing step have the highest level of
75 solids. (Moreno et al. 2014b; Petzold et al., 2016). The process can be performed in
76 successive stages to increase the concentration index (Aider and Ounis, 2012). This
77 technology could have economic advantages. This technology may be more efficient than
78 other techniques if the amount of solids trapped in the diluted phase decreases (Aider
79 and de Halleux, 2009). Some challenges remain in the industrial application of block
80 freeze-concentration related to increasing the separation efficiency and establishing the
81 effect of the technique on the bioactive compounds, bioactivity, and sensory attributes
82 among other quality characteristics of the products. Moreover, a sensorial analysis
83 identifies the parameters of smell and flavour, employing different notes, (Lu et al., 2009)
84 to help identify the changes in green tea extract before and after freeze-concentration.

85 BFC has been studied on several fluids (Moreno et al., 2014a, 2013; Orellana-Palma et
86 al., 2017; Petzold et al., 2016, 2015). However, to our best knowledge, no references have
87 yet been found on the block freeze-concentration of green tea. Thus, this paper aimed to
88 evaluate the effect of three stages of block freeze-concentration on the solids content,
89 polyphenol content, catechin content, antioxidant activity and sensory quality of aqueous
90 green tea extract.

91 **2. Material and Methods**

92

93 **2.1. Materials**

94 Green tea extract was obtained from commercial dried tea leaves supplied by Agrícola
95 Himalaya (Bitaco) from Cali Colombia. First, 130 g of dried leaves were placed in 1 L of
96 distilled water at 80 °C for 30 min to obtain a solution with an initial solid content of 4%
97 (w/w) ; then, extraction was performed according to the protocol proposed by Vuong et al.,
98 (2011). The solution was stored at 4°C. The °Bx of all samples was measured by a
99 refractometer (Atago Pal 100, Tokyo, Japan) and a correlation between °Bx and solid
100 content was obtained. The solids content (Cs) was determined in an oven at $103 \pm 2^\circ\text{C}$
101 (NTC 4396,1998). The equation obtained was $Cs = 0.789 \cdot \text{°Bx}$ ($R^2 = 0.99$). Cs represent the
102 total amount of solids present in the tea extract. All measurements were performed in
103 triplicate.

104

105 2.2. Freeze-concentration tests

106 The block freeze-concentration tests were performed on the equipment located in the
107 laboratories of Universidad de La Sabana, Chia, Colombia, developed by Moreno et al
108 2014a (Figure 1). First, 300 mL of tea extract at 4 % (w/w) was placed in the vessel (1) to
109 be complete frozen and consequent thawed by using as the cooling and heating fluid,
110 ethylene glycol provided by the thermostatic baths (2) (Cole Parmer, Vernon Hills, USA)
111 (4,5) with a Cooling and heating power at 20 C of 800 W and 1 kW respectively. The bath
112 temperature was set to -25 °C, and the freezing time was 8 h. After the freezing stage, the
113 thawing stage was conducted by heating the external jacket of the container with the
114 heating fluid settled at 20 °C (3). Ten thawed fractions were collected in separated
115 containers (4) in the thawing stage by gravitational separation and weighed with a scale
116 (5) (Mettler Toledo, Greifensee, Switzerland). The thawing time varied between 4 and 6 h.
117 Finally, the concentration was measured by refractometry (Atago model PAL-100; Tokyo,
118 Japan) and converted to a content of solids with the correlated equation $C_s = 0.789 * \text{°Bx}$.
119 This equation was generated by this work as explained before.

120 The experimental design was performed in three stages as shown in Figure 2. Two
121 streams with the mixing of some collected fractions were obtained after freeze
122 concentration, one diluted and other concentrated. The definition of the number of
123 fractions to be mixed was performed according to the results shown in figure 2. The
124 fractions in which the concentration index (defined in equation 1) was greater than one
125 were mixed and identified as the concentrated stream (liquid fraction). The fractions with a
126 concentration index less than one were mixed and identified as the diluted stream (ice
127 phase). The diluted streams were discarded. The concentrated phase was used to the
128 next stage. The first stage was preformed four times and the second stage twice to
129 complete the volume necessary to the next stage. A total of three stages with seven tests
130 were performed as shown in Figure 2. The overall process was performed in triplicate.

131

132 2.3. Evaporation test

133 An evaporation test was performed to compare the catechin concentration results with a
134 method of concentrating at higher temperatures. The evaporation process was performed
135 using 100 g of green tea extract (4% w/w) obtained under the same conditions as the

136 extract used in the freeze-concentration tests. The sample was evaporated at 92°C, the
 137 boiling point at Bogotá, using a beaker on a heating plate (Thermo Fischer Scientific,
 138 Waltham, USA). The sample was evaporated at 92 °C for the necessary time to obtain the
 139 same concentration that results in the three stages of BFC. The solids content was
 140 measured by refractometry. The samples were stored at 4 °C until further analysis.

141 **2.4. Response variables**

142 The following response variables were calculated to evaluate the behaviour of the freeze
 143 concentration tests:

144 **Concentration index (CI):** This factor was defined as the ratio between the solids content
 145 in the liquid fraction and the solids content in the initial fraction (Moreno et al., 2014a).

$$146 \quad CI = \frac{C_{sLIQ}}{C_{s0}} \quad (1)$$

147 Where, CI is the concentration index, C_{sLIQ} is the total solid content (%w/w) in the
 148 individual liquid fraction and C_{s0} is solid content of the initial solution.

149 **Thawing fraction (f):** This was defined as the ratio between the mass thawed and the
 150 initial mass, and this variable can be used to follow the development of the process
 151 (Moreno et al., 2014a).

$$152 \quad f = \frac{m_{LIQ}}{m_0} \quad (2)$$

153 where m_{LIQ} and m_0 are the liquid and initial mass, respectively.

154
 155 **Solid Yield (Y):** This factor was defined to analyse the solids recovery; it was calculated
 156 as the ratio between the solid mass in the liquid and the solid mass present in the initial
 157 solution (Moreno et al., 2013):

$$158 \quad Y = \frac{m_{sLIQ}}{m_{s0}} \quad (3)$$

159 where m_{sLIQ} is the solid mass in the recollected liquid, and m_{s0} is the solid mass in the initial
 160 solution.

161

162 **Concentration efficiency (eff):** This was defined as the concentration of solids recovery
 163 in the liquid solution with respect to the concentration of solids present in the ice phase.
 164 The equation is as follows (Gulfo et al., 2014):

165

$$166 \quad \quad \quad eff = \frac{C_{sLIQ} - C_{sICE}}{C_{sLIQ}} * 100 \quad (4)$$

167 Where, C_{sLIQ} and C_{sICE} are the total solid content (%w/w) in the liquid and ice fractions
168 respectively.

169 **The average distribution coefficient:** This is the ratio between the solids in the ice phase
170 and the solids in the liquid phase (Moreno et al., 2014b).

171

$$172 \quad \quad \quad k = \frac{C_{sICE}}{C_{sLIQ}} \quad (5)$$

173

174 **Productivity:** This factor is defined as the rate of separation of water from the initial
175 solution (Moreno et al., 2015).

176

$$177 \quad \quad \quad Productivity = m_{ice} * \frac{(1 - C_{sICE})}{100 * (m_0 * t)} \quad (6)$$

178 Where, m_{ice} is the mass of the ice produced during freeze concentration, C_{sICE} is the total
179 solid concentration in the ice, m_0 is mass of the initial solution, and t is the total processing
180 time of the freeze concentration in hours. The productivity of the operation represent the
181 rate of water separation from the solution achieved by the freeze concentration process.

182 **2.5. Bioactive compounds and bioactivity of green tea extract**

183 The following measurements were performed for each of the samples before and after the
184 three stages of BFC and the evaporation concentration.

185 **Catechin content**

186 The catechins were measured by high-performance liquid chromatography (HPLC). The
187 column used was a Zorbax SB-C8 with a UV detector at 280 nm, the mobile phases were
188 methanol (25%) and trifluoroacetic acid (TFA) (57%), and the catechins were reported in
189 mg/mL of green tea extract (Gadkari et al., 2014).

190 **Total polyphenol content**

191 The polyphenol content was measured by Folin-Ciocalteu according to ISO
192 14502-1 (2005). A sample of 1 mL of tea extract was mixed with 1 mL of gallic acid at
193 different concentrations and 5 mL of Folin-Ciocalteu reagent; the reaction was left for 60
194 min at room temperature, and then 4 mL of 7.5% Na_2CO_3 was added. The absorbance
195 was measured at 765 nm, using a spectrophotometer (probe 600 Merk, Darmstadt,
196 Germany), and a blank sample was included. The results were expressed in mg equivalent
197 gallic acid per gram of tea leaf (dry basis).

198 **Antioxidant activity**

199 The antioxidant activity was measured by three different methods: DPPH (2,2-Diphenyl-1-
200 picrylhydrazyl), ABTS (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid)) and ORAC
201 (oxygen radical absorbance capacity). Different methods were used to show the
202 reproducibility of the results.

203 The DPPH technique was performed using 2 mL of green tea extract and mixed with
204 DDPH reactive and a 2 mL of solution ethanol; the absorbance was measured at 517 nm,
205 and the results were expressed as grams of TEAC Trolox per 100 grams of tea leaves or
206 mL of solution. (Yu et al., 2007). The ABTS technique was performed using 0.025 mL of
207 green tea extract mixed with a previously prepared Trolox ethanolic solution. The mixture
208 was left for approximately 2 h in a dark room; then, the samples were read in a
209 spectrometer at 734 nm (Damiani et al., 2014). The results were expressed as mmol/L
210 Trolox equivalents per 100 grams of tea leaves. Finally, the ORAC technique was
211 performed using 20 µl of sample and mixed with Trolox reactive. The pH was adjusted to
212 7.7, and the samples were incubated in boxes with 96 plates for 10 min at 37°C; the
213 measurements were performed at 538 nm. The results were expressed as µmol of Trolox
214 equivalents per mL of tea extract (µmol TE/100 mL) (Liu et al., 2018).

215 **2.6. Sensory analysis**

216 A panel of seven trained judges between 25 and 60 years at the Laboratory of Sensory
217 Analysis of Universidad de Antioquia (Medellin, Colombia) evaluated the sensory
218 properties of the initial and freeze-concentrated green tea extract. The samples were re-
219 dissolved and standardized at the same final solid content (1.5 % w/w) This content was
220 settled to compare the sensory quality of a possible tea beverage produced from the
221 freeze concentrated extract. Panellists evaluated the appearance, texture, taste, flavour,
222 and odour according to the Colombian National Standard NTC 3501 and NTC 3932.

223 **2.7. Statistical analysis**

224

225 Data were analysed using SAS9.2 (SAS Institute Inc., Cary, USA) software package. All
226 tests were performed in triplicate. Data were reported with the deviation, and the variance
227 (ANOVA) was evaluated to identify the differences among the data with a level of
228 significance of 95%. Pearson's correlation coefficients were used to compare the
229 relationship between the bioactive compounds and antioxidant activity.

230 3. Results and discussion

231 3.1. Freeze-concentration tests

232 The first analysis corresponds to the solid content of the individual thawing fractions. The
233 results of the concentration index (CI) for each stage of BFC at the ten fractions obtained
234 following the thawing process are shown in Figure 3. The concentration index represents
235 the number of times the initial concentration of the solution was increased for each
236 collected fraction during the thawing stage. It is important to note that each thawing
237 fraction was analysed separately, thus the CI in the figure 3 represents the individual
238 concentration index. The behaviour was similar for the three stages. The graph shows
239 that the solid content of the first fractions recovered in the thawing stage had the highest
240 concentration, with CI values of approximately 2.6. This is because the drops of liquid that
241 were being separated in the first thawed fractions were enriched with the solids eluted
242 from the frozen matrix during the thawing stage (Nakagawa et al., 2010; Robles et al.,
243 2016). Consequently, the solid content decreased at each thawing fraction until the
244 concentration index values approached 0.

245 The second analysis corresponds to the result of the three stages of freeze concentration.
246 The horizontal line of the figure 3 represents the case in which $CI=1$, which indicates that
247 the fractions that are being separated have the same solid content than the initial solution.
248 At this point, two streams can be separated. The first, with the highest solid content,
249 composed by the mixture of the first fractions is the concentrated phase, and the last, with
250 the lowest solid content, containing the last fractions is the diluted phase. The average f at
251 $CI=1$ of each test was determined and showed in the table 1. and a mass balance was
252 made at this f , to calculate the final solid content of each stream as shown in Figure 4. The
253 values are expressed as a percent w/w. The solid content in the liquid phase increased
254 from 4% (w/w) to 14.1% (w/w) in the three stages, whereas the solids content in the ice
255 phase was 5.86% (w/w); the overall concentration efficiency of the process was 81.57%.
256 The results are similar to those reported for the BFC of other products (Moreno et al.,
257 2014b, Petzold et al., 2015).

258 The response variables of the block freeze-concentration at the average f value when $CI=1$
259 are shown in Table 1. These values can be calculated with the equations 1 to 6. The total
260 solids content increased by 3.52-fold in the three stages. In each stage the concentration
261 index was approximately 1.5, but in the second stage this value was higher. The Y_{stage}
262 represents the solid yield, or the amount of solids that can be recovered from the initial

263 solution at the end of each BFC stage. In the first two stages it was similar; 78% of the
264 solids were recovered. In the third stage the value decreased slightly due to the difficulty of
265 recovering solids in more concentrated solutions. Thus, there was always a similar solids
266 recovery. The highest value of the average distribution coefficient was obtained in the third
267 stage. This is because the concentration was higher at the third stage, and the solids
268 increased the viscosity and reduced the mobility of solids during the thawing stage,
269 allowing more solids to be trapped in the ice (Dunaway et al., 2010; Moreno et al., 2014a;
270 Nakagawa et al., 2010; Osorio et al., 2018). Finally, the productivity of the operation was
271 0.04 kg water removed per kg extract per hour. This value is lower than that reported by
272 Moreno et al. 2015 for coffee extract due to the lower temperature needed to completely
273 freeze the green tea sample.

274 The results demonstrated that BFC is an effective technique for concentrating tea and
275 increasing the solids content. Similar values were reported by Moreno et al. (2014) in
276 coffee extract. (Moreno et al., 2014a); however, the processing time is relatively high,
277 approximately 8 h for each BFC. Similar results were obtained in coffee extract (Moreno et
278 al. 2015). The high processing time is maybe the greatest disadvantage of block freeze
279 concentration. Several alternatives in the thawing and separation stages such as
280 microwave thawing, vacuum, and centrifugal separation are being explored to increase the
281 concentration efficiency or reduce the processing time (Orellana-Palma et al., 2017;
282 Petzold et al., 2016).

283 Finally, a third analysis was done in this section. A multistage process of block freeze
284 concentration was proposed. Figure 5 shows a scheme of three BFC stages and the solids
285 content of the concentrated and diluted phases. The arrows pointing downwards represent
286 the ice phase (diluted), and the right arrows the concentrated phase which enter to the
287 new stage. BCF3 effluent (5.83% (w/w)) can be recirculated to the stage 2 to recover the
288 solids contained in that stage. In contrast, the ice obtained from stages 1 and 2, because
289 they are very diluted, can be defined as final effluents. A mass balance was calculated
290 with this process strategy and the overall concentration efficiency and concentration index
291 were calculated. In addition, an experimental equation to calculate the concentration index
292 as a function of the concentration of liquid at each stage was used. An overall process
293 efficiency of 85.3% can be achieved with this process configuration, with a concentration
294 index of 3.52, an f value of 0.16 and a solute yield of 0.56. In contrast, the process without
295 recirculation achieved a concentration efficiency of 81.1%, an f value of 0.11 and a solute

296 yield of 0.41. The results showed an improve of the process. Moreover, the multistage
297 process can be applied with a higher number of stages to reach a final concentration to
298 obtain a more concentrated extract. The limit is determined by the solid-liquid equilibrium
299 of the system. Further research on the topic is necessary. This kind of product can be
300 commercialized as a concentrated liquid to be reconstituted after storage, or even as a
301 product prior to freeze-drying to obtain freeze-dried green tea extract.

302 **3.2. Biocompounds and bioactivity of the extract**

303

304 Catechins, polyphenols, and antioxidant activity in the extract before and after block
305 freeze-concentration and the comparison between the values obtained with evaporation
306 are shown in Table 2. In general, the results indicate that both bioactive compounds and
307 bioactivity were significantly increased at each stage of BFC. The table shows the value of
308 the concentration index reached at the third stage (CI Stage 3). This value corresponds to
309 how many times the concentration of the compound or the activity was increased with
310 respect to the initial concentration or activity. The polyphenol content and catechin
311 concentration were increased by a similar factor as the increase in total solids content. The
312 catechins were slightly more concentrated than the total solids. This could be obtained by
313 a possible selective separation due to differences in polarity or molecular size. A further
314 experimentation is recommended to explain this result and to determine the useful of this
315 selectivity. This result showed that the BFC preserved the bioactive compounds of the
316 extract. In contrast, the evaporation technique used with the same solids content showed a
317 decrease in most of the bioactive compounds and bioactivity ($CI < 1$).

318 Similarly, the antioxidant activity measured by DPPH, ABTS and ORAC was increased
319 after BFC but with a lower concentration index than the bioactive compound preservation.
320 However, the CI obtained for the antioxidant activity of BFC was higher than when using
321 the evaporation technique. The higher CI obtained by ORAC, ABTS and DPPH presented
322 similar values of CI, between 1.48 and 2.02 for BFC; in contrast, evaporation obtained
323 values ranging from 0.59 to 1.58. The DPPH values were higher with respect to the other
324 two techniques (Kopjar et al., 2015); similar values were found in matrices such as yogurt
325 (Jaster et al., 2018) and yerba mate (Dudonné et al., 2009). The catechins increased by
326 4.45-fold, but the activities increased by 1.48 to 2.02; hence, the prolonged exposure of 12
327 h to cold could affect the ability to capture free radicals. On the other hand, the
328 polyphenols were increased; thus, the technique of freeze-concentration is highly efficient

329 in preserving these bioactive compounds; similar results were found for matrices such as
330 wine (Zhang et al., 2016). In contrast, evaporation decreased the concentration of
331 bioactive compounds and the bioactivity of the green tea extract.

332 The results of the correlation analysis between bioactive compounds and antioxidant
333 activity in BFC are shown in Table 3. Pearson's correlation coefficient is reported. This
334 value represents the linear relationship between the variables, and it can be used to
335 establish a correlation between the compounds and the bioactivity. A good correlation
336 between the bioactive compounds and the antioxidant activity was found. This result
337 indicates that the enhancement of the activity after BFC can be explained by the increase
338 in the catechin and polyphenol contents. In addition, a direct correlation between catechins
339 and polyphenols of 0.983 was obtained; catechins have a correlation with antioxidant
340 activities with values between 0.771-0.936, and polyphenols have a correlation with values
341 between 0.733-0.953. BFC preserves the compounds and the functional quality of green
342 tea extract. The evaporation test did not yield a good correlation.

343 **3.3. Sensory analysis**

344 A sensory evaluation of green tea extract can directly reflect the quality of the samples.
345 The sensory profile of extract tea in a concentrated liquid after three stages of BFC and
346 without treatment is shown in the spider web diagram in Figure 6. The panellists evaluated
347 27 parameters among flavour (F), odour (O), texture (T), appearance (A), and sensorial
348 sensation (Ss), and they did not perceive any significant difference in 18 of the 27
349 evaluated parameters. Some parameters of the freeze-concentrated sample obtained a
350 higher score than the initial extract at the same concentration and other parameters a
351 lower value. However, the overall quality was statistically preserved in the tea extract
352 before and after freeze concentration. This result shows that BFC is an effective technique
353 for preserving the sensory quality of tea extract.

354

355 Despite the overall quality preservation, some attributes such as tobacco, bitterness, green
356 colour, sweetness, herbal flavour and sweet odour were parameters that presented a
357 significant difference with $p < 0.05$. This means, the probability to reject the hypothesis of
358 the statistical difference is less than 5%. The overall quality did not show statistical
359 differences. The changes in the freeze-concentrated extract, compared with the extract
360 without treatment, can be related to the volatile compounds found in the polyphenols and

361 other bioactive compounds (Ahmad et al., 2015; Wu et al., 2017) that were present in high
362 quantity in the BFC extract, as the samples were diluted with water at the same
363 concentration as the extract without treatment; at the same time, exposure to temperatures
364 below zero can cause changes in odour (Zhang et al., 2016). Similar results were obtained
365 for other matrices such as coffee (Moreno et al., 2015) and strawberry pulp (Jaster et al.,
366 2018). These results show that the sensorial quality can be preserved using block freeze-
367 concentration.

368

369 **4. Conclusions**

370 It was possible to concentrate the green tea extract (*Camellia sinensis*) from 4 % (w/w)
371 solids to 14.1 % (w/w) after three stages of block freeze-concentration. The efficiency of
372 concentration was 81.3% after three stages and the solute yield 42%. These values
373 improve to 85.3% and 56% respectively when a recirculation strategy is applied. The
374 concentration of the bioactive compounds, catechins, and polyphenols can be increased
375 during freeze concentration. In contrast, these compounds were degraded during
376 concentration by evaporation. The antioxidant activity was improved with BFC. A
377 correlation between the compounds and bioactivity was found. The sensory profile of most
378 attributes was preserved during freeze concentration. The overall sensory quality of the
379 beverage was preserved by BFC. The multistage process can be applied to obtain higher
380 concentrations depending on the number of stages. The results show that block freeze-
381 concentration is an efficient technique for increasing the solids and bioactive compound
382 contents, increasing the antioxidant activity and preserving the sensory quality of green tea
383 extract.

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Table 1 Initial solids concentration, concentration index, solid yield and distribution coefficient at each stage of BFC for green tea extract, at CI=1

Stage	C _{S_{LIQ}} (% w/w)	C _{S_{ICE}} (% w/w)	Y	CI	k	f	Eff(%)	Productivity (kg water/(kg extract. h))
1	5.20 ± 0.00 ^c	1.25 ± 0.01 ^c	0.79 ± 0.062 ^a	1.53 ± 0.025 ^b	0.25 ± 0.075 ^b	0.65 ± 0.029 ^a	75.38 ± 4.21 ^a	0.043 ± 0.002 ^b
2	8.94 ± 0.01 ^b	2.08 ± 0.03 ^b	0.77 ± 0.075 ^a	1.73 ± 0.084 ^a	0.24 ± 0.064 ^b	0.45 ± 0.058 ^b	76.79 ± 3.52 ^a	0.044 ± 0.004 ^b
3	14.1 ± 0.11 ^a	5.76 ± 0.18 ^a	0.68 ± 0.007 ^b	1.55 ± 0.078 ^b	0.43 ± 0.013 ^a	0.40 ± 0.009 ^b	59.10 ± 2.87 ^b	0.046 ± 0.001 ^a

Average values in the same column with different superscript letters are significantly different ($p < 0.05$).

Table 2. Catechin content, total polyphenols content, and antioxidant activity at each stage of BFC of green tea extract compared with evaporation at 92°C.

Block freeze concentration					
Stage	Initial	Stage 1	Stage 2	Stage 3	CI stage 3
Solid concentration (% w/w)	4.0 ± 0.04 ^d	5.2 ± 0.06 ^c	8.9 ± 0.09 ^b	14.1 ± 0.11 ^a	3.53 ± 0.06
Catechins (mg/mL)	6.25 ± 0.58 ^d	11.30 ± 0.95 ^c	16.42 ± 1.38 ^b	27.80 ± 0.45 ^a	4.45 ± 0.50
Polyphenols (mg eq GAE/mL)	14.48 ± 0.69 ^d	18.71 ± 1.64 ^c	34.88 ± 3.75 ^b	50.33 ± 6.31 ^a	3.48 ± 0.60
DPPH (mg eq Trolox/mL)	95.90 ± 2.76 ^b	148.11 ± 4.67 ^b	157.78 ± 5.36 ^a	166.08 ± 4.69 ^a	1.73 ± 0.10
ABTS (mg eq Trolox/mL)	68.32 ± 1.98 ^d	75.61 ± 2.04 ^c	97.64 ± 3.22 ^b	101.41 ± 3.44 ^a	1.48 ± 0.09
ORAC (mg eq Trolox/mL)	50.74 ± 2.92 ^d	78.52 ± 6.01 ^c	85.54 ± 3.28 ^b	102.48 ± 3.29 ^a	2.02 ± 0.19
Evaporation					
Solid concentration (% w/w)	4.0 ± 0.02 ^d	5.2 ± 0.03 ^c	8.9 ± 0.12 ^b	14.1 ± 0.24 ^a	3.53 ± 0.06
Catechins (mg/mL)	4.65 ± 0.12 ^a	0.91 ± 0.12 ^b	0.81 ± 0.09 ^c	0.71 ± 0.07 ^d	0.15 ± 0.01
Polyphenols (mg eq GAE/mL)	12.01 ± 0.24 ^b	10.48 ± 1.40 ^b	16.64 ± 2.27 ^a	8.08 ± 1.23 ^c	0.67 ± 0.11
DPPH (mg eq Trolox/mL)	91.54 ± 6.87 ^a	79.25 ± 3.74 ^b	63.48 ± 1.27 ^c	54.35 ± 2.51 ^d	0.59 ± 0.06
ABTS (mg eq Trolox/mL)	63.40 ± 1.81 ^d	98.79 ± 2.16 ^c	101.45 ± 1.35 ^b	100.73 ± 3.51 ^a	1.58 ± 0.09
ORAC (mg eq Trolox/mL)	44.32 ± 2.75 ^d	58.22 ± 2.49 ^b	66.79 ± 1.55 ^a	54.31 ± 2.43 ^c	1.23 ± 0.12

Average values in the same row with different superscript letters are significantly different ($p < 0.05$).

Table 3. Correlations between bioactive compound concentration and antioxidant activity.

	Catechins	Polyphenols	DPPH	ABTS	ORAC
Catechins	1.000	0.983	0.771	0.905	0.936
Polyphenols	0.983	1.000	0.733	0.953	0.901
DPPH	0.771	0.733	1.000	0.810	0.944
ABTS	0.905	0.953	0.810	1.000	0.897
ORAC	0.936	0.901	0.944	0.944	1.000

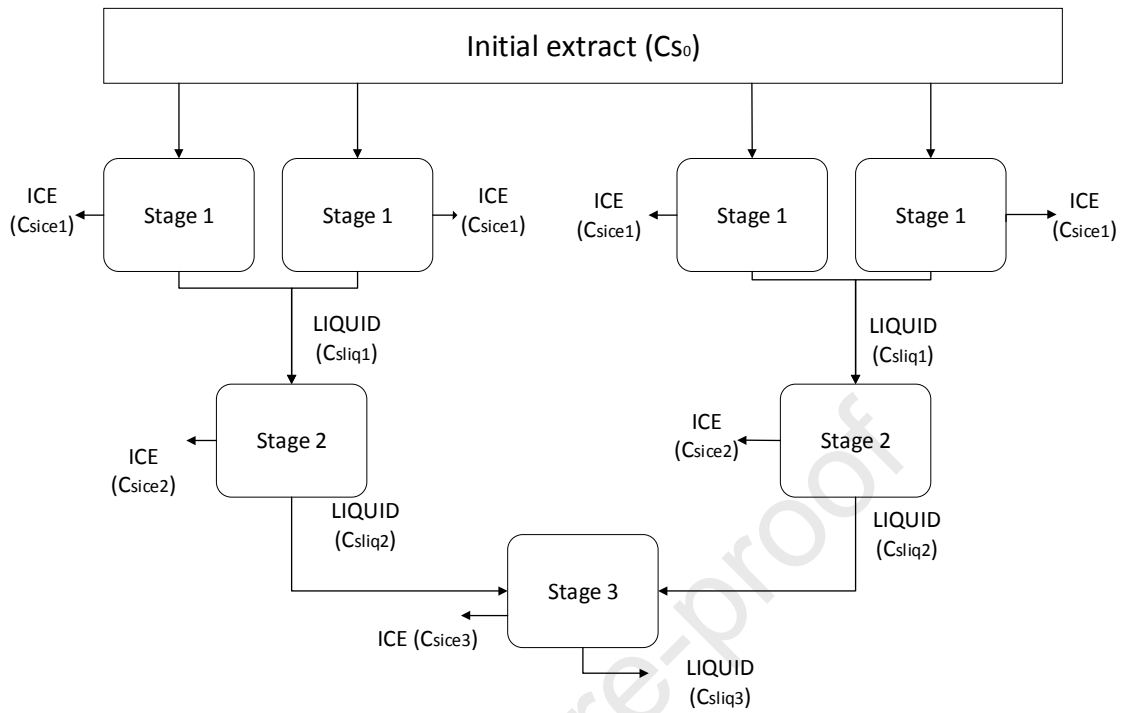


Figure 2. Multistage block freeze concentration process applied in the tests. C_{s0} (initial solids concentration), C_{sice} (solids concentration in the ice phase), and C_{sliq} (solids concentration in the liquid phase). ICE streams were discarded.

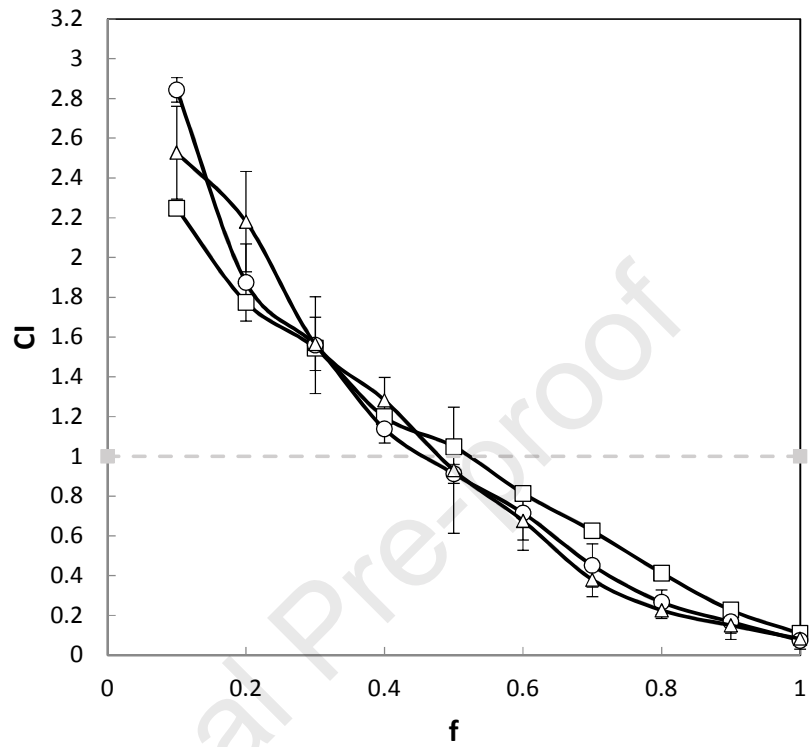


Figure 3. Concentration index as a function of thawing fraction; (□) First stage (○) second stage, and (△) third stage

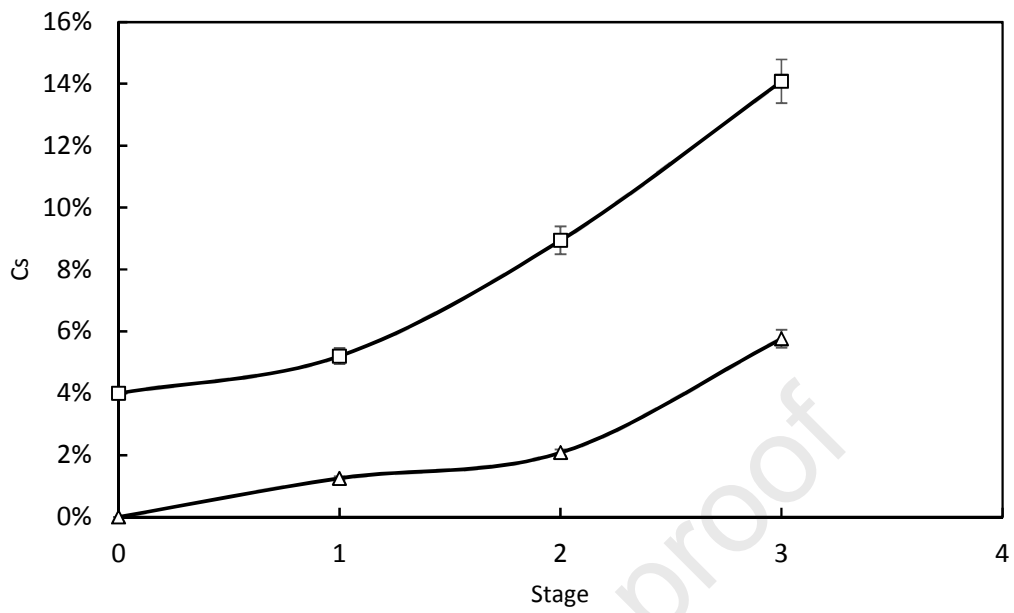


Figure 4. Solids concentration in liquid and ice at each BFC stage. (Δ) Ice phase and (\square) Liquid phase.

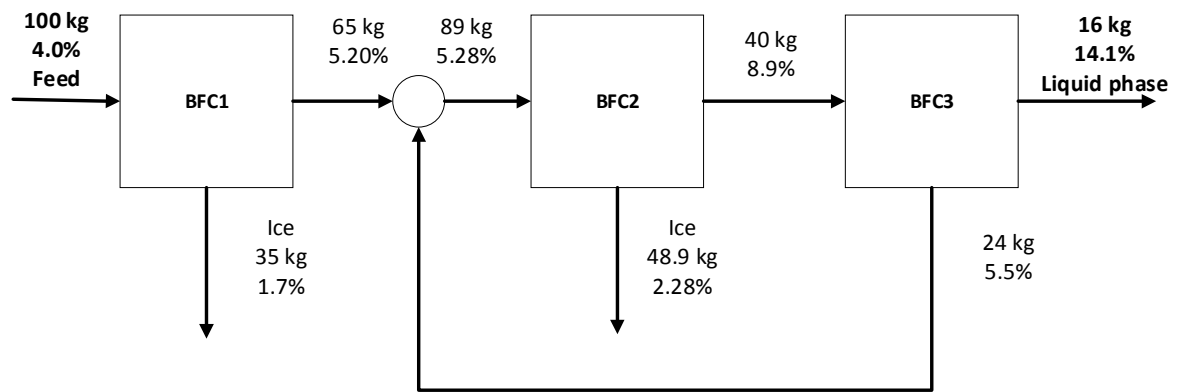


Figure 5. The proposed process of freeze-concentrating green tea extract using block freeze-concentration (BFC). Ice streams of stages 1 and 2 are final effluents. The liquid phase in the stage 3 is the final concentrated product. Solid concentration is expressed as % (w/w),

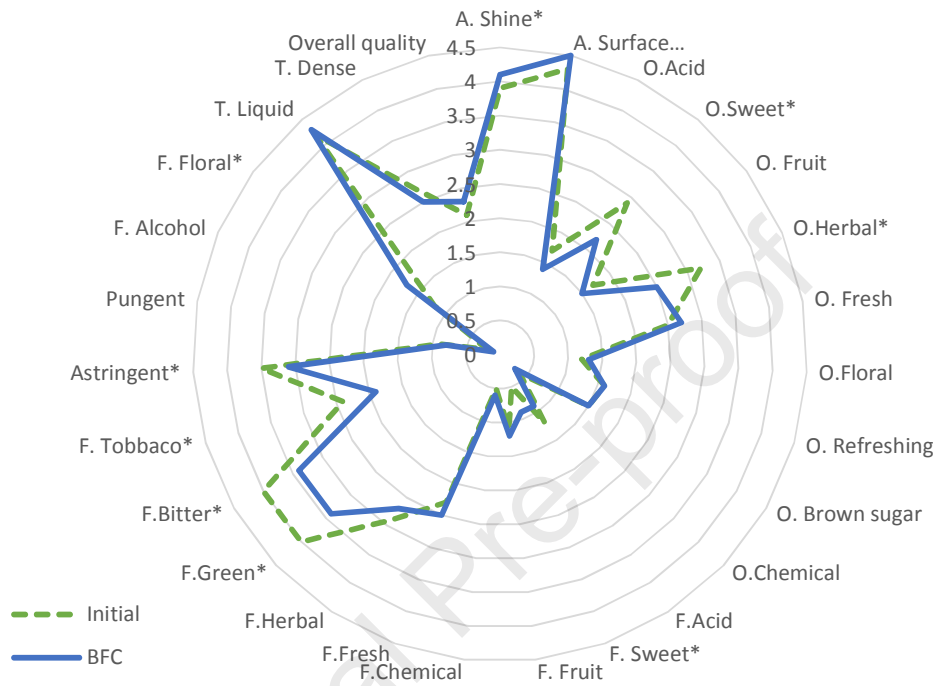


Figure 6. Sensory profile of the liquid after three stages of (—) BFC and (---) the initial green tea extract without treatment at the same solid content. Parameters marked (*) showed a significant difference between BFC and the initial extract.

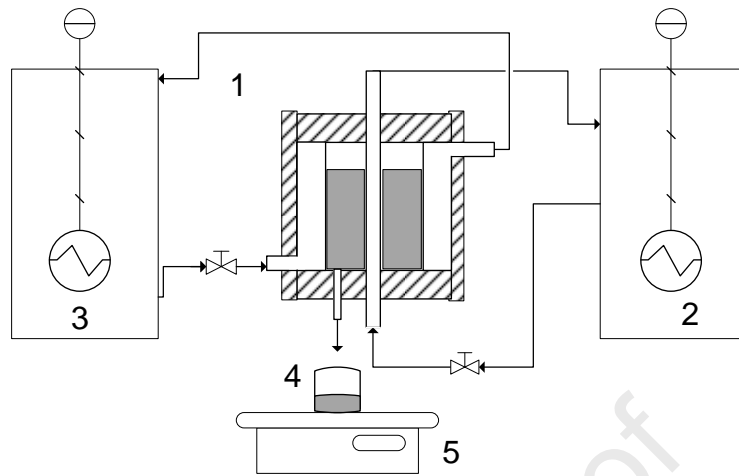


Figure 1. Experimental set-up for block freeze-concentration

Adapted from (Moreno et al., 2014a)

Highlights

Green tea extract was freeze-concentrated by the block technique.

The content of bioactive compounds was increased after the concentration.

The technique preserved the functional properties of the green tea extract.

The sensory attributes of the extract were preserved.

Journal Pre-proof

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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