

1 **Feasibility of S-rich streams valorization through a two-step**
2 **biosulfur production process**

3 M. Mora^{a*}, E. Fernández-Palacios^a, X. Guimerà^b, J. Lafuente^a, X. Gamisans^b,
4 D. Gabriel^a

5 ^aGENOCOV Research Group, Department of Chemical, Biological and Environmental
6 Engineering, Escola d'enginyeria, Universitat Autònoma de Barcelona, 08193 Bellaterra, Spain

7 ^bDepartment of Mining Industrial and ICT Engineering, Universitat Politècnica de Catalunya,
8 Avinguda de les Bases de Manresa 61-73, 08240 Manresa, Spain.

9 (*corresponding author: mabel.mora@uvic.cat)

10

11 **ABSTRACT**

12 A bioscrubbing process named SONOVA has been developed, tested and assessed herein
13 to valorize flue gases containing SO_x. The process consists in a first scrubbing stage, to
14 absorb and oxidize SO₂ to sulfate, followed by a two-step biological stage. It consists of
15 (1) an up-flow anaerobic sludge (UASB) reactor to reduce sulfate to sulfide with crude
16 glycerol and (2) a continuous stirred tank reactor (CSTR) to partially oxidize sulfide to
17 elemental sulfur (S⁰). SONOVA integrates the reutilization of resources, using the
18 effluent of the biological stage as a sorbent agent and the residual heat of flue gases to
19 dry the product. S⁰ is then obtained as a value-added product, which nowadays is
20 produced from fossil fuels. In this research, SO₂ concentrations up to 4000 ppm_v were
21 absorbed in 2 s of gas contact time in the spray-scrubber with removal efficiencies above
22 80 %. The UASB reduced up to 9.3 kg S-Sulfate m⁻³ d⁻¹ with sulfide productivities of 6
23 kg S m⁻³ d⁻¹ at an hydraulic retention time (HRT) as low as 2h. Finally, CSTR was fed
24 with the UASB effluent and operated at HRT ranging from 12h to 4h without biomass
25 wash-out. Sulfide was fully oxidized to S⁰ with a productivity of 2.3 kg S m⁻³ d⁻¹ at the

26 lowest HRT tested. Overall, this research has explored not only maximum capabilities of
27 each SONOVA stage but has also assessed the interactions between the different units,
28 which opens up the possibility of recovering S⁰ from harmful SO_x emissions, optimizing
29 resources utilization and costs.

30 **Keywords:** Biosulfur, bioscrubber, SO_x emissions, integral process, UASB, flue gas
31 valorization

32 **1. INTRODUCTION**

33 The combustion of sulfur containing matter results in SO_x formation which causes health
34 impacts, acid deposition in the environment and visibility depletion if released to the
35 atmosphere (EPA, 2019). Pigments industry, coal-based power generation plants or waste
36 incineration facilities are some examples of the industries generating SO_x containing
37 emissions. Pigments industry, in particular, utilizes a wide diversity of chemical
38 compounds in innumerable formulations to manufacture products such as Ultramarine
39 blue and Prussian blue, pigments which are mainly synthesized from sulfur and other
40 additives such as natural clay or copper, respectively (Eastaugh et al. 2007). The Global
41 pigments market size was valued at 20023 million USD in 2015 registering a Compound
42 Annual Growth Rate (CAGR) of 4.1 % during the forecast period 2022 (Nathwani, 2016).
43 Unfortunately, this industry generates harmful waste effluents since most of the
44 manufacture processes occur at extreme conditions of temperature (up to 1000 °C - 1200
45 °C). As an example, the synthesis of S based pigments generates large amounts of SO₂
46 emissions, besides other byproducts such as nitrogen oxides and particles, that must be
47 treated before their release to the atmosphere.

48 One of the most implemented treatment to remove SO_x from flue gases is wet flue gas
49 desulfurization (WFGD) which (1) is costly due to the use of alkaline absorbents and (2)
50 produces an alkaline sulfate-sulfite rich effluent that must be further treated before its

51 release to the environment (Srivastava and Jozewicz, 2001). In this sense, there is an
52 arising need to propose and develop environmentally friendly and feasible technologies,
53 both from technological and economical points of view, to valorize S-rich emissions. This
54 is the case of SO_x contained in flue gases or S-rich waste liquid effluents, that could be
55 transformed into elemental sulfur, a value-added product currently obtained from the
56 petrochemical industry. Still and all, the high operating costs of recovery processes,
57 together with demanding energy inputs, and in particular low market acceptance of the
58 recovered material, handicap resource recovery (Kataki et al., 2016).

59 To this aim, a promising integrated bioscrubbing process, namely the SONOVA process,
60 is proposed in this work to valorize SO₂ contained in flue gases. The process consists of:
61 (1) the scrubbing of SO₂ from flue gas using slightly alkaline absorbents, and the
62 treatment of scrubber liquor through (2) the biological reduction of sulfite and/or sulfate
63 to sulfide (using crude glycerol as carbon source and catalyzed by sulfate reducing
64 bacteria) and (3) the partial oxidation of sulfide to elemental sulfur (performed under
65 micro-aerophilic conditions and catalyzed by sulfide oxidizing bacteria). The biological
66 stage of this process (sulfate reduction and sulfide partial oxidation) could be also applied
67 to the treatment of sulfate-laden effluents, even without organic content, such as pulp and
68 paper industry effluents or saline wastewater. The competitiveness of the overall
69 bioscrubbing process proposed in this work is based on the use of biological processes to
70 recover elemental sulfur as a value added product reusing energy (energy contained in
71 flue gases can be reused for elemental sulfur drying) and resources (the effluent from the
72 bioscrubbing process can be reused for boosting SO_x absorption). In general, the most
73 well-known application of S-rich effluents valorization, instead of to recover elemental
74 sulfur, is to treat acid mine drainage (AMD) since this type of waste contains metals,
75 commonly precious or heavy metals, and sulfate. The biological reduction of sulfate

76 contained in AMD allows precipitating metals with sulfide to afterwards be valorized or
77 not depending on the added value of the recovered metals (Christensen et al., 1996; Jong
78 and Parry, 2003; Neculita et al., 2007). Apart from this last application, only few studies
79 can be found in the literature focused on sulfur recovery from SO_x emissions or S-rich
80 effluents (Chuang et al., 2005; Jiang et al., 2013; Sun et al., 2018; Zhang et al., 2018),
81 and most of them require either using easily biodegradable and/or expensive carbon
82 resources or treating low S loads (< 2 kg m⁻³ d⁻¹). Jiang et al. (2013) reported excellent
83 results applying the SANI process although required sewage and nitrogen sources to
84 valorize S-containing effluents. Zhang et al. (2018) reported a smart alternative to the
85 SANI process, targeting biosulfur production, to treat specifically sulfate-deficit
86 wastewater and also required the use of domestic wastewater for sulfate reduction; and
87 Sun et al. (2018) developed a novel bio-FGD system configuration to eliminate SO₂ and
88 NO from flue gases although requiring the application of a quartz sulfur condenser to
89 separate the final product and concluding that, the process could be profitable if the
90 operational parameters could be further optimized and if wastewater was available. Other
91 studies propose both biological steps (sulfate reduction and sulfide oxidation) in a single
92 reactor with successful results and less investment costs but probably with complex
93 downstream procedures for sulfur recovery or low sulfur purities (Philip and Deshusses,
94 2003; Xu et al., 2012).

95 Regardless the product targeted from S-rich effluents valorization, the supply of an
96 organic carbon source is required if the first biological step, i.e. the reduction of sulfate,
97 is performed heterotrophically. Many organic sources have been used in other studies to
98 reduce sulfate such as VFAs (Celis-Garcia et al., 2007; Garcia-Solares et al., 2014),
99 ethanol (Nevatalo et al., 2010) or lactate (Bertolino et al., 2014) among others (Liamleam
100 and Annachhatre, 2007). However, industrial applications are every time more limited by

101 the costs of the carbon source to get not only technically feasible processes but also
102 economically feasible processes. The use of wastes is an economical alternative to
103 efficiently reduce sulfate (Jian et al., 2013; Barrera et al., 2014; Fernandez-Palacios et al.,
104 2019; Mora et al., 2018; Sahinkaya et al., 2013). As an example, crude glycerol, an
105 organic waste that has been traditionally valorized as biogas through anaerobic digestion
106 (Lopez et al., 2009; Viana et al., 2012) or as biohydrogen through dark fermentation
107 (Haron et al., 2018), has been also proven to be a potential electron donor for sulfate
108 reduction (Mora et al., 2018; Santos et al., 2018; Fernández-Palacios et al., 2019). Crude
109 glycerol is an attractive waste for being an inexpensive carbon source and highly available
110 since it is the byproduct of the biodiesel production process. As reported by Kolesarova
111 et al. (2011) and Yang et al. (2012) a surplus of crude glycerol is forecasted due to the
112 growth of the biodiesel industry. Crude glycerol is also impure since it can contain
113 methanol, soaps, FAMES (fatty acid methyl esters) and inorganic elements (calcium,
114 phosphorous, magnesium, sulfur). However, in the SONOVA process the utilization of
115 crude glycerol avoids the supplementation of nutrients and trace metals to the anaerobic
116 sludge, which are generally necessary in anaerobic digestion processes (Singh et al.,
117 1999), since they are all contained in the organic source.

118 Regarding partial sulfide oxidation (the last step of the SONOVA process) several works
119 have been published reporting elemental sulfur recovery with suspended biomass
120 (Krishnakumar et al., 2005; Zytoon et al., 2014; Zhang et al., 2018). However, few of
121 them treats effluents from sulfate reduction processes using waste organic effluents,
122 which implies the presence of by-products from biological degradation and, frequently,
123 high free ammonia loads besides the absence of nitrate or nitrite. Moreover, the quality
124 of the elemental sulfur produced is not usually determined; neither proposed or tested the
125 downstream procedure to recover and purify biosulfur, which is important since it is the

126 final product of the process. This is the main reason why sulfide oxidizing biomass is
127 cultivated as a suspended culture in a continuous stirred tank reactor (CSTR) in this study,
128 instead of being cultivated as an immobilized culture.

129 Then, the aim of this research was to demonstrate the feasibility of a bioscrubbing process
130 to valorize SO₂ contained in flue gases through the development, operation and
131 optimization of a sequential Scrubber-UASB-CSTR system to obtain a valuable product,
132 as is elemental sulfur, using crude glycerol. The process proposed in this work is
133 challenging since it targets not only the optimization of SO₂ absorption but also the sulfate
134 reduction minimizing the production of methane. The process also couples the cultivation
135 of suspended sulfide oxidizing biomass to separate more efficiently the elemental sulfur
136 produced. Furthermore, the development of this research allowed obtaining the maximal
137 capabilities of each of the stages besides valuable data from different operating scenarios.

138 **2. MATERIALS AND METHODS**

139 **2.1 Experimental setup and assays performed to assess SO₂ absorption efficiencies**

140 A lab-scale spray-scrubber was constructed and operated separately to characterize the
141 wet-absorption of SO₂ under different operating conditions. Detailed information and the
142 schematic of the absorption unit is presented in the Supplementary Material (Section S1,
143 Figure S1).

144 A series of absorption assays were performed with two different sorbents (1) slightly
145 alkaline solution and (2) effluent of the biological process. The second sorbent was used
146 in order to evaluate the effect of its composition over the SO₂ absorption efficiency
147 compared to that obtained with the alkaline solution. The characterization of SO₂ wet-
148 absorption was performed by assessing the influence of both the gas contact time and gas-
149 absorbent (L/G) ratio. The experimental conditions of wet-absorption tests are shown in
150 Table 1.

151

152 **Table 1.** Experimental conditions tested during the characterization of SO₂ absorption

Absorbent	Inlet SO₂ (ppmv)	L/G (L/m³)	Gas contact time (s)	pH_{initial}
Alkaline absorbent	2000	5 - 15	0.6 - 4	8.0
Bioprocess effluent	2000 - 4000	5 - 15	0.6 - 4	7.4

153

154 Since the SONOVA process considers the recovery of energy from flue gases to obtain a
155 dry bioproduct, the temperature of flue gases to be valorized is anticipated to be below
156 80°C. A set of experiments was previously performed at temperatures ranging from 40°C
157 to 80°C to assess the SO₂ removal efficiency (see Table S1). It was proven that under the
158 conditions set in the present study, temperature was not affecting the performance of the
159 absorption unit and hence this parameter was not controlled.

160 A Labview interface was used for the monitoring of SO₂ concentration at the outlet gas
161 and pH and ionic conductivity in the liquid effluent. Experiments were performed until
162 constant SO₂ concentration, feeding fresh absorbents continuously to ensure constant pH
163 and composition in the liquid phase during SO₂ absorption. The performance of the SO₂
164 absorption process was evaluated in terms of removal efficiency. To ensure experiments
165 accuracy, three replicate measurements were made, obtaining relative errors below 5%.
166 In this paper only average values are shown.

167 **2.2 Experimental setup and operating conditions for biosulfur production**

168 The experimental setup for biosulfur production, from a mimic of S-rich wastewater,
169 consisted of a sequence of two reactors (Figures S2 and S3, Supplementary Material).
170 The first reactor was an up-flow anaerobic sludge bed (UASB) reactor for sulfate
171 reduction with a total volume of 2.5L and an effective volume (sludge bed volume) of
172 1L. Start-up and initial operation of the UASB were performed as described in a previous

173 work (Fernández-Palacios et al., 2019). As previously reported (Mora et al., 2018), the
174 optimal conditions found to promote sulfidogenesis from a methanogenic granular sludge
175 using crude glycerol as the organic carbon source were: T=35°C, pH=8.5, COD/S-Sulfate
176 ratio above 7.0 g O₂ g⁻¹ S-SO₄²⁻, microaerophilic conditions and sulfide concentrations
177 under 250 mg S²⁻ L⁻¹. Nevertheless, COD/S-Sulfate ratio in the influent was set below 6
178 g O₂ g⁻¹ S-SO₄²⁻ since it has been demonstrated that the limitation of COD promotes
179 sulfidogenesis and minimizes biogas production in sulfidogenic UASB reactors (Mora et
180 al., 2018). The UASB was fed with synthetic mineral medium for a better evaluation of
181 biosulfur production from a controlled sulfate-rich influent. Detailed information about
182 the experimental setup and operating conditions can be found in the Supplementary
183 Material (section S2).

184 The second reactor was a glass-made continuous stirred tank reactor (CSTR) with a
185 volume of 6 L. The CSTR was used for biosulfur production, under microaerophilic
186 conditions, from the partial oxidation of sulfide produced in the UASB. Then, the influent
187 of the CSTR was the effluent of the UASB. Moreover, the supernatant of the CSTR
188 effluent was tested as liquid absorbent for SO₂ removal in the absorption unit (Table 1,
189 bioprocess effluent). Microaerophilic conditions in the CSTR were assured by
190 transferring the oxygen from the headspace to the liquid phase through mechanic
191 agitation. The pH in the reactor was monitored and registered.

192 The startup of the UASB-CSTR system was performed with two different inocula to
193 speed up the startup of both reactors with specific biomass. The UASB was inoculated
194 with settled granular sludge from an anaerobic digester treating wastewater in a paper
195 recycling company. As reported by Pol et al. (2004), the use of granular sludge allows
196 loading rates in UASB reactors far beyond the common loading rates applied in
197 conventional activated sludge processes due to two main factors: a) the superior settling

198 characteristics and b) the high specific activities of granular sludge. On the contrary, the
 199 CSTR was inoculated with 6 L of sulfide oxidizing sludge obtained from a bioscrubber
 200 for biogas desulfurization.

201 The abovementioned reactors were operated under pseudo-steady conditions in order to
 202 assess the feasibility of the process. Two different sets of short-term assays were
 203 performed in the experimental setup to explore the limits of the two-step bioprocess. The
 204 first set was performed in the UASB during 60 h and consisted of varying the sulfate inlet
 205 concentration every 12 h (from 100 mg S L⁻¹ to 1000 mg S L⁻¹) maintaining a COD/S-
 206 Sulfate ratio of 15 g O₂ g⁻¹ S. The second set of assays was performed in the CSTR during
 207 4 days and consisted of decreasing the HRT (from 12 h to 4 h) by changing the liquid
 208 level and hence the reaction volume in the CSTR. In Table 2 the summary of the operating
 209 conditions and the short-term assays conditions are presented.

210

211 **Table 2.** Conditions tested in the two-step experimental setup for biosulfur production

	Period I	Period II	Ending period	SLR Assays	HRT** Assays
Reactor	UASB	UASB	UASB	UASB	CSTR
Time (d)	0 - 99	99 - 115	115-120	65	117-120
HRT_{UASB} (h)	1.81±0.25	2.19±0.14	2.29±0.25	1.74	2.14±0.18
HRT_{CSTR} (h)	12	12	12	12	12 8 6 4
S inlet (mg S L⁻¹)	235±17		451±7	100 250 500 750 1000	398±2

SLR (kg S m ⁻³ d ⁻¹)	2.42±0.54	2.47±0.39	4.63±0.48	1.4	
				3.5	0.8
				7.0	1.2
				10.5	1.6
				14	2.4
				22	
OLR (kg O ₂ m ⁻³ d ⁻¹)	10.23±2.29	12.72±1.12	15.3±1.63	53	2.8
				98	4.2
				151	5.5
				200	8.3
COD/S (g O ₂ g ⁻¹ S)	3.90±0.89	5.35±0.61	5.63±0.68	15	3.5

212 *SLR and OLR are sulfate and organic loading rates calculated taking into account the
 213 granular sludge bed volume. **Values calculated for HRT assays in the CSTR take into account
 214 the inlet sulfide and COD concentrations contained in the UASB effluent.

215

216 The ending period consisted of increasing both the SLR and OLR in the system,
 217 maintaining the COD/S-Sulfate ratio, to increase the COD concentration in the effluent
 218 of the UASB and, subsequently, to assess the COD removal efficiency in the CSTR
 219 during the HRT assays. The SLR range treated in the CSTR during the HRT assays is
 220 referred to the sulfide loading rate considering the concentration of sulfide present in the
 221 effluent of the UASB during the HRT assays. The OLR range treated in the CSTR during
 222 the HRT assays was also calculated based on the COD concentration present in the
 223 effluent of the UASB during the HRT assays.

224 **2.3 Jar tests for biosulfur precipitation and sedimentation**

225 A series of jar tests were performed with different combinations of coagulants and
 226 flocculants to assess the precipitation and sedimentation efficiency of biological sulfur.
 227 Each test was performed at room temperature (18-20 °C) with 200 mL of effluent from
 228 the CSTR (pH = 7.4). Agitation cycles of 1 min mixing at 200 rpm followed by 1 min
 229 mixing at 70 rpm and 15 min without mixing were applied to evaluate the performance

230 of each additive added to the samples. Two different cationic coagulants were initially
231 tested, one with linear molecular structure (FL4820) and another with slightly branched
232 molecular structure (FL3050) both supplied by SNF Group, France. Five different
233 concentrations of each coagulant were applied (from 0.1 to 1 %), as recommended by the
234 supplier. The supernatant turbidity was also analyzed at the beginning and at the end of
235 each test to assess the efficiency of the process. Afterwards, the coagulant showing better
236 results, in terms of % settled solids, was combined with different dosages of cationic
237 (from 0.03 to 0.3 % of FO435, SNF Group, France) and anionic (from 0.1 to 1 % of
238 HIMOLOC, Derypol, Spain) flocculants, as recommended by the suppliers. Settled
239 solids, obtained from those tests showing optimal results, were also analyzed to determine
240 sulfur purity and hence to assess the effect of coagulant and flocculant addition to the
241 quality of the final product.

242 **2.4 Analytical methods**

243 *2.4.1 Liquid phase analyses*

244 Sulfate (SO_4^{2-}) and thiosulfate ($\text{S}_2\text{O}_3^{2-}$) concentrations were analyzed off-line by ion
245 chromatography using suppressed conductivity detection (ICS-2000, Dionex
246 Corporation). Chemical Oxygen Demand (COD) was analyzed off-line using COD
247 commercial kits and a photometer (Lovibond®). Ionic conductivity and pH were
248 continuously measured in the scrubber with a multi-analytic probe (Multi 3420, WTW,
249 Germany). Oxidation-reduction potential (ORP) and pH were monitored in the UASB
250 and CSTR with probes (Crison pH5333 and ORP5353, Hach Lange, Spain) connected to
251 a multimeter (Crison MM44, Hach Lange, Spain). Sulfide concentration was analyzed
252 off-line with a sulfide selective electrode connected to a benchtop meter (Symphony,
253 VWR) after conditioning the samples with sulfide antioxidant buffer (SAOB). The SAOB
254 composition was (g L^{-1}): ascorbic acid (35) and EDTA (67) dissolved in NaOH (2M).

255 VFA were analyzed by gas chromatography (7820A, Agilent Technologies) equipped
256 with a DB-FFA column and using a flame ionization detector (FID) with helium as carrier
257 gas. Prior to VFA analyses, samples were prepared as follows (Montpart, 2014). A
258 volume of 0.6 mL of sample was pipetted to a 1.5 mL glass vial. Afterwards, 0.15 mL of
259 a preserving solution and 0.75 mL of deionized water were added to the vial and kept at
260 -20 °C until analyzed. The preserving solution was meant to deproteinize the sample and
261 to be used as internal standard. It contained (g/L): HgCl₂ (2), ortophosphoric acid (33.7)
262 and hexanoic acid (2) (internal standard).

263 *2.4.2 Gas phase analyses*

264 SO₂ concentration in the outlet gas phase of the scrubber was performed using an
265 electrochemical SO₂ sensor (SO₂-B4, Alphasens, UK) with a measuring range from 0 to
266 2000 ppmv. The measurement of higher SO₂ concentrations required flue gas dilution,
267 performed by two flowmeters (2100, Tecfluid, Spain). CH₄, CO₂ and H₂S contained in
268 the gaseous effluent of the UASB were analyzed by gas chromatography (HP 5890,
269 Hewlett Packard) equipped with a Porapack Q column and a thermal conductivity
270 detector (TCD) with helium as carrier gas. The volume of the gas produced in the UASB
271 reactor was analyzed following the Gas Bag Method (GBM) presented in Ambler and
272 Logan (2011) using gas chromatography (7820A, Agilent Technologies) equipped with
273 an HP-Mole Sieve column, a thermal conductivity detector (TCD) and using argon as the
274 carrier gas. The GBM procedure consisted basically of measuring the initial composition
275 of the collected gas in the bag, adding a known volume of tracer gas (nitrogen gas in this
276 case) and analyzing the new composition. From these two analyses (before and after the
277 tracer injection) the initial volume of gas could be calculated from mass balances (Baeza
278 et al., 2017).

279

280 *2.4.3 Solid phase analyses*

281 The sludge contained in the riser of the UASB was sampled after 50 days of operation
282 and was analyzed, together with a biosulfur sample, to obtain the granular size distribution
283 (GSD) using laser diffraction based technology (Mastersizer 2000, Malvern Panalytical
284 Ltd.) and total and volatile suspended solids (TSS and VSS, respectively) following the
285 standard method analysis (APHA, 2005). Biosulfur (centrifuged sample from the CSTR)
286 and sedimented solids from jar tests were also analyzed to obtain the main characteristics
287 of the product. Sulfur purity was determined from the elemental analysis of centrifuged
288 and lyophilized samples with a Flash EA 2000 CHNS instrument (Thermo Fisher
289 Scientific) connected to a microbalance (MX5, Mettler Toledo). In the case of biosulfur,
290 a thermogravimetric analysis (TGA) was also performed using a differential scanning
291 calorimeter (449 F1 Jupiter, NETZSCH) in order to assess the effect of heat and air over
292 the stability of biosulfur. DSC allows measuring enthalpy changes in samples due to
293 changes in their physical and chemical properties as a function of temperature or time.
294 Information about the melting point of biosulfur or the energy associated to its oxidation
295 can be obtained with TGA.

296 **3. RESULTS AND DISCUSSION**

297 **3.1 Technical feasibility and operation limits of the biological units**

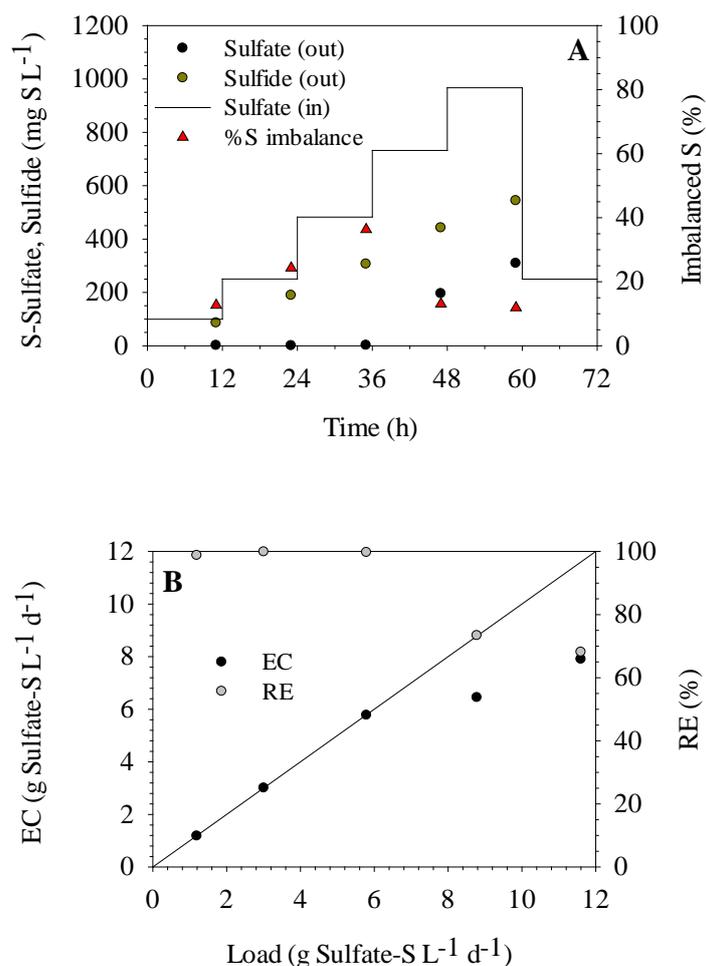
298 The technological feasibility of the sequential UASB-CSTR experimental setup utilized
299 for biosulfur production was assessed in this work. Both reactors were connected from
300 the beginning of the operation and their performance monitored during 120 days. In the
301 Supplementary Material (section S3) the overall performance of the biological units is
302 described in detail.

303 Short-term SLR assays (Table 2) were performed after 65 days of operation when the S-
304 RE achieved in the UASB was 100 %. The results obtained from this first set of short-

305 term assays in the UASB are presented in Figure 1. As can be observed in Figure 1A, the
306 S-EC of the UASB reactor, operated at such a low HRT (2h) and under non limiting COD
307 availability, allowed treating inlet concentrations of sulfate as high as 1000 mg S L⁻¹ with
308 68.1 % S-RE (maximum elimination capacity of 389 mg S L⁻¹ h⁻¹ or 9.3 kg S m⁻³ d⁻¹).
309 This value is comparable to that obtained by Bijmans et al. (2008) (9.7 kg S m⁻³ d⁻¹) using
310 formate and higher than the S-EC obtained by Nagpal et al. (2000), Nevatalo et al. (2010)
311 and Kaksonen et al. (2006) using ethanol at 30-35 °C (S-EC up to 3.8 kg S-Sulfate m⁻³ d⁻¹).
312 The high S-EC obtained in this work is most probably related with the absence of the
313 liquid phase recirculation during the UASB operation. The presence of sulfide in the
314 recirculation could interfere negatively in the hydrolysis of crude glycerol, mainly
315 happening in the first part of the sludge bed as previously reported by Fernandez-Palacios
316 et al. (2019), worsening the performance of the UASB.

317 Regarding the sulfur mass balance it was observed that sulfide production did not
318 correspond to sulfate reduction. As can be observed from the experimental data presented
319 in the Supplementary Material (section S3), sulfur imbalance tends to increase when COD
320 increases. This phenomenon is caused by the production of organosulfur compounds,
321 which leads to require further research in order to enhance the yield of sulfate reduction
322 to sulfide, instead of to other intermediate compounds. The production of intermediate
323 compounds, such as organosulfur compounds, causes a decrease of elemental sulfur
324 productivity and an increase of oxygen consumption in the CSTR.

325



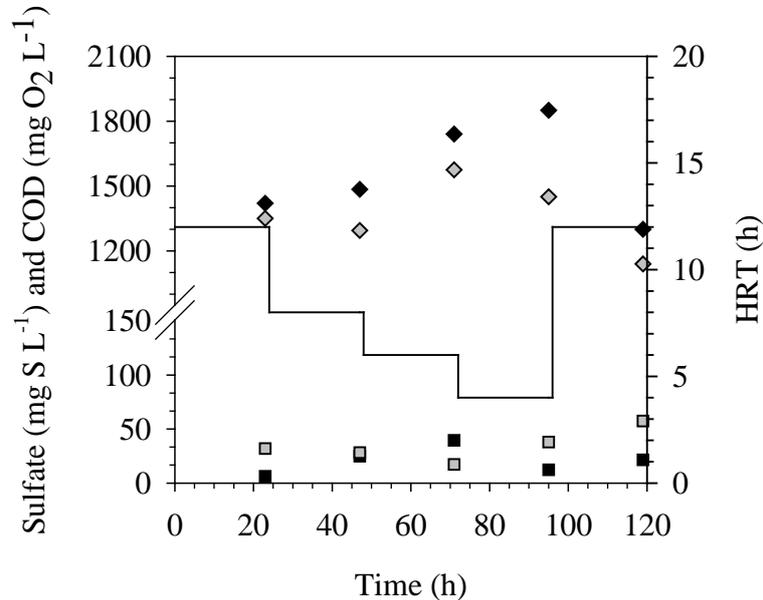
326 **Figure 1.** Sulfate loading rate assays performed in the UASB. A) Sulfate and sulfide
 327 concentrations in each hydraulic steady state and B) S-EC and S-RE obtained in each hydraulic
 328 steady state.

329

330 On the other hand, the HRT assays (Table 2) were performed at the end of the operation,
 331 once the SLR and OLR were doubled (Ending period, Table 2). The aim of this last
 332 operation period was to obtain higher COD concentration in the effluent of the UASB
 333 since until day 115 the COD was almost depleted (section S3, Figure S4B, Supplementary
 334 Material). The COD increase allowed assessing 1) the competence between heterotrophic
 335 and autotrophic bacteria and 2) the COD removal efficiency (COD-RE) in the CSTR

336 under different HRT. In Figure 2, the results obtained from this second set of short-term
337 assays are presented.

338



339

340 **Figure 2.** HRT assays performed in the CSTR. HRT (solid line), sulfate in the influent (black
341 square), sulfate in the effluent (grey square), COD in the influent (black diamond) and COD in
342 the effluent (grey diamond).

343

344 Sulfide was not detected at any HRT tested. Sulfate in the effluent of the CSTR was
345 similar or higher than sulfate in the influent which indicated the full capacity of the culture
346 to oxidize partially sulfide to sulfur. Sulfur productivities up to 95 mg S L⁻¹ h⁻¹ (or 2.3 kg
347 S m⁻³ d⁻¹) were obtained at the lowest HRT tested (4 h). COD was also monitored and it
348 was observed that the CSTR had a certain capability of COD degradation although the
349 maximum removal efficiency calculated was 26.3 % at the lowest HRT tested. On the
350 one hand, this result could indicate that heterotrophic bacteria were outcompeting SOB
351 for the oxygen. However, the high dilution rate set in the CSTR together with the
352 increasing SLR could also improve the adsorption of the organic matter on the surface of
353 the elemental sulfur particles (Meerburg et al., 2015). As has been reported in other

354 studies, biosulfur has hydrophobicity and contain organic compounds on its surface such
355 as long-chain polythionates or proteins (Kleinjan et al., 2003). Then, biosulfur
356 composition and structure probably enhanced COD adsorption improving COD-EC and
357 COD-RE at such a low HRT (4 h).

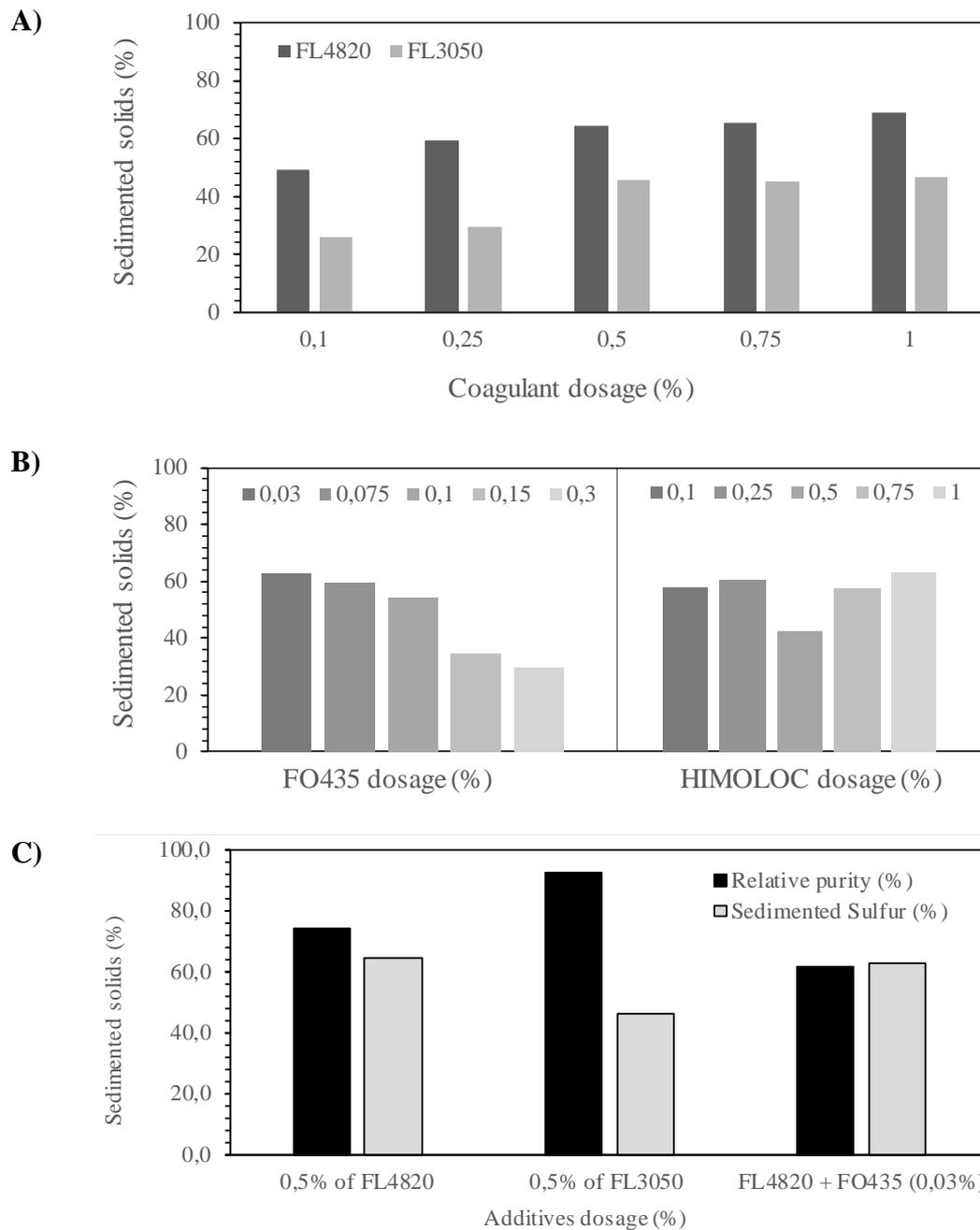
358 VSS concentration in the CSTR at HRTs of 12 h and 4 h was 660 ± 10 mg L⁻¹ and 555 ± 7
359 mg L⁻¹ which confirmed, together with the absence of sulfide and the production of
360 sulfate, that the biomass was not washed-out at such a low HRT (4 h). The TSS and VSS
361 concentrations at each HRT had alike values indicating that the suspended matter sampled
362 in the CSTR was only organic (mainly biomass and sulfur).

363 **3.2 Biosulfur quality and recovery process**

364 Biosulfur was analyzed to obtain more information about the characteristics of the
365 product and to propose an adequate procedure for its recovery. The analysis of the particle
366 size distribution was performed as a first step. From this analysis a volume weighted mean
367 of 39.8 μm, with a specific surface area of 0.748 m² g⁻¹, was obtained. This size was much
368 bigger (10x) than that reported by Janssen et al. (2009) and Kostrytsia et al. (2018) for
369 biosulfur particles, although biosulfur produced in this study did not show any settling
370 capacity in 1 h of Imhoff test.

371 The elemental analysis of biosulfur was also performed to obtain its purity and organic
372 content. Two samples of suspended biosulfur were centrifuged at 10000 rpm during 2
373 min. The humidity content measured after the centrifugation of both samples was 46 %.
374 Then, the first sample was lyophilized and homogenized (S₁, named raw biosulfur). The
375 second sample was also lyophilized and homogenized after the layer of biomass deposited
376 on the centrifuged sulfur was removed (S₂). The result of the elemental analysis is
377 presented in Table S2 (section S4, Supplementary Material).

378 Sulfur content in S_1 was 87.5 %, which could be already adequate for some applications
379 of this product. Instead, the sulfur purity in S_2 was almost 95 % with carbon content lower
380 than 3.5 %. This result indicates that an effective centrifugation of both solids could be
381 an adequate treatment to have a biosulfur with high quality; however, costs associated to
382 this specific separation process are considerably higher than those required for
383 sedimentation. As mentioned above, raw biosulfur sedimentation through the addition of
384 chemical agents (coagulants and flocculants) is an economical alternative to
385 centrifugation. In this case, Jar tests were certainly helpful to evaluate the % of biosulfur
386 recovery (% of sedimented solids) and its final purity (Figure 3) after a coagulation-
387 flocculation-sedimentation process. As can be observed in Figure 3A, the addition of the
388 coagulant FL4820 had better results in terms of percentage of sedimented solids (SS)
389 compared to coagulant FL3050. FL4820 presented at least 50 % of SS with the lowest
390 dosage (0.1 %), while the addition of FL3050 uniquely generated 46.8 % of SS at the
391 highest dosage tested (1 %). Although the optimal dosage of FL4820 was 0.5 % (64.4 %
392 of SS), flocculation tests (Figure 3B) were performed with the maximal dosage tested
393 since that was the one showing the highest % of SS (68.8%). In Figure 3B it can be
394 observed that the addition of flocculants did not improve the % of SS. Results obtained
395 with FO435 showed that the addition of cationic flocculants even worsened the % of SS,
396 which was 49.3 % with a dosage of 0.03 % FO435. A different behavior was observed
397 with the addition of HIMOLOC; nevertheless, the maximum % of SS (63.3 % of SS)
398 obtained at the highest dosage (1 %) was still lower than that obtained adding uniquely
399 the coagulant FL4820. These results highlighted the positive effect that minimal dosages
400 of coagulant have over the sedimentation of solids without requiring other additives.
401



402 **Figure 3.** Effect of coagulants and flocculants over the sedimentation and quality of biosolids
 403 produced in the CSTR. A) Addition of coagulants (FL4820 and FL3050) and B) addition of
 404 flocculants (FO435 and HIMOLOC) combined with 1 % of FL4820 and C) comparison of purity
 405 and % of settled sulfur obtained from the tests showing best results.

406

407 Other studies have been conducted to investigate the colloidal biogenic sulfur

408 flocculation. However, it is still a great challenge. As an example, Chen et al. (2015)

409 obtained successful results under different conditions than those set in this study. They
410 found that the biogenic sulfur could be efficiently flocculated and separated although at
411 pH 4.73, stirring speed of 129 r/min and a flocculant dose of 2.42 mg polyaluminum
412 chloride mg^{-1} S. Jansen et al. (1996) also reported the difficulties associated to the
413 aggregation of biosulfur (due to the multiple functional groups that it has) and the need
414 to operate sulfide-oxidizing bioreactors at high loading rates to enhance the settleability
415 of the sulfur sludge. Yuan et al. (2014) stated that the colloidal nature of biosulfur requires
416 the addition of cationic coagulants to be separated. This is the same result obtained in the
417 present study although worst % of sedimented solids have been obtained, probably due
418 to the characteristics of the UASB effluent (salts, organic matter and organosulfur
419 compounds content). As can be observed, a wide range of results about the recovery of
420 biogenic sulfur are available in the literature indicating that there is not a definitive
421 solution for all the biosulfur producing systems.

422 Settled solids from tests performed with 0.5 % of coagulant addition (both FL4820 and
423 FL3050) and with FL4820 and 0.03 % of FO435 were analyzed to obtain the sulfur purity
424 (Figure 3C). The elemental analysis of the settled biosulfur confirmed that the addition
425 of coagulant and flocculants presented better % of SS than those obtained with the unique
426 addition of FL3050, although the sulfur purity in the product was not so satisfactory (61.5
427 % sulfur purity). Dosages of 0.5 % of FL3050 presented high sulfur purities, nevertheless
428 the % of SS was only 45.9 %. Then, the best performance was obtained with the addition
429 of FL4820, which allowed obtaining a product with a sulfur purity of 74.3 % and 64.4 %
430 of SS. However, a better approach is required to separate as much biosulfur as possible
431 with the less investment in chemicals dosage which worsen the economic benefits of the
432 process and the quality of the final product. Then, further studies should be conducted in

433 order to test tailor-made coagulants and flocculants for this specific application in order
434 to achieve higher % of SS and biosulfur purities.

435 Finally, the thermogravimetric analysis of non-lyophilized S₁ was performed since
436 lyophilization is not considered an economically feasible alternative to treat a product
437 without an important added value such as pharmaceutical or food products. To obtain
438 dried biosulfur a thermal treatment should be done. However, biosulfur could be melted
439 and oxidized to SO₂ when heated in the presence of air. For this reason, the
440 thermogravimetric analysis was performed not only with air but also with an inert gas. In
441 Figure S6 (section S5, Supplementary Material) the results from the gravimetric analysis
442 are presented. Profiles shown in Figure S6A (thermogravimetry performed with inert gas)
443 revealed that the melting point of biosulfur was 125°C and, also, that biosulfur was
444 volatilized over 175°C. From Figure S6B (air) it was observed that, biosulfur was
445 oxidized to SO₂ at 225°C (enthalpy of -4513 J g⁻¹). Then, it was concluded that an
446 appropriate process to recover biosulfur from the CSTR could be a sequence of settling
447 after coagulant addition and air drying at T<100°C. However, further research should be
448 done in order to obtain higher % of biosulfur recovery and to assess the effect of chemical
449 agents addition over the characteristics of the final product.

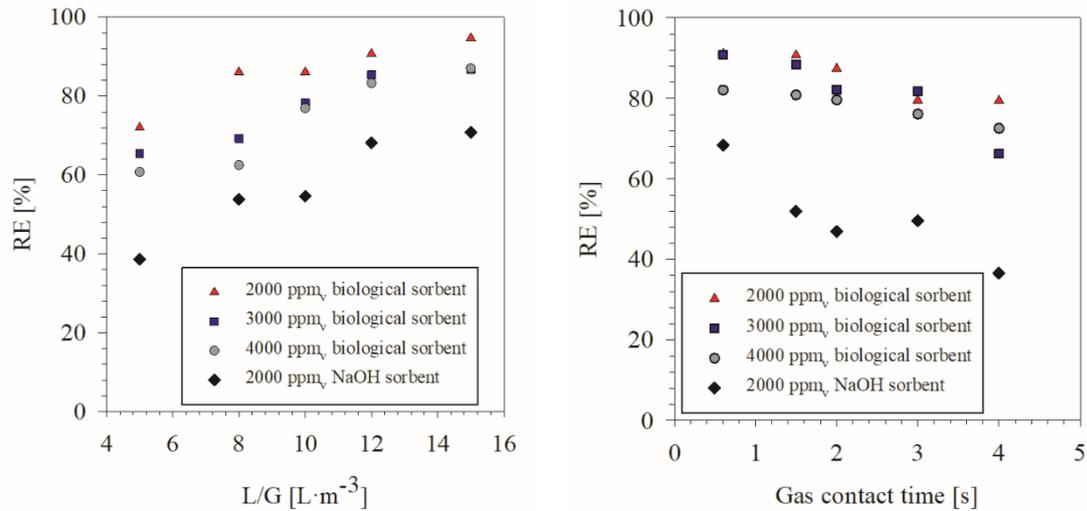
450 **3.3 Assessment of the SO₂ absorption system**

451 Results obtained from the absorption tests performed in the lab-scale spray scrubber under
452 different conditions are presented in Figure 4. It can be observed that L/G ratio and gas
453 contact time had a strong influence over the RE for each SO₂ concentration and type of
454 sorbent tested.

455

A)

B)



456 **Figure 4.** Removal efficiencies of SO₂ absorption. A) At a gas contact time of 0.6 s depending
 457 on the L/G ratio and B) at a L/G ratio of 7.5 L m⁻³ depending on the gas contact time.

458

459 The RE of 2000 ppm_v of SO₂ using a slightly alkaline absorbent prepared from a NaOH
 460 solution at pH 8.0 rose from 38.7 to 70.9% when L/G ratio was increased from 5 to 15 L
 461 m⁻³. The value of RE obtained in the spray-scrubber revealed low mass transfer of SO₂
 462 from gas to liquid phase at a gas contact time of 0.6 s. The increasing volumetric flowrate
 463 of the absorbent increased the diameter of the droplets inside the scrubber and thus the
 464 gas-liquid interface, reducing mass transfer limitations and enhancing absorption
 465 efficiency. On the other hand, the increment of the contact time from 0.6 to 10 s (at a
 466 constant L/G ratio of 7.5 L m⁻³) resulted in a strong RE decay, from 68.4 to 20.6%. At a
 467 constant L/G ratio, turbulences in gas-liquid contactor are increased at high gas flowrate,
 468 enhancing SO₂ mass transfer to the liquid bulk.

469 The residual alkalinity contained as dissolved carbonate in the bioprocess effluent was
 470 favorably used in the absorption unit to reduce reagents consumption and operational
 471 costs, as well as to improve SO₂ RE. Results obtained using specifically this absorbent
 472 for SO₂ removal showed an improvement on RE between a 25 and a 50%. Then, treating
 473 2000 ppm_v of SO₂ at a gas contact time of 0.6 s had associated a RE increase to 94.7 %

474 and 72.0 % at a L/G ratio of 15 and 5 L m⁻³, respectively. In the same way, SO₂ absorption
475 at a L/G ratio of 7.5 L m⁻³, and at gas contact time between 0.6 and 4 s, also presented
476 higher RE (ranging from 91.0 and 74.4 %). The performance of the bioprocess effluent
477 as an absorbent for SO₂ treatment was also evaluated at higher SO₂ concentrations. The
478 removal efficiencies for the treatment of 3000 ppm_v of SO₂ were only 8-9% lower than
479 the obtained for a flue gas of 2000 ppm_v of SO₂. Additionally, almost an identical
480 absorption efficiency was observed treating 4000 ppm_v of SO₂. These results highlighted
481 the suitability of residual alkalinity contained in the bioprocess effluent for SO₂
482 absorption, obtaining higher RE than using NaOH based absorbent. The reuse of
483 bioprocess waste as regenerated absorbent enabled the reduction of reagents consumption
484 in the absorption unit. In addition, buffer capacity of the regenerated absorbent allowed
485 higher efficiencies also for the treatment of high SO₂ loads. Therefore, the use of
486 bioprocess effluent as absorbent showed an excellent performance to reduce fluctuations
487 in scrubber RE due to variations in SO₂ inlet concentration.

488 **3.4 Economical feasibility of the SONOVA process implementation: the pigments** 489 **industry as a case study**

490 The integrated process for SO₂ bioscrubbing proposed in this work (SONOVA process)
491 has opened a window of opportunity to valorize sulfur contained in flue gases, which are
492 generated in this type of industrial activities. The technical feasibility of the integrated
493 SO₂ bioscrubbing process has been already demonstrated in this work; however, it is also
494 essential to assess the economic feasibility of the integrated process before it is
495 implemented at larger scales. For this purpose, a case study is proposed to calculate
496 investment costs, operational costs and the return on investment associated to a
497 hypothetical project. The operational conditions of the case study process are presented

498 in Table S3 (section S5, Supplementary Material), which are based on the findings
 499 reported in this work.

500 The biogas production from the UASB was also estimated to be 10% of the liquid effluent
 501 treated in the unit. The dimensions of the process and the treatment capacity could target
 502 the valorization of up to 20000 m³ h⁻¹ of flue gas, with SO₂ concentrations ranging from
 503 3000-4000 ppmv. Considering that any company generating flue gases with SO₂ must
 504 have already installed a scrubber to treat the emissions, the capital costs and operating
 505 costs of the S-rich effluent valorization process have been estimated in 2,021.000 USD
 506 and 1,471.897 USD per year, respectively (Aeris Tecnologías Ambientales, SL). Annual
 507 savings in sulfur, natural gas, sodium hydroxide and waste management have also been
 508 estimated in 400100 USD. This result means that the return on investment would be 5
 509 years, specifically for the case study proposed in this work. Table 3 shows a summary of
 510 the operational costs associated to the industrial activity considered in this case study
 511 before and after the implementation of SONOVA.

512

513 **Table 3.** Operational costs of an industrial activity producing 20000 m³ h⁻¹ of flue gas with SO₂
 514 concentration ranging from 2000 to 3000 ppm_v before the implementation of the SO₂ bioscrubber
 515 (case A) and after the implementation of the bioscrubber (case B)

Treatment technology	Concept	Consumption	Cost	Operational costs (USD year ⁻¹)
CASE A: Scrubber	Elemental sulfur	870 Tn year ⁻¹	150 USD Tn ⁻¹	130 500
	Natural Gas	1 871 323 Nm ³ year ⁻¹	0.354 USD Nm ³	662 448
	NaOH 50%	1529 Tn year ⁻¹	527 USD Tn ⁻¹	805 783
	Waste management	50605 m ³ year ⁻¹	5.4 USD m ⁻³	273 267
TOTAL ANNUAL COSTS				1 871 998 USD
CASE B: SONOVA	Elemental sulfur ¹	350 Tn year ⁻¹	150 USD Tn ⁻¹	525 000
	Natural Gas ²	1 697 223 Nm ³ year ⁻¹	0.354 USD Nm ³	600 817
	NaOH 50% ³	844 Tn year ⁻¹	527 USD Tn ⁻¹	445 038
	Waste management ⁴	0 m ³ year ⁻¹	5.4 USD m ⁻³	0
	Crude Glycerol	2 660 Tn year ⁻¹	100 USD Tn ⁻¹	266 002

Electricity ⁵	1 191 360 kW year ⁻¹	0.088 USD kW ⁻¹	104 840
TOTAL ANNUAL COSTS			1 471 897 USD

516 *¹Recovered sulfur from the SONOVA process is reused as a raw material for pigments production resulting*
517 *in operational savings. ²Methane produced in the UASB supplies part of the natural gas purchased for the*
518 *production process resulting in operational savings. ³Alkalinity contained in the recirculated effluent*
519 *reduces chemicals supply in the scrubber resulting in operational savings. ⁴S-rich effluent generated in the*
520 *scrubber is not treated through a waste management external company but valorized resulting in*
521 *operational savings. ⁵Electricity requirements associated uniquely to the implementation of the SONOVA*
522 *process mainly for pumping and aeration.*

523

524 **4. CONCLUSIONS**

525 Overall, the process proposed in this study to produce biosulfur from flue gases
526 containing SO₂ and crude glycerol has been demonstrated to be feasible from the
527 technological point of view. A polluted gas with SO₂ concentration ranging from 2000
528 ppm_v to 4000 ppm_v has been optimally treated in a spray-scrubber, obtaining RE higher
529 than 80 % at a gas contact time as low as 2 s, by reusing the effluent generated in the
530 biological stage (CSTR). Moreover, sulfur loads up to 9.3 kg S m⁻³ d⁻¹ were treated in the
531 UASB setting an HRT of 2h. The CSTR was fed with the effluent from the UASB and
532 operated at an HRT as low as 4h resulting in 100% of sulfide partial oxidation.

533 On the other hand, the economic analysis performed in this study showed that an
534 industrial activity generating 20000 m³ h⁻¹ of flue gas, with SO₂ concentration ranging
535 from 2000 to 3000 ppm_v, would have a ROI of 5 years with the implementation of the
536 integrated SO₂ bioscrubber. Moreover, the recirculation of the liquid effluent
537 (consumption of chemical agents reduced by almost 50 %) besides the recovery of
538 elemental sulfur from flue gases (60 % of the raw material recovered applying the
539 SONOVA process) makes this technology, not only technologically and economically
540 feasible, but also beneficial from the environmental point of view.

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