

1 **Quality attributes and shelf-life of vacuum packaged potato strips (*Solanum tuberosum*) as**  
2 **affected by edible coatings, thermal and non-thermal treatments**

3

4 R. D.A. Amaral<sup>1,2</sup>, I. Achaerandio<sup>1</sup>, B. C. Benedetti<sup>2</sup>, M. Pujolà<sup>1\*</sup>

5 <sup>1</sup>Department d'Enginyeria Agroalimentària i Biotecnologia, Escola Superior d'Agricultura de  
6 Barcelona, Universitat Politècnica de Catalunya BarcelonaTech. Barcelona, Spain

7 <sup>2</sup>Faculty of Agricultural Engineering, University of Campinas (UNICAMP), Brazil

8 Email address: [montserrat.pujola@upc.edu](mailto:montserrat.pujola@upc.edu) (M.Pujola)

9

10 **Abstract**

11 The aim of this work was to investigate the effect of using ultrasound or edible coating as a possible  
12 alternative to blanching on the quality of vacuum-packaged potato strips. The treatments assessed  
13 were blanching (85°C, 3.5 min), coating with 2 % alginate and sonicating (42 kHz, 5 min) in an  
14 ultrasonic bath containing a 2 % citric acid solution. Vacuum-packaged samples were stored up to 12  
15 days at 3 ±1°C. The pH, polyphenol oxidase (PPO) activity, sugars and microbial load were assessed.  
16 Also, the colour, shear-force and dry matter of the treated and fried potato strips as well as the oil  
17 adsorption and acrylamide after frying were evaluated. The PPO activity of the treated samples was  
18 not significantly different over time (p>0.05). The treatments applied did not affect the attributes of  
19 the fried potato strips over time; there were no significant changes in oil absorption, acrylamide  
20 content or colour (p>0.05). However, the visual quality of sonicated packaged potato strips was  
21 significantly better than that of the other treatments after storage. The loss of the texture of blanched  
22 potatoes was remarkable (p<0.05) before and after frying. Sonicated samples maintained mesophilic  
23 bacteria counts better than blanched and alginate coated vacuum-packaged potato strips.

24

25

26

27 **Keywords:** ultrasound, blanching, PPO, alginate, French fries.

28

## 29 **1. Introduction**

30 Several treatments have been used to replace the current methods of prolonging shelf life and reducing  
31 the loss in quality of fresh-cut products. Blanching is a method that is commonly used in minimally  
32 processed potatoes to prevent enzymatic browning by inactivating polyphenol oxidase, promoting a  
33 more uniform colour after frying, limiting oil absorption and improving texture (Severini, Baiano, De  
34 Pilli, Romaniello, & Derossi, 2003; Moreira, Castell-Perez, & Barrufet, 1999). However, it is also  
35 known that this thermal treatment could lead to a loss of firmness and a loss of other attribute, such as  
36 nutrients, flavour and colour (Alvarez, Canet, & Tortosa, 2001).

37 The use of edible coatings and non-thermal technologies, such as ultrasound, can be a good alternative  
38 to avoid or minimize this loss of quality. Studies involving the use of ultrasound for mango juice  
39 showed improvements in the carotenoid content and phenolic compounds, in addition to a reduction in  
40 the microbial load (Santhirasegaram, Razali, & Somasundram, 2013). These authors also stated that  
41 this treatment is a good alternative to a thermal treatment and indicates the combination with other  
42 thermal and non-thermal technologies. Ultrasound has been shown to reduce the loss of firmness of  
43 kiwifruit (Meng, Zhang, & Adhikari, 2014) and plum fruit during storage (Chen & Zhu, 2011).  
44 Similarly, Amaral, Benedetti, Pujola, Achaerandio, & Bachelli (2015) applied ultrasound to vacuum-  
45 packaged potatoes and found that this treatment affected the tuber microstructure, but did not affect  
46 the firmness, and no changes were observed in colour. The activity of PPO and POD decreases when  
47 ultrasound (40 kHz) is combined with ascorbic acid (1%) in fresh-cut apples stored for 12 days at 10°C  
48 (Jang & Moon, 2011). However, the effectiveness of ultrasound varies according to the frequency and  
49 power used, type of microorganism and enzyme, pH, temperature, and fruit or vegetable to which it is  
50 applied (Bilek & Turantaş, 2013; São José et al., 2014).

51 However, edible coatings are widely used in industry for whole and fresh-cut products to preserve  
52 quality. Alginate has been used to reduce weight loss and microbial load in carrots (Amanatidou,  
53 Slump, Gorris, & Smid, 2000), to maintain the quality and prolong the shelf life of fresh-cut apples  
54 (Rojas-Graü et al., 2007), and to reduce the stress of failure and browning during the storage of fresh-  
55 cut mangoes (Chiumarelli, Ferrari, Sarantópoulos, & Hubinger, 2011). Therefore, the association of  
56 alginate and nanomaterials can exhibit beneficial effects on the quality of shiitake mushrooms during

57 extended storage (Jiang, Feng, & Wang, 2013). Therefore, non-thermal processing technologies and  
58 edible coatings could be alternatives to thermal processing methods, but specific studies are needed for  
59 each product. For this reason, the aim of this work is to evaluate the effects of alginate and the  
60 combined use of ultrasound and citric acid as alternatives to blanching on the quality attributes of  
61 minimally processed potatoes and fried potatoes maintained under refrigeration for 12 days.

62

## 63 **2. Materials and methods**

64 Potatoes (*Solanum tuberosum* L. cv. Asterix) were acquired from Mercabarna (Mercado de  
65 Abastecimientos de Barcelona SA.). Tubers were selected, washed in tap water to remove surface dirt,  
66 dried and stored in the dark at a cool temperature ( $12\pm 2$  °C). Potato tubers that were free of defects  
67 were hand-peeled, cut into rectangular strips with a cross-section of 10x10 mm with a manual slicer  
68 and rinsed in distilled water. Any post-harvest chemical washing was applied. Then, strips were  
69 randomly assigned into four groups.

### 70 *2.1 Treatments*

71 Four treatments were applied to the raw material: i) for the control treatment, strips were rinsed in  
72 distilled water 1:4 (w/v) and ii) the strips were blanched in hot water, 1:4 (w/v) at 85°C for 3.5 minutes  
73 (Pedreschi, Moyano, Kaack, & Granby, 2005); iii) for the ultrasound treatment, the potato strips were  
74 dipped in an ultrasound bath (40 kHz frequency and 200 W of generation power, P-SELECTA  
75 3000617, Barcelona, Spain) filled with citric acid solution 2% (w/v) at 1:4 (w/v) for 5 minutes. The  
76 samples were stored for 10 minutes at room temperature for these treatments and then dried with paper  
77 towels. In the alginate treatment (iv), the strips were dipped in a coating solution with sodium alginate  
78 2% (w/v), glycerol 1% (w/v) and calcium lactate 2% (w/v) for 3 minutes (Chiumarelli et al., 2011).  
79 Coated strips were drained at  $16\pm 2$ °C for 1 hour to dry the coating material.

80 After the treatments, portions of  $100\pm 5$  g were vacuum-packed (-98 kPa, in a vacuum sealer VM-18  
81 ORVED S.p.A., Italy) in coextruded polyamide/high density polystyrene bags (Coex. PA/PEHD-  
82 70/150; thickness: 22  $\mu$ m; O<sub>2</sub> transmission rate: 8 cm<sup>3</sup>/m<sup>2</sup> dbar at 25°C) and stored at  $3\pm 1$ °C for up to  
83 12 days. Two replicates of  $100\pm 5$  g of vacuum-packaged potatoes were assessed for each treatment  
84 and sampling date.

85 The frying process was conducted in sunflower oil at  $180\pm 5^{\circ}\text{C}$  for 6 min (this time was fixed  
86 according to the palatability of the fried strips) in an electrical fryer (Taurus Professional Compact,  
87 Oliana, Spain) at a proportion of 4:1 (w:w) (Gökmen & Palazoğlu, 2009). After frying, the strips  
88 drained for 1 min and were then placed at room temperature on absorbent paper for 10 min to remove  
89 excess oil.

## 90 *2.2. Ultrasound equipment*

91 The ultrasound bath was made of a welded aluminium sheet, with a capacity of 9 L, and the  
92 dimensions were 12 cm x 46 cm x 12 cm (height x width x depth). The equipment had four steel cone-  
93 shape transducers (45 mm/38 mm in diameter; 47 mm length). The experiments were conducted in  
94 batch mode in a non-refrigerated system. The operating conditions had previously been optimized, and  
95 the distribution of the ultrasound in the bath was uniform (Amaral et al., 2015). The increase of the  
96 temperature in the water was less than  $2^{\circ}\text{C}$  after 5 min of treatment.

## 97 *2.3 Analytical determinations*

98 All of the analyses were carried out on days 1, 4, 8 and 12 for vacuum-packaged and fried potatoes.  
99 Vacuum-packaged potato samples (control, blanched, ultrasound and alginate treatments) were  
100 characterized in triplicate. For PPO activity, total sugar and acrylamide samples were collected,  
101 immediately frozen at  $-20^{\circ}\text{C}$  and subsequently freeze-dried at  $-54^{\circ}\text{C}$  and 0.07 mbar vacuum for 40 h  
102 by Telstar Cryodos -50 freeze dryer (1 KVA of potency, model 2G-6, Telstar, Barcelona, Spain).

103 The dry matter content was determined by drying 5 g of potato at  $65^{\circ}\text{C}$  for 24 h. (AOAC 931.04). The  
104 pH was measured with a potentiometer according to AOAC (981.12). Polyphenol oxidase (PPO)  
105 activity was determined in 3 g of lyophilized potato samples by determining the absorbance increase at  
106 410 nm over a period of 3 minutes. The results were expressed as units of enzymatic activity. One unit  
107 of PPO activity was defined as a change in absorbance at 400 nm per min and mL of enzymatic  
108 extract. The initial reaction rate was estimated from the linear portion of the plotted curve.

109 Sugars were extracted and measured by the procedure used in vegetables with slight modifications  
110 (Hernandez, Gonzalez-Castro, Alba, & de la Cruz Garcia, 1998). Samples consisting of 2 g of potato  
111 were extracted by refluxing for 30 min with 20 mL of 70% ethanol. The extract was vacuum-filtered,  
112 and the filtrate was filled to 25 mL with 70 % ethanol. A 5-mL aliquot of the solution was passed

113 through a Waters Sep-Pak C column, filtered (0.45  $\mu\text{m}$  pore size membrane), and then injected onto  
114 the HPLC (Hewlett Packard series 1100). The instrument was equipped with a Beckman 110B injector  
115 and a Beckman Refraction Index Detector (RID). Fructose, glucose and sucrose separation were  
116 performed using a Phenomenex Luna column (250 x 4.6 mm i.d.) at a constant temperature of 28°C  
117 using isocratic elution of acetonitrile-water (78:22 v/v). The flow rate was 1.2 mL·min<sup>-1</sup>. The average  
118 of the results of three replications was expressed as g·kg<sup>-1</sup> of lyophilized weigh (LW).

119 In fried potatoes, oil uptake determination consisted of a Sohlex extraction of the dried potato sample  
120 and gravimetric quantification of the oil content (AOAC 934.05), and dry matter was also determined  
121 under the same conditions for vacuum-packaged potatoes.

122 Acrylamide was assessed in fried potato strips following the extraction procedure used by Yang,  
123 Achaerandio & Pujolà (2016). GC analysis of the extract samples was performed on an AutoSystem  
124 gas chromatograph equipped with a flame ionization detector (FID) (Hewlett Packard 5890 series II).  
125 The column used was an Agilent HP-FFAP capillary (length = 25 m, i.d.= 0.2 mm, and film thickness  
126 = 0.3  $\mu\text{m}$ ), and the analysis conditions were as follows: the initial column temperature was set at 100  
127 °C for 0.5 min and then raised at a gradient of 10°C/min to 200 °C. The temperatures of the injector  
128 and detector were set to 250 and 260 °C, respectively; helium was used as the carrier gas at a flow rate  
129 of 1 mL/min, splitless for 1 min, and the injection volume was 1  $\mu\text{L}$ . The results were expressed as  
130  $\mu\text{g}\cdot\text{kg}^{-1}$  of lyophilized weight (LW).

### 131 *2.3 Physical properties*

132 The instrumental texture of the potato strips (vacuum-packaged and fried) was measured using a  
133 TA.TXplus texture analyser (Stable Microsystems Ltd., Godalming, UK) equipped with a 30-kg load  
134 cell and connected to a Warner Bratzler blade set at a speed of 1 mm·s<sup>-1</sup>. Twenty measurements of  
135 each treatment for each day were made. Shear force was measured as the maximum shear strength  
136 value and expressed as the maximum force (N).

### 137 *2.4 Colour evaluation*

138 The colour of the vacuum-packaged and fried potato strips was measured using a Minolta CR-400  
139 colourimeter (Osaka, Japan) in the CIE laboratory space. The L\* (lightness), a\* (greenness [-] to  
140 redness [+]), and b\* (blueness [-] to yellowness [+]) were recorded and evaluated. Five readings were

141 taken at two sites on the surface of the potato strips for each treatment and replicated. The parameters  
142 of hue angle ( $H^*$ ) and chroma ( $C^*$ ) were calculated as  $H^* = \tan^{-1}(b^*/a^*)$  and  $C^* = (a^{*2}+b^{*2})^{1/2}$ . Colour  
143 differences ( $dL^*$  and  $dH^*$ ) were calculated by subtracting the  $L^*H^*$  values for the sample from the  
144 control samples on the first day of analysis. The results were expressed as the mean value.

#### 145 *2.5 Microbiological analyses*

146 Analysis of Coliforms, Enterobacteriaceae, mesophilic aerobic microorganisms, yeast and moulds  
147 were carried out according to ISO 4832:2006, ISO 21528-2:2004, ISO 4833:2003 and ISO  
148 21527:2008, respectively. Two bags were evaluated for each treatment and each day of analysis.

#### 149 *2.6 Statistical analysis*

150 The data reported are the mean of duplicate experiments. A statistical study of variations after  
151 application of treatments and during storage was carried via two-way ANOVA using Minitab (v.17,  
152 MINITAB Inc., State College, PA) at a 95% confidence level. The differences between samples were  
153 determined using Tukey's least significant difference test. Pearson's correlation analysis was carried  
154 out to study the relationships between variables.

155

### 156 **3. Results and discussion**

#### 157 *3.1 Vacuum packaged potato strips*

158 Colour changes were evaluated by the lightness ( $L^*$ ) and Hue angle ( $H^*$ ) parameters, and colour  
159 differences were compared to the control on the first day of storage. The  $L^*$  values were between  
160 55.33 for alginate 2% treated samples at 12 days of storage and 66.02 for blanched samples after 4  
161 days of storage. The immersion of strips in sodium alginate solution promoted reduction of the  
162 lightness of the potatoes and, according to (Calder, Kash, Davis-Dentici, & Bushway, 2011), this is a  
163 signal of browning. The higher the negative difference, the higher the degree of browning obtained.  
164 Moreover, significant differences were observed for alginate 2 % after 8 days of storage ( $p<0.05$ , data  
165 not shown). Table 1 shows the lightness difference ( $dL^*$ ) between samples. There was an increase for  
166 blanching with storage, while alginate 2% had the opposite behaviour. Blanched potato samples  
167 showed higher values of the Hue angle ( $124.72^\circ$ ) immediately after processing and during the storage  
168 ( $p<0.05$ , data not shown). The Hue angle of the other samples of vacuum-packaged potatoes was

169 approximately 100°, as observed by (Fernandes, Soratto, Evangelista, & Nardin, 2010). The authors  
170 also stated that this value indicates a yellow pulp of the potato, which is desirable for frying. The Hue  
171 angle difference (dH\*) is especially notorious for blanching (Table 1), whereas in the alginate 2 % and  
172 US + citric acid treatments, there were no significant differences during the time of storage. This  
173 difference indicated the effect of blanching, resulting in a brownish yellow colour. Adams & Brown  
174 (2007) stated that the after-cooking darkening of potatoes may be due to a reaction in which a  
175 coloured chlorogenic acid-ferric iron complex is formed.

176 The dry matter (DM) content is very important for fried potato texture, and the use of this parameter  
177 has been found to be appropriate for predicting various texture attributes and for giving information  
178 about potatoes after the application of treatments (Arvanitoyannis, Vaitsi, & Mavromatis, 2008). In  
179 agreement with our results, Tajner-Czopek, Figiel, & Carbonell-Barrachina (2008) observed that the  
180 initial dry matter content of potato tubers suitable for frying was 208.1 g·kg<sup>-1</sup>, and the DM content of  
181 potato strips after blanching was 203 g·kg<sup>-1</sup>. In this study, the initial DM content varied from 175.2 to  
182 202.9 g·kg<sup>-1</sup>. No statistically significant differences were found (p <0.05), either between treatments  
183 or storage times, in the content of DM in vacuum-packaged potatoes (Table 1).

184 Initial values of the shear force of control vacuum-packaged potatoes were approximately 7 N (Table  
185 1). This value is slightly lower than the value found by Feltran, Lemos, & Vieites (2004) for the  
186 Asterix potato cultivar (8.15 N). As expected, the blanching treatment had the greatest effect on the  
187 texture of the vacuum-packaged potatoes. According to Alvarez, Canet & Tortosa (2001) and García-  
188 Segovia, Andrés-Bello, & Martínez-Monzó (2008), the blanching temperature had an influence on the  
189 softening and texture of potato tissue by starch gelatinization as well as on changes in pectic  
190 substances. The statistically significant decrease (p<0.05) of the shear force in the blanching treatment  
191 was maintained throughout the 12 days of storage at 3±1°C.

192 As expected in a cultivar suitable for French fries, the Asterix cultivar had a low reducing sugar  
193 content, less than 3 g·kg<sup>-1</sup> (FoodDrinkEurope, 2011). However, Rodriguez-Saona & Wrolstad (1997)  
194 and Fernandes et al. (2010) noted that the content of sugars is not enough to explain or predict the  
195 quality of fried potatoes. Fernandes et al. (2010) concluded that despite Markies and Mondial potato  
196 cultivars showing higher and lower sugar contents, respectively, these varieties presented an

197 acceptable colour after being fried. In our results, sucrose was the major sugar ( $0.40 - 0.24 \text{ g}\cdot\text{kg}^{-1}$ ),  
198 and the content of reducing sugars, in general, was slightly lower ( $0.11 - 0.22 \text{ g}\cdot\text{kg}^{-1}$ ) on the first day of  
199 storage (Table 1). In the case of sucrose, statistically significant differences ( $p < 0.05$ ) were found at  
200 four days of storage, with the sucrose content in the alginate treated samples significantly lower than  
201 in the blanching and control samples. In general, the storage time involved a reduction of the sucrose  
202 content in all of the treatments tested. This decrease can possibly be attributed to the hydrolytic  
203 processes of the disaccharides.

204 The initial pH of the vacuum-packaged potatoes was between 4.99 and 5.96 (US + citric 2 % and  
205 blanching, respectively). The use of citric acid at 2% in the ultrasonic bath involved lowering the pH  
206 of the potato samples to 0.72 ( $p < 0.05$ ), which was maintained during storage until the eighth day  
207 (Figure 1). This behaviour was in accordance with the results reported by Calder et al. (2011), who  
208 also observed a reduction of the pH of fresh-cut potato slices after 1 minute of immersion in solutions  
209 of sodium acid sulphate at 3% and citric acid at 3 %.

210 Blanching was the most effective treatment to inhibit polyphenol oxidase (PPO) activity. The  
211 application of US + citric acid 2% and alginate 2% treatments to potato strips involved non-significant  
212 differences in PPO activity. During storage, the PPO activity of coated samples was similar to that of  
213 the control samples, but there were statistical differences in the sonicated samples (Figure 1). The  
214 enzymatic activity of PPO can be limited by pH adjustment and/or the use of chelators or reducing  
215 agents (Girelli, Mattei, Messina, & Tarola, 2004). Contrary to this statement, in our results, there was  
216 no effect of US-citric acid 2% on the PPO activity of the treated samples. Wang et al. (2015) observed  
217 that a higher PPO activity does not always correlate with more browning. The same results were  
218 obtained by Cantos, Tudela, Gil, & Espn (2002) in potato strips (8 x 8 mm) stored at 4°C for 6 days.  
219 Jiang et al. (2013) used sodium alginate in mushrooms, and they observed similar patterns of PPO  
220 activity in the coated and control samples. Additionally, the authors obtained a positive synergetic  
221 effect of the alginate coating (2% and 3%) and 100% of  $\text{O}_2$  in the inhibition of PPO activity ( $p < 0.05$ ).

222 Sodium alginate 2 % was not an effective treatment for reducing the microbial load in vacuum-  
223 packaged potatoes (Figure 2). Nevertheless, the combination of US-citric acid 2 % was useful in the  
224 reduction of total coliforms, Enterobacteriaceae and aerobic bacteria. However, the reduction of yeasts



225 and moulds was lower. The inactivation of microorganisms could result from a combination of  
226 physical and chemical mechanisms, which occur during cavitation and cause the formation of free  
227 radicals and H<sub>2</sub>O<sub>2</sub> (Oyane et al., 2009). In a recent review, Cebrián, Mañas, & Condón, (2016) noted  
228 that the treatment medium pH had little or no influence on microbial resistance to manosonication and  
229 non-thermal UV treatment. From our results, the reduction of microbial counts may be due to the  
230 effect of cavitation in bacteria cells.

### 231 *3.2 Fried potato strips*

232 The frying process entailed an increase of dry matter content in the potato samples on the first day of  
233 storage (430.1-452.4 g·kg<sup>-1</sup>, Table 2). This increase was similar for all treatments and throughout the  
234 storage time; no statistically significant differences were found (p >0.05). The opposite behaviour was  
235 observed in the shear force of the fried potato. The shear force decreased with the frying process, but  
236 no significant differences between treatments and storage time were found. This finding is consistent  
237 with previous reports by Calder et al. (2011), who also did not find significant differences in the shear  
238 force of fried potatoes, although the control and the citric acid treated fried potato slices had a dried  
239 and case-hardened appearance on the surface of the raw fries over storage time.

240 The colour of the fried potatoes (evaluated with the L\* and H\* parameters) decreased after frying. The  
241 lightness on the first day of storage ranged between 60.35 to 54.68 and 90.54 to 83.15 for hue angle  
242 according to the treatment carried out. Non-significant differences were found between treatments  
243 (p<0.05). A positive difference of lightness for US + citric acid 2 % and negative differences of L\* for  
244 blanching, alginate treatment and the control were found (Table 2). The same behaviour was observed  
245 in the H\* parameters.

246 The frying process involved oil absorption in the potato strips and entailed, as previously discussed, an  
247 increase of DM. Moreno, Brown, & Bouchon (2010) noted that fried products reached the state of  
248 equilibrium after 10 min of cooling, when the competition between the drainage of oil from the  
249 surface and suction of oil from the crust had ceased. Our results are in agreement with their results. In  
250 our work, after frying, the potato strips were drained for 1 min and were then placed at room  
251 temperature for 10 min.

252 Controlling the frying time and oil temperature may reduce acrylamide formation because it could  
253 reduce Maillard reactions and minimize the hydrolysis of sucrose during frying (Medeiros Vinci,  
254 Mestdagh, & De Meulenaer, 2012). The content of acrylamide in fried potatoes was between 3.00 to  
255 4.42 mg·kg<sup>-1</sup> at the initial time and 2.89 to 3.81 mg·kg<sup>-1</sup> at 12 days of storage at 3±1°C. Non-  
256 significant differences were found in the acrylamide content in the treated fried potatoes and  
257 throughout storage time at 3°C for 12 days (p<0.05). These values are in agreement with the results  
258 obtained by Yang, Achaerandio, & Pujolà (2016) for French fries of the Kennebec and Agria  
259 cultivars, but were higher than those of the Red Pontiac cultivar.

### 260 *3.3 Correlation between parameters*

261 Significant Pearson correlations between parameters analysed in vacuum-packaged potato and fried  
262 potato strips are shown in Table 3. The study of Pearson correlations noted that the PPO activity was  
263 positively correlated (p < 0.01) with some of the physical and chemical parameters, such as pH, shear  
264 force, DM and hue angle (vacuum-packaged and fried potato strips), and negatively correlated with  
265 the sucrose content (p<0.05). However, the major presence of microorganisms in vacuum-packaged  
266 potatoes may promote the PPO enzymatic activity (p < 0.001; r = 0.858 and 0.804 for coliforms and  
267 Enterobacteriaceae, respectively).

268 Acrylamide formation was highly correlated with pH, dH\* and dL\*. The higher the content of  
269 acrylamide, the lower the lightness (L\*) and higher the hue angle (H\*) in fried potato strips. Related to  
270 pH, an increase in pH implied a decrease of the acrylamide content, as other works noted  
271 (FoodDrinkEurope, 2011).

272 A higher pH and DM content were observed in vacuum-packaged potatoes, and the absorption of oil  
273 in fried potatoes increased, while the shear force decreased (p<0.05).

274 The sucrose content in vacuum-packaged potato strips had opposite effects on the PPO activity and oil  
275 absorption (p< 0.05, r = -0.593, PPO activity and p<0.01, r = 0.932, oil absorption): a higher content  
276 of sucrose tended to increase oil absorption, but the PPO activity decreased. Similar effects on oil  
277 absorption were observed with the total sugar content (p<0.05 r = 0.838).

## 278 **4. Conclusions**

279 A non-thermal treatment (5 min, ultrasound bath 42 kHz) with citric acid 2% was an efficient  
280 treatment for maintaining the quality attributes of vacuum-packaged potato strips. Sonication coupled  
281 with an acid medium inhibited microbial growth (Enterobacteriaceae, coliforms and mesophilic  
282 bacteria) and maintained the visual appearance and texture of the samples up to 12 days of storage at  
283  $3\pm 1^{\circ}\text{C}$ . The use of an edible coating (sodium alginate 2%) did not favour the maintenance of the  
284 microbial quality and promoted the reduction of the lightness and increased the browning of the strips.  
285 The blanching treatment (3.5 min at  $85^{\circ}\text{C}$ ) was the treatment that had the higher inhibitory effect on  
286 PPO activity. However, blanching had significant and negative effects on the colour and shear force in  
287 vacuum-packaged potato strips. In general, none of the treatments assessed in this research involved  
288 significant differences in oil absorption, colour, shear force or acrylamide content in the fried potato  
289 strips.

290

## 291 **5. References**

- 292 Adams, J. B., & Brown, H. M. (2007). Discoloration in raw and processed fruits and vegetables.  
293 *Critical Reviews in Food Science and Nutrition*, 47(3), 319–333.  
294 <http://doi.org/10.1080/10408390600762647>
- 295 Alvarez, M. D., Canet, W., & Tortosa, M. E. (2001). Kinetics of thermal softening of potato tissue (cv.  
296 Monalisa) by water heating. *European Food Research and Technology*, 212(5), 588–596.  
297 <http://doi.org/10.1007/s002170100295>
- 298 Amanatidou, a., Slump, R. a., Gorris, L. G. M., & Smid, E. J. (2000). High Oxygen and High Carbon  
299 Dioxide Modified Atmospheres for Shelf-life Extension of Minimally Processed Carrots. *Journal*  
300 *of Food Science*, 65(JANUARY), 61–66. <http://doi.org/10.1111/j.1365-2621.2000.tb15956.x>
- 301 Amaral, R. D. A., Benedetti, B. C., Pujola, M., Achaerandio, I., & Bachelli, M. L. B. (2015). Effect of  
302 Ultrasound on Quality of Fresh-Cut Potatoes During Refrigerated Storage. *Food Engineering*  
303 *Reviews*, 7(2), 176–184. <http://doi.org/10.1007/s12393-014-9091-x>
- 304 Arvanitoyannis, I. S., Vaitis, O., & Mavromatis, A. (2008). Physico-chemical and sensory attributes in  
305 conjunction with multivariate analysis of two potato (*Solanum tuberosum* L.) cultivars after 90  
306 days of storage: An exploratory authentication study. *International Journal of Food Science and*

307 *Technology*, 43(11), 1960–1970. <http://doi.org/10.1111/j.1365-2621.2008.01799.x>

308 Bilek, S. E., & Turantaş, F. (2013). Decontamination efficiency of high power ultrasound in the fruit  
309 and vegetable industry, a review. *International Journal of Food Microbiology*, 166(1), 155–162.  
310 <http://doi.org/10.1016/j.ijfoodmicro.2013.06.028>

311 Calder, B. L., Kash, E. A., Davis-Dentici, K., & Bushway, A. A. (2011). Comparison of sodium acid  
312 sulfate to citric acid to inhibit browning of fresh-cut potatoes. *Journal of Food Science*, 76(3),  
313 164–169. <http://doi.org/10.1111/j.1750-3841.2011.02082.x>

314 Cantos, E., Tudela, J. A., Gil, M. I., & Espn, J. C. (2002). Phenolic Compounds and Related Enzymes  
315 Are Not Rate-Limiting in Browning Development of Fresh-Cut Potatoes Phenolic Compounds  
316 and Related Enzymes Are Not Rate-Limiting in Browning Development of Fresh-Cut Potatoes.  
317 <http://doi.org/10.1021/jf0116350>

318 Cebrián, G., Mañas, P., & Condón, S. (2016). Comparative Resistance of Bacterial Foodborne  
319 Pathogens to Non-thermal Technologies for Food Preservation. *Frontiers in Microbiology*,  
320 7(May), 1–17. <http://doi.org/10.3389/fmicb.2016.00734>

321 Chen, Z., & Zhu, C. (2011). Combined effects of aqueous chlorine dioxide and ultrasonic treatments  
322 on postharvest storage quality of plum fruit (*Prunus salicina* L.). *Postharvest Biology and*  
323 *Technology*, 61(2-3), 117–123. <http://doi.org/10.1016/j.postharvbio.2011.03.006>

324 Chiumarelli, M., Ferrari, C. C., Sarantópoulos, C. I. G. L., & Hubinger, M. D. (2011). Fresh cut  
325 “Tommy Atkins” mango pre-treated with citric acid and coated with cassava (*Manihot esculenta*  
326 Crantz) starch or sodium alginate. *Innovative Food Science & Emerging Technologies*, 12(3),  
327 381–387. <http://doi.org/10.1016/j.ifset.2011.02.006>

328 Feltran, J. C., Lemos, L. B., & Vieites, R. L. (2004). Technological quality and utilization of potato  
329 tubers. *Scientia Agricola*, 61(6), 593–597. <http://doi.org/10.1590/S0103-90162004000600006>

330 Fernandes, A. M., Soratto, R. P., Evangelista, R. M., & Nardin, I. (2010). Qualidade físico-química e  
331 de fritura de tubérculos de cultivares de batata na safra de inverno. *Horticultura Brasileira*,  
332 28(3), 299–304. <http://doi.org/10.1590/S0102-05362010000300010>

333 FoodDrinkEurope. (2011). Acrylamide Toolbox 2011. Confederation of the Food and Drink Industries

334 of the EU (FoodDrinkEurope), (September), 2–47.

335 García-Segovia, P., Andrés-Bello, A., & Martínez-Monzó, J. (2008). Textural properties of potatoes  
336 (*Solanum tuberosum* L., cv. Monalisa) as affected by different cooking processes. *Journal of*  
337 *Food Engineering*, 88(1), 28–35. <http://doi.org/10.1016/j.jfoodeng.2007.12.001>

338 Girelli, A. M., Mattei, E., Messina, A., & Tarola, A. M. (2004). Inhibition of Polyphenol Oxidases  
339 Activity by Various Dipeptides. *Journal of Agricultural and Food Chemistry*, 52(10), 2741–  
340 2745. <http://doi.org/10.1021/jf0305276>

341 Gökmen, V., & Palazoğlu, T. K. (2009). Measurement of evaporated acrylamide during frying of  
342 potatoes: Effect of frying conditions and surface area-to-volume ratio. *Journal of Food*  
343 *Engineering*, 93(2), 172–176. <http://doi.org/10.1016/j.jfoodeng.2009.01.011>

344 Hernandez, J. L., Gonzalez-Castro, M. J., Alba, I. N., & de la Cruz Garcia, C. (1998). High-  
345 performance liquid chromatographic determination of mono- and oligosaccharides in vegetables  
346 with evaporative light-scattering detection and refractive index detection. *Journal of*  
347 *Chromatographic Science*, 36(June), 293–298.

348 Jang, J.-H., & Moon, K.-D. (2011). Inhibition of polyphenol oxidase and peroxidase activities on  
349 fresh-cut apple by simultaneous treatment of ultrasound and ascorbic acid. *Food Chemistry*,  
350 124(2), 444–449. <http://doi.org/10.1016/j.foodchem.2010.06.052>

351 Jiang, T., Feng, L., & Wang, Y. (2013). Effect of alginate/nano-Ag coating on microbial and  
352 physicochemical characteristics of shiitake mushroom (*Lentinus edodes*) during cold storage.  
353 *Food Chemistry*, 141(2), 954–960. <http://doi.org/10.1016/j.foodchem.2013.03.093>

354 Medeiros Vinci, R., Mestdagh, F., & De Meulenaer, B. (2012). Acrylamide formation in fried potato  
355 products - Present and future, a critical review on mitigation strategies. *Food Chemistry*, 133(4),  
356 1138–1154. <http://doi.org/10.1016/j.foodchem.2011.08.001>

357 Meng, X., Zhang, M., & Adhikari, B. (2014). The Effects of Ultrasound Treatment and Nano-zinc  
358 Oxide Coating on the Physiological Activities of Fresh-Cut Kiwifruit. *Food and Bioprocess*  
359 *Technology*, 7(1), 126–132. <http://doi.org/10.1007/s11947-013-1081-0>

360

361 Moreira, R. G., Castell-Perez, M. E., & Barrufet, M. A. (1999). *Deep-fat Frying: Fundamentals and*

362 *Applications*. Gaithersburg: Aspen Publishers.

363 Moreno, M. C., Brown, C. A., & Bouchon, P. (2010). Effect of food surface roughness on oil uptake  
364 by deep-fat fried products. *Journal of Food Engineering*, *101*(2), 179–186.  
365 <http://doi.org/10.1016/j.jfoodeng.2010.06.024>

366 Oyane, I., Takeda, T., Oda, Y., Sakata, T., Furuta, M., Okitsu, K., ... Nishimura, R. (2009).  
367 Comparison between the effects of ultrasound and  $\gamma$ -rays on the inactivation of *Saccharomyces*  
368 *cerevisiae*: Analyses of cell membrane permeability and DNA or RNA synthesis by flow  
369 cytometry. *Ultrasonics Sonochemistry*, *16*(4), 532–536.  
370 <http://doi.org/10.1016/j.ultsonch.2009.01.001>

371 Pedreschi, F., Moyano, P., Kaack, K., & Granby, K. (2005). Color changes and acrylamide formation  
372 in fried potato slices. *Food Research International*. <http://doi.org/10.1016/j.foodres.2004.07.002>

373 Rodriguez-Saona, L. E., & Wrolstad, R. E. (1997). Influence of potato composition on chip color  
374 quality. *American Potato Journal*, *74*(2), 87–106. <http://doi.org/10.1007/BF02851555>

375 Rojas-Graü, M. A., Raybaudi-Massilia, R. M., Soliva-Fortuny, R. C., Avena-Bustillos, R. J., McHugh,  
376 T. H., & Martín-Belloso, O. (2007). Apple puree-alginate edible coating as carrier of  
377 antimicrobial agents to prolong shelf-life of fresh-cut apples. *Postharvest Biology and*  
378 *Technology*, *45*(2), 254–264. <http://doi.org/10.1016/j.postharvbio.2007.01.017>

379 Santhirasegaram, V., Razali, Z., & Somasundram, C. (2013). Effects of thermal treatment and  
380 sonication on quality attributes of Chokanan mango (*Mangifera indica* L.) juice. *Ultrasonics*  
381 *Sonochemistry*, *20*(5), 1276–1282. <http://doi.org/10.1016/j.ultsonch.2013.02.005>

382 São José, J. F. B. de, Andrade, N. J. de, Ramos, A. M., Vanetti, M. C. D., Stringheta, P. C., & Chaves,  
383 J. B. P. (2014). Decontamination by ultrasound application in fresh fruits and vegetables. *Food*  
384 *Control*, *45*, 36–50. <http://doi.org/10.1016/j.foodcont.2014.04.015>

385 Severini, C., Baiano, A., De Pilli, T., Romaniello, R., & Derossi, A. (2003). Prevention of enzymatic  
386 browning in sliced potatoes by blanching in boiling saline solutions. *LWT - Food Science and*  
387 *Technology*, *36*(7), 657–665. [http://doi.org/10.1016/S0023-6438\(03\)00085-9](http://doi.org/10.1016/S0023-6438(03)00085-9)

388

389 Tajner-Czopek, A., Figiel, A., & Carbonell-Barrachina, Á. A. (2008). Effects of potato strip size and

390 pre-drying method on french fries quality. *European Food Research and Technology*, 227(3),  
391 757–766. <http://doi.org/10.1007/s00217-007-0784-8>

392 Wang, Q., Cao, Y., Zhou, L., Jiang, C.-Z., Feng, Y., & Wei, S. (2015). Effects of postharvest curing  
393 treatment on flesh colour and phenolic metabolism in fresh-cut potato products. *Food Chemistry*,  
394 169, 246–254. <http://doi.org/10.1016/j.foodchem.2014.08.011>

395 Yang, Y., Achaerandio, I., & Pujolà, M. (2016). Influence of the frying process and potato cultivar on  
396 acrylamide formation in French fries. *Food Control*, 62, 216–223.  
397 <http://doi.org/10.1016/j.foodcont.2015.10.028>

398

**Table1**

Color differences, Dry matter content, Shear force, Reducing sugar and Sucrose content of vacuum packaged potato strips stored at 3±1°C for 12 days

Parameters	Treatments	Storage time			
		Day 1	Day 4	Day 8	Day 12
dL*	Control	-	-0.25	0.76	-0.28
	Blanching	-1.05	4.10	1.34	1.83
	US-citric 2%	0.88	-0.04	0.43	-0.65
	Alginate 2%	-5.82	-7.32	-5.65	-6.59
dH*	Control	-	1.27	0.89	0.47
	Blanching	24.33	24.33	27.34	26.29
	US-citric 2%	1.62	2.20	1.69	1.74
	Alginate 2%	0.94	1.21	0.52	0.69
DM g.kg <sup>-1</sup>	Control	192.0±11.3 Aa	175.8±1.5 Aa	172.5±2.1 Aa	190.3±4.0 Aa
	Blanching	202.9±1.6Aa	196.0±9.2 Aa	203.7±12.8 Aa	199.5±2.9 Aa
	US-citric 2%	185.6±6.0 Aa	185.7±17.5 Aa	182.2±22.7Aa	178.8±4.0 Aa
	Alginate 2%	175.2±11.6 Aa	156.7±16.3 Aa	155.4±23.6 Aa	171.5±20.8Aa
Shear force (N)	Control	7.00±0.05Aa	7.43±0.12Aa	7.07±0.17Aa	7.11±0.13Aa
	Blanching	5.19±0.36Ba	5.34±0.35Ba	5.90±0.66Aa	6.11±0.06Ba
	US-citric 2%	6.53±0.17Aa	6.68±0.15Aa	7.14±0.11Aa	6.81±0.26ABa
	Alginate 2%	6.87±0.02Aa	7.10±0.12Aa	6.87±0.01Aa	6.83±0.01ABa
Reducing sugars g.kg <sup>-1</sup> FW	Control	0.20±0.02Aa	0.18±0.06Aa	0.17±0.01ABa	0.10±0.04Ba
	Blanching	0.17±0.01Aa	0.11±0.03Aa	0.18±0.00ABa	0.19±0.01ABa
	US-citric 2%	0.23±0.05Aa	0.15±0.03Aa	0.14±0.02Ba	0.27±0.01Aa
	Alginate 2%	0.11±0.05Aa	0.13±0.03Aa	0.22±0.02Aa	0.13±0.01Ba
Sucrose g.kg <sup>-1</sup> FW	Control	0.40±0.01Aa	0.27±0.02Bb	0.27±0.02Ab	0.18±0.03Ac
	Blanching	0.39±0.02Aa	0.38±0.03Aa	0.31±0.08Ab	0.22±0.00Ac
	US-citric 2%	0.25±0.03Aa	0.25±0.01BCa	0.24±0.01Aa	0.17±0.02Ab
	Alginate 2%	0.24±0.07Aab	0.15±0.02Cc	0.16±0.01Ab	0.21±0.01Aab

US-citric 2% = Ultrasound bath with citric acid solution 2%, Alginate 2% = Sodium alginate 2% with glycerol 1% and calcium lactate 2%. Color parameters: dL\* =difference of Luminosity dH\*= difference of Hue angle. Data represents the mean ±standard error. Different capital letters in the same column indicates statistical differences (p<0.05). Different small letters in the same file indicates statistical differences (p<0.05).



**Table 2**

Physical parameters (dL\*,dH\* and Shear force ), dry matter content, oil absorption and acrylamide content in fried potato strips stored at 3±1°C for 12 days

Storage time	Treatments	dL*	dH*	DM g.kg <sup>-1</sup>	Shear force (N)	Oil Absorption g.kg <sup>-1</sup> FW	Acrylamide mg.kg <sup>-1</sup> FW
Day 1	Control	-	-	444.5±18.0Aa	1.27±0.19Aa	177.9±18.5Aa	3.93±0.82Aa
	Blanching	-1.99	-5.03	452.0±11.5Aa	1.14±0.22Aa	172.8±21.0Aa	3.80±0.69Aa
	US-citric 2%	1.46	1.51	452.4±29.8Aa	1.64±0.25Aa	155.9±2.2Aa	3.00±0.32Aa
	Alginate2%	-4.21	-5.88	430.1±17.9Aa	1.40±0.14Aa	155.1±1.9Aa	4.42±0.11Aa
Day 12	Control	-1.14	-2.08	449.1±2.7Aa	1.55±0.31Aa	146.2±1.1Aa	3.45±1.14Aa
	Blanching	-3.55	-9.03	489.2±31.8Aa	1.23±0.01Aa	159.3±12.5Aa	3.62±1.50Aa
	US-citric 2%	0.23	0.55	469.4±0.3Aa	1.91±0.20Aa	143.0±3.0Aa	2.89±0.25Aa
	Alginate2%	-2.49	-7.92	431.7±14.1Aa	1.76±0.04Aa	139.1±11.5Aa	3.81±1.10Aa

US-citric 2% = Ultrasound bath with citric acid solution 2%, Alginate 2% = Sodium alginate 2% with glycerol 1% and calcium lactate 2%. Color parameters: dL\* =difference of luminosity dH\*= difference of Hue angle. Data represents the mean ±standard error. Different capital letters in the same column indicates statistical differences (p<0.05). Different small letters in the same file indicates statistical differences (p<0.05).

**Table 3**

Significant Pearson correlations between the studied factors in vacuum-packaged potato and fried potatoes strips

Vacuum-packaged potato		Fried potato		
	PPO		Acrylamide	Oil absorption
dH*fried	0.703**	dH*fried	0.742*	-
H*	0.901**	dL*fried	-0.895*	-
pH	0.783**	pH	0.779*	0.703*
DM	0.692*	DM	-	0.932*
Shear force	0.842**	Shear force fried	-	-0.838*
Shear force fried	0.787**			
Sucrose	-0.593*	Sucrose	-	0,932**
		Total sugars	-	0.838*
Enterobacteriaceae	0.858**			
Coliforms	0.804**			

\*p< 0.05, \*\*p<0.01.

## Figure caption

**Figure1.** PPO activity (%) (bars) and pH (points) of vacuum-packaged potato strips stored at  $3\pm 1^\circ\text{C}$  along time.

Control (white bar), ultrasound + citric acid 2% (painted bar), alginate 2% (grey bar) and blanching (black bar).

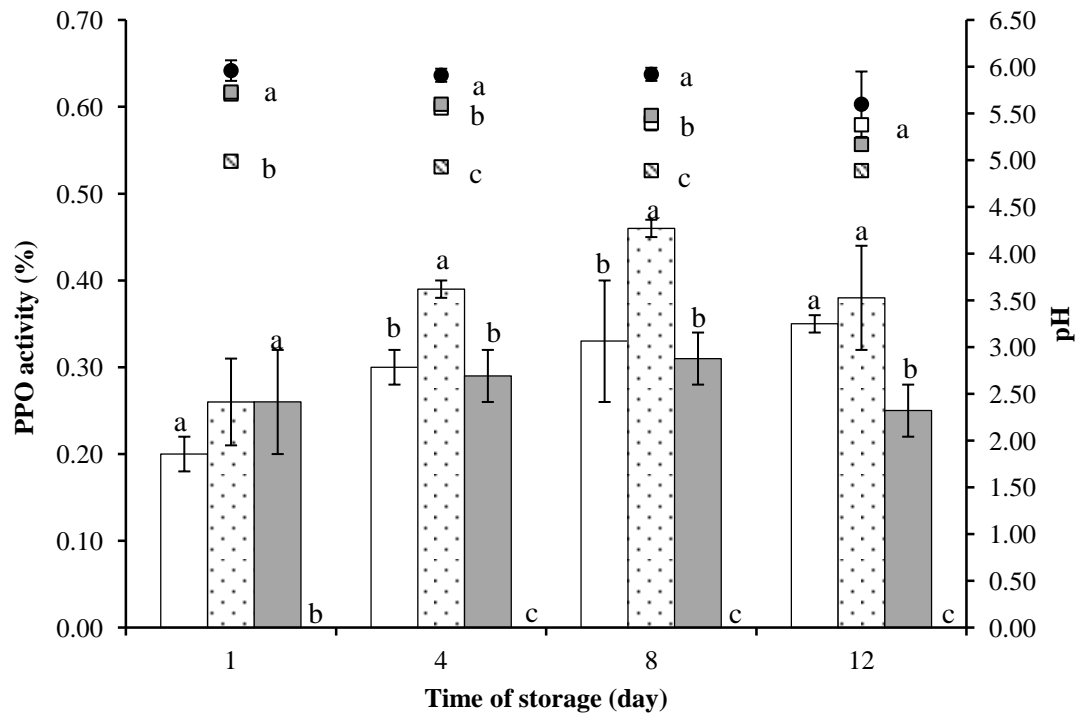
Values are the average of two experiments and triplicate samples  $\pm$  error deviation. Different lower-case letters indicate statically different groups by Tukey's test ( $p < 0.05$ ) between treatments the same day of analysis.

**Figure2.** Reduction of Mesophilic bacteria (A), Coliforms (B), Enterobacteriaceae (C) and yeast and moulds (D) counts ( $\log_{10}\text{UFC}\cdot\text{g}^{-1}$ ) of vacuum-packaged potato strips stored at  $3\pm 1^\circ\text{C}$  along time.

Control count at day 1 was used as reference value (1). Control (white bar), ultrasound + citric acid 2% (painted bar), alginate 2% (grey bar) and blanching (black bar).

Values are the average of two experiments and duplicate samples  $\pm$  error deviation. Different lower-case letters indicate statically different groups by Tukey's test ( $p < 0.05$ ) between treatments and day of analysis.

**Figure 1**



**Figure 2**

