

1 **CLASSIFICATION OF POTATO CULTIVARS TO ESTABLISH**
2 **THEIR PROCESSING APTITUDE**

3 Yali Yang, Isabel Achaerandio, Montserrat Pujolà *

4 *Department d'Enginyeria Agroalimentària i Biotecnologia, Escola Superior d'Agricultura de*
5 *Barcelona, Universitat Politècnica de Catalunya BarcelonaTech, Barcelona, Spain*

6 **Corresponding author at: Universitat Politècnica de Catalunya BarcelonaTech (UPC),*
7 *Departament d'Enginyeria Agroalimentària i Biotecnologia, Campus Baix Llobregat, Edifici D4,*
8 *C/Esteve Terradas, 8, 08860 Castelldefels, Barcelona, Spain. Tel.: +34 93 552 1089; FAX: +34 93*
9 *552 1121.*
10 *e-mail address: montserrat.pujola@upc.edu (M. Pujolà).*

11 **Abstract**

12 **BACKGROUND:** The commercial potato cultivars are diverse, not only in their physical
13 characteristics but also in their nutritional compositions and their content of functional
14 compounds (resistant starch (RS), total phenolic content (TPC) and antioxidant activity (AA)),
15 but there is little information about these differences. The aim of this study was to characterise
16 the nutritional value (focusing in carbohydrates and functional compounds) and instrumental
17 parameters of eight potato cultivars consumed in Spain and to determine whether these
18 parameters are useful for classifying the cultivars.

19 **RESULTS:** Significant Pearson's correlations were found due to the common and complex
20 interactions between the constituents of potatoes and their properties ($p < 0.05$). Principal
21 component analysis (PCA) revealed the correlations among the physicochemical properties, and
22 the first two principal components explained 56.84% of the variance among the cultivars studied.

23 **CONCLUSIONS:** The eight cultivars could be classified into three groups: (1) Red Pontiac,
24 Caesar, Kennebec, Agria and Cherie cultivars; (2) Agata and Monalisa cultivars and (3) Spirit
25 cultivar. Our results demonstrated that certain nutritional and functional parameters indicated
26 the potential efficacy of different cultivars to satisfy the nutritional needs of consumers,
27 improving the knowledge on the biochemical basis of potato processing to obtain higher quality
28 products.

29

30 **Key words:** Total starch (TS); resistant starch (RS); total phenol content (TPC); reducing sugars;
31 organic acids

INTRODUCTION

32

33 Potatoes (*Solanum tuberosum* L.) are a primary component of the human diet. More than 4000
34 varieties of potatoes are cultivated throughout the world. Despite the huge number of potato
35 varieties available, few cultivars have been commercialised. These potato varieties were
36 selected for their feasibility to be marketed and stored. A regular tuber size, high production
37 ratio, good storage ability, multi-purpose use and consumer acceptance are the main
38 characteristics of the potatoes that producers and sellers have chosen to improve their profits
39 and reduce the waste of potatoes. In addition, the food industry prefers cultivars that can be
40 stored for longer periods. In Spain, the rate of potato consumption is 23.13 kg per person per
41 year and involves mainly stored potatoes with a white skin. The predominant variety of potato
42 that is consumed is the Monalisa (26%) cultivar, followed by the Agata (16%) and Kennebec
43 (16%) cultivars, and then the Caesar (8%), Agria (4%) and Red Pontiac (3%) cultivars
44 [<http://www.magrama.gob.es>].¹

45 Freshly harvest potato tubers contain approximately 800 g·kg⁻¹ water and 200 g·kg⁻¹ dry matter.
46 The dry matter of potato tubers is composed of various substances, including starch, sugars,
47 nitrogenous compounds, lipids, organic acids, phenolic compounds, mineral substances and
48 non-starch polysaccharides. The starch content of potatoes varies with the cultivar and its
49 growth stage. As a major constituent of potatoes, starch contributes to the texture, consistency
50 and organoleptic qualities of foods prepared using them.² Starches are generally classified as a
51 rapidly digestible, slowly digestible or resistant starch (RS), depending on the rate and extent of
52 its digestion.³ RS is not digested in the small intestine, therefore reaching the colon, resulting in
53 the production of short-chain fatty acids, particularly butyric acid produced by the fermentation
54 of resistant starch. RS was recently recognised as a valuable contributor to dietary fibre intake
55 that is better tolerated than are other soluble fibres. The total sugar content of a potato is
56 approximately 0.05 g·kg⁻¹ of fresh weight, which is in the form of reducing monosaccharides,
57 such as D-glucose and D-fructose, and non-reducing disaccharides, such as sucrose.⁴ Low
58 storage temperatures may cause an increase in the sugar content of most potato cultivars. The

59 organic acids in potato tubers are carbonaceous compounds with high metabolic activity, and
60 some of them are related to each other. The main organic acids in potato tubers are citric, malic,
61 tartaric, oxalic, fumaric and succinic acids. The content of citric acid in potato tubers is higher
62 than that of the other acids, and it plays an important role as an antioxidant by inhibiting the
63 oxidative enzymatic browning process.⁵

64 The phenolic compounds are members of a large group of minor plant products that play an
65 important role in determining the organoleptic properties. In potato tubers, the phenolic
66 compounds are located mainly in the skin and adhesive tissue cortex, and the concentrations of
67 phenolic components decrease toward the centre of the tuber.⁶ However, potato skins are often
68 wasted as by-product of the potato-processing industry (i.e. the frying potato chips) or by
69 consumers. Lopez-Cobo et al.⁷ studied the phenolic compounds of potato skin and stated that the
70 potato skin could be merit more attention. Physicochemical properties contained in potato tubers
71 that may act as antioxidants in the human diet have been intensively studied.^{8,9} High positive
72 correlations between the level of antioxidant activity (AA) and the total phenol content (TPC)
73 were found by Reyes et al.¹⁰ who stated that phenolic compounds were mainly responsible for
74 the antioxidant activity of potatoes. The antioxidant activities of potatoes vary among the potato
75 genotypes with varying flesh colours, and purple potatoes tend to be associated with a high level
76 of total antioxidant activity.¹¹

77 In summary, potatoes are a healthy component of a varied diet because they are a relatively low-
78 calorie food containing a variety of dietary fibre nutrients and antioxidants. However, there is
79 scarce information about the correlations of textural parameters, carbohydrate components,
80 phenolics and antioxidant capacity of the potato cultivars commonly consumed. The main
81 purposes of this study were (1) to evaluate the **instrumental parameters**, carbohydrates and
82 functional content of eight commercial cultivars consumed in Spain, focusing on the starch
83 profile (RS, TS), the functional content (TPC, AA), **colour and texture** and (2) to establish
84 possible correlations between nutrients as a distinguishing feature of different potato varieties
85 for determining the aptitude of being processed.

87 **Chemicals**

88 All the chemicals and solvents used were of analytical grade. Oxalic acid, fumaric acid, tartaric
89 acid, malic acid, citric acid, lactic acid, D-(+)-fructose, D-(+)-glucose, sucrose, gallic acid 1-
90 hydrate, Folin-Ciocalteu reagent, sodium dihydrogen phosphate, sodium carbonate anhydrous,
91 HPLC-grade acetonitrile and 37% (v/v) chloridric acid were purchased from Panreac Applichem
92 (IWT group, Barcelona, Spain). Methanol and anhydrous di-sodium hydrogen phosphate were
93 supplied by Scharlau Chemie (Scharlab S.L. Barcelona, Spain). Thermostable amyloglucosidase
94 (GOPOD) reagent buffer, GOPOD reagent enzymes, D-(+)-glucose standard solution,
95 amyloglucosidase and pancreatic- α -amylase were purchased from Megazyme International
96 (Ireland). Fluorescein, 2, 2-azobis (2-amidinopropane) dihydrochloride (AAPH) and 6-hydroxy-
97 2, 5, 7, 8-tetramethylchromane-2-carboxylic acid (Trolox) were obtained from Acros Organics
98 (Thermo Fisher Scientific Inc. Waltham, MA, USA).

99

100 **Sample preparation**101 *Potato samples*

102 Potato tubers (*Solanum tuberosum* L.) of eight cultivars that are commercialised in Spain (Agata,
103 Agria, Caesar, Cherie, Kennebec, Monalisa, Red Pontiac and Spirit) were obtained from
104 Mercabarna (Mercados de Abastecimientos de Barcelona S.A., Barcelona, Spain). All the
105 potatoes were grown in Europe and had the same post-harvest storage conditions. Randomly
106 chosen samples of 10 kg of each potato cultivar were provided. Table 1 shows the physical
107 characteristics of the potato cultivars. The average weight of the tubers ranged from 134 to 337
108 g, except for the Cherie cultivar, the weight of which was less than 100 g. The colour of the skin
109 of the tubers was white-yellow, except for that of the Red Pontiac and Cherie cultivars, which
110 was red. The flesh colour ranged from white to yellow. Two of the cultivars (Agria and Caesar)
111 were defined as mealy, which might be due to their high starch content, and the rest (Agata,

112 Cherie, Kennebec, Monalisa, Red Pontiac and Spirit) were defined as multi-purpose (useful for
113 boiled and baked products).

114 Each selected potato tuber was washed and then, potatoes were hand-peeled and the potato flesh
115 was cut into small strips (1×1×5 cm). The potato flesh strips and the skin peels were lyophilised
116 using a freeze-drying instrument, Cryodos-45 (Terrasa, Spain), and packed in plastic bags, and
117 the lyophilised powders were maintained at -20 °C until further use.

118 *Extraction of phenolic compounds, sugars and organic acids*

119 Methanol extracts were obtained using the procedure described by Andre et al,¹² with slight
120 modifications. In brief, the lyophilised powder of each sample was mixed and chilled, and 1 g
121 was removed for extraction. Solid-liquid extraction was performed using 10 mL of acidified
122 methanol/water (80:20 v/v) containing 0.1 g/L HCl by sonication (Bandeline Sonoplus GM70,
123 Germany) for 14 min in an ice bath. After sonication, the extracts were centrifuged at 2.313 xg
124 for 10 min at 4 °C (Selecta, Spain). The supernatant was filtered through Whatman No.1 filter
125 paper, and the pellet was re-extracted as before. Two liquid extracts were mixed and were
126 evaporated to dryness using a rotary evaporator at 40 °C (Laborata 4000 Efficient, Germany).
127 The organic residue was brought to 5 mL with acidified water containing 0.1 g/L HCl and was
128 stored at -20 °C until further analysis. All the samples were extracted in triplicate. The potato
129 skin and flesh extracts were used for analysis of the phenolic compound content and antioxidant
130 capacity. The flesh extracts were also used for the measurement of sugar and organic acid
131 content.

132

133 **Instrumental analysis**

134 *Colour evaluation*

135 The colour of the potato samples was measured using a MINOLTA CR-400 colorimeter
136 (Minolta camera, Osaka, Japan) in the CIE lab space. The L* (lightness), a* (greenness [-] to
137 redness [+]), and b* (blueness [-] to yellowness [+]) values were evaluated at three different
138 positions. The values for the parameters of hue angle (H°) and chroma (C*) were calculated as

139 follows: $H^{\circ} = \tan^{-1}(b^*/a^*)$ and $C^* = (a^{*2} + b^{*2})^{1/2}$. Six measurements were taken for each potato
140 sample, and the results were expressed as the mean values \pm standard deviation.

141 *Texture analysis: Texture profile and shear force analyses*

142 A texture profile analysis (TPA) of fresh samples was performed using a 75 mm diameter
143 cylinder plunger probe (P/75). Six potato cylinders (length 10 mm, diameter 10mm) were
144 analysed for each potato sample, The following parameters were set: test speed of $0.83 \text{ mm} \cdot \text{s}^{-1}$,
145 a rest period of 5 s between the two cycles and a trigger force of 5 g. The maximum extent of
146 deformation was 40% of the original length.¹³ Four TPA parameters were determined from each
147 curve as described by Bourne,¹⁴: hardness (peak force of the first compression cycle),
148 cohesiveness (ratio of positive force area during the second compression to that during the first
149 compression), springiness (ratio of the time duration of force input during the second
150 compressing to that obtained during the first compression), and chewiness (hardness multiplied
151 by cohesiveness multiplied by springiness). Six potato cylinders were analysed for each potato
152 sample, and the results were expressed as the mean value \pm standard deviation.

153 The flesh shear force of the potato cultivars was measured using a texture analyser (TA.XT plus,
154 Stable Microsystems, Surrey, UK) equipped with a Wartner-Blazer probe, as previously
155 described.¹⁵ The test conditions were: test speed of $1 \text{ mm} \cdot \text{s}^{-1}$, target distance of 22 mm into the
156 samples and trigger force of 2 g. The potato strips were cut vertically by the probe, and the shear
157 force was set as the maximum force and expressed in N. Six potato strips were tested for each
158 potato sample, and the results were expressed as the mean values \pm standard deviation.

159

160 **Potato component analysis**

161 *Dry matter and pH*

162 The dry matter content was determined using the gravimetric method (AOAC 931.04). The
163 results were expressed as $\text{g} \cdot \text{kg}^{-1}$. The pH was determined according to AOAC method 981.12.
164 All the analyses were conducted in triplicate, and the results were expressed as the mean values
165 \pm standard deviation.

166 *Determination of the total starch and resistant starch contents*

167 The total and resistant starch contents were analysed through their enzymatic hydrolysis to D-
168 glucose using the AOAC 996.11 (amyloglucosidase/ α -amylase method) and AOAC 2002.02
169 methods, respectively. D-(+)-Glucose was oxidised to D-gluconate, which was quantitatively
170 measured from the absorbance at 510 nm of a colorimetric reactant. The results of the total and
171 resistant starch analyses were expressed as $\text{g}\cdot\text{kg}^{-1}$ of the lyophilised weight (LW). Each sample
172 was analysed in triplicate.

173 *Sugar analysis*

174 The content of sugars in the potato samples was analysed using a high-performance liquid
175 chromatography (HPLC) Hewlett Packard series 1100 instrument equipped with a Beckman
176 110B injector and a Beckman Refraction Index Detector (RID). The separation was performed
177 using a Phenomenex Lunacolumn (250 x 4.6 mm i.d.), following the method of Rodriguez-
178 Galdon et al.¹⁶ Briefly, the liquid extracts were filtered through a 0.45 μm membrane filter, and
179 20 μL of each filtrate was injected. The mobile phase consisted of acetonitrile/water (78:22 v/v),
180 and the flow rate was $1.2\text{ mL}\cdot\text{min}^{-1}$. Individual sugars were identified by comparing their
181 retention times with those of external standards, and the quantification was conducted through
182 calibration using the external standard. Each extract was analysed in triplicate. The sugar
183 contents were expressed as $\text{g}\cdot\text{kg}^{-1}$ of LW.

184 *Organic acid analysis*

185 The organic acid contents were determined using the same protocol used for analysis of the
186 sugar contents, except that detection was conducted using a Beckman 166 UV-Vis detector set
187 to 210 nm, as described by Rodriguez-Galdon et al.¹⁶ The organic acid contents were expressed
188 as $\text{g}\cdot\text{kg}^{-1}$ of LW.

189 *Total phenolic content analysis*

190 The total phenolic content of the extracts of potato skin and flesh was determined using a
191 modified version of the Folin-Ciocalteu assay.¹⁷ Gallic acid was used as the standard and was
192 diluted with distilled water to obtain the range of concentrations appropriate for a standard curve.

193 20 μL of sample extract, gallic acid calibration standard or the blank material was placed in a
194 plastic cuvette with 1.58 mL of distilled water and 100 μL of Folin-Ciocalteu reagent. After 5
195 min of incubation, 300 μL of 200 $\text{g}\cdot\text{L}^{-1}$ sodium carbonate was added, and the solution was
196 incubated for 2 h at room temperature. Then, the absorbance at 765 nm was measured using the
197 Nicolet Evolution 300 spectrophotometer (Thermo electron Corporation, UK). The results were
198 expressed as g Gallic acid equivalent per kg of LW ($\text{g GAE}\cdot\text{kg}^{-1}$ LW).

199 *Analysis of the antioxidant activity of the potato skin and flesh extracts*

200 The antioxidant activity of the extracts was evaluated using the oxygen-radical absorbance
201 capacity (ORAC) assay, as described by Gorjanovic et al.¹⁸ Briefly, 13.5 μL of each diluted
202 sample or standard solution (Trolox at five different concentrations, ranging from 6.25 to 500
203 μM) was mixed with 135 μL of 9.57×10^{-2} μM fluorescein in a well of a 96 cell plate, and the
204 plate was incubated at 37 $^{\circ}\text{C}$ for 10 minutes; then, 50 μL of 153 mM AAPH (2, 2-azobis (2-
205 amidinopropane) dihydrochloride) was added to each well to initiate the reaction. The
206 fluorescence was measured every 2 min for 120 min at 37 $^{\circ}\text{C}$ using emission and excitation
207 wavelength of 485 and 530 nm, respectively, using a microplate fluorescence reader (SynergyTM
208 Multi-detection Microplate Reader, BIO-TEK Instruments, USA) until the decay of the kinetic
209 curve was completed. The area under the curve (AUC) and the net AUC ($\text{AUC}_{\text{standard}} - \text{AUC}_{\text{blank}}$)
210 was calculated for each standard or sample. A blank measurement was taken and a linear
211 standard curve (5 standards) was created for every run. The results were expressed as g of
212 Trolox equivalents (TE) per kg of LW. The reagents, standards and diluted samples were
213 prepared using phosphate buffer, pH 7.40. Each extract was analysed in triplicate.

214

215 **Statistical analyses**

216 The data reported were the mean values of the results of triplicate analyses. The coefficient of
217 variation of the chemical data was less than 5%. The variations among the contents of the potato
218 components were evaluated using a one-way analysis of variance (ANOVA). The differences
219 between mean values were evaluated using the HSD Tukey test with a 95% confidence interval.

220 Pearson's correlation analysis was conducted to study the relationships among the variables.
221 Both analyses were conducted using Minitab 16 Statistical software (MINITAB Inc., State
222 College, PA, USA). A principal component analysis (PCA) was conducted to determine the
223 relationships among the variables that were analysed using STAT-ITCF statistical software
224 (Bordeaux, France).

225 **RESULTS AND DISCUSSION**

226 **Instrumental parameters**

227 *Colour characteristics*

228 Colour is an important quality parameter for raw potato tubers that is affected by certain pre-
229 harvest and post-harvest factors and by the type of cultivar. The chroma and hue-angle values of
230 the tubers of the different potato cultivars varied considerably (Table 2). The chroma values of
231 the yellow and light-yellow tubers were higher than those of the white tubers, with the exception
232 of the Spirit tuber. The tuber chroma values increased from the Red Pontiac to the Agria (13.47
233 to 30.29, respectively) cultivars. Our results were similar to those of Cabezas-Serrano et al,¹⁹
234 who found that the chroma values of the tubers of five industrial potato cultivars ranged from
235 17.1 to 31.0. Among the cultivars analysed, the highest Hue-angle value was obtained for Agata
236 tubers (light yellow), followed by Monalisa (light yellow) and Spirit (white) tubers, and the
237 lowest value was obtained for Cherie (yellow) tubers. The Hue-angle values of the tubers of the
238 other cultivars were not significantly different ($p>0.05$).

239 *Texture: TPA and shear force*

240 The values obtained from the TPA of the raw potatoes varied considerably among the eight
241 cultivars (Table 2). Their hardness ranged from 123.89 N to 156.80 N, and these results were
242 consistent with those of Bordoloi et al.²⁰ Spirit showed a greater hardness and shear force,
243 whereas Agata showed the lowest hardness and shear force of the tested cultivars. Therefore,
244 significant correlation was found between the shear force and hardness ($r=0.838$; $p<0.05$). In
245 addition, the Spirit cultivar also had the highest cohesiveness, springiness and chewiness values
246 in all cultivars. The values of cohesiveness and springiness of all cultivars tested were smaller

247 than other parameters, but these values were significantly different ($p < 0.05$). The shear force
248 also significantly correlated with chewiness ($r = 0.708$; $p < 0.05$). The differences in this textural
249 property among the potato cultivars may be due to their differing dry-matter content.²¹ The
250 texture is a very important parameter in cooking quality of potatoes. Mealiness was used to
251 describe texture of boiled, baked and oven-fried potatoes, while hardness was also judged
252 regarding quality.²² According to Kaur et al,²³ potato cultivars with greater mealiness showed
253 higher hardness and cohesiveness. The texture changes during the cooking are associated with
254 the gelatinization and retrogradation behaviour of starch and with enzymatic and non-enzymatic
255 changes of pectin.²²

256

257 **Dry matter and carbohydrate contents of the commercial potatoes**

258 The dry-matter content of the potatoes ranged from $230 \text{ g} \cdot \text{kg}^{-1}$ for the Spirit cultivar to $184 \text{ g} \cdot \text{kg}^{-1}$
259 for the Agata cultivar. All the cultivars, except the Agata variety, contained more than $200 \text{ g} \cdot \text{kg}^{-1}$
260 of dry matter, and significant differences in the dry-matter contents of the cultivars were found,
261 as shown in Table 3 ($p < 0.05$). Usually, commercial varieties of potatoes are selected according
262 to their dry-matter content, but this parameter can be affected by the storage and growing area. **S**
263 Stored Agata and Agria tubers from Italy had similar contents of dry matter,¹⁹ but Agria tubers
264 purchased in New Zealand had a higher content of dry matter.²⁰

265 Starch is the main component of dry matter, with a concentration of approximately $600 \text{ g} \cdot \text{kg}^{-1}$.²⁴
266 The content of total starch, which is also the major carbohydrate component of potatoes, ranged
267 from 681 (Agata) to $752 \text{ g} \cdot \text{kg}^{-1}$ of LW (Spirit), as shown in Table 3. The cultivated potatoes had
268 a mean content of $188 \text{ g} \cdot \text{kg}^{-1}$ of starch on a fresh weigh basis (FW) that ranged from 100 to 300
269 $\text{g} \cdot \text{kg}^{-1}$.²⁵ Among the studied potatoes with the starch content lower than that of the mean value,
270 the tubers of the Spirit and Agata cultivars contained $173 \text{ g} \cdot \text{kg}^{-1}$ and $125 \text{ g} \cdot \text{kg}^{-1}$ of starch at FW.
271 The total starch contents observed in this study were similar to those reported by Bordoloi et
272 al.²⁰ for four commercial varieties.

273 The resistant starch in raw potatoes is defined as RS type 2 (RS2), which comprises

274 ungelatinised resistant granules that are slowly hydrolysed by α -amylase.²⁶ In this study, the
275 content of resistant starch of the potatoes ranged from 482 (Agria) to 610 g·kg⁻¹ (Cherie) of LW
276 (Table 3). Previous reports noted that fresh potatoes contain 691-764 g·kg⁻¹ of RS,^{27,28} but there
277 was scarce data about the differences among potato cultivars. Our results showed that the
278 potatoes of the cultivars studied contained less RS than was previously reported and there were
279 significant differences between the RS contents of the cultivars ($p < 0.05$). Notably, the fraction
280 of resistant starch comprised 65-85% of the total starch in the potatoes of the eight cultivars
281 assessed.

282 Significant differences were found in the sugar content of the potatoes of the eight cultivars
283 tested ($p < 0.05$). The concentration of glucose (1.20-23.50 g·kg⁻¹ LW) was slightly higher than
284 fructose in all of the cultivar samples. Their fructose content ranged from 0.80 to 14.1 g·kg⁻¹ LW,
285 and sucrose content ranged from 1.40 to 4.5 g·kg⁻¹ LW. Plata-Guerrero et al.²⁹ analysed the
286 content of glucose, fructose and sucrose of the potatoes of four commercial potato cultivars, and
287 they also found that the content of glucose (0.28-1.20 g·kg⁻¹ FW) was higher than that of
288 fructose (0.27-0.90 g·kg⁻¹ FW) and sucrose (0.26-0.40 g·kg⁻¹ FW). These results may be due to
289 the high concentration of fructokinase in potatoes; this enzyme could redirect fructose into the
290 hexose-phosphate pathway.³⁰ Our data showed that the reducing sugars (glucose and fructose)
291 content of the tubers ranged from 0.20 to 37.6 g·kg⁻¹ LW and the reducing sugars content of the
292 Agata and Monalisa tubers were significantly higher than those of the other cultivars. Wegener
293 et al.³¹ analysed a variety of coloured potato cultivars and found that the content of reducing
294 sugars in their tubers ranged from 0.26 to 0.75 g·kg⁻¹ FW. Whereas the cultivar type and storage
295 conditions had a notable effect on the equilibrium between free sugars and starch in tubers,
296 Kumar et al.³⁰ did not observe changes in the concentration of reducing sugars in tubers during
297 the first three days of storage at low temperatures (4-6 °C). Endo et al.³² observed that during
298 storage at temperatures lower than 8 °C, the content of reducing sugars increased markedly in all
299 the cultivars studied. Furthermore, reducing sugars affect potato processing such that the higher
300 the reducing sugars content, the higher the level of browning after frying. Additionally,

301 Muttucumaru et al.³³ proposed that in potatoes, sugars are more important than free asparagine
302 as acrylamide precursors because other amino acids present in potatoes may play a role in the
303 formation of this contaminant. Therefore, the tubers of commercialised cultivars that develop a
304 high content of reducing sugars during storage may be a potential risk for acrylamide formation
305 during cooking (i.e. baking or frying).

306

307 **Correlations among the carbohydrate content, dry-matter content and texture of**
308 **commercial potatoes**

309 Pearson's correlations were calculated to evaluate the relationships among the components of
310 the potatoes. The total starch content of the potatoes was highly positively correlated with their
311 dry matter content ($r=0.722$; $p<0.01$), in agreement with the results of Bordoloi et al.²⁰ However,
312 a less significant correlation was found between the dry matter and resistant starch content of
313 the potatoes ($r=0.441$; $p=0.031$), which may be due to the content of resistant starch being
314 dependent on the physicochemical properties of the starch.³⁴ Finally, the level of correlation
315 between the contents of resistant starch and total starch of the potatoes ($r=0.417$; $p<0.01$) was
316 similar to that found for the RS and DM contents.

317 As expected, the glucose content was significantly positively correlated with the fructose
318 content ($r=0.993$; $p<0.01$). There were negative correlations between the reducing sugar content
319 and content of total starch ($r=-0.515$; $p<0.01$) and dry matter ($r=-0.727$; $p<0.05$).

320 The potatoes that had higher starch and higher dry-matter contents (such as Spirit potatoes) were
321 observed to be much harder and more cohesive than those with low levels of these components.

322 A significantly positive Pearson's correlation was found between the hardness and the shear
323 force levels and the dry matter content of raw potato tubers ($r=0.824$; $p<0.05$ and $r=0.915$;
324 $p<0.01$, respectively), whereas the correlations between the hardness and the shear force levels
325 and the total starch content were also high ($r=0.700$ and 0.734 , respectively; $p<0.05$).

326

327 **Organic acids and pH**

328 As Table 4 shows, the pH values of the tubers ranged from 5.89 to 6.30. The correlation
329 between the organic acid contents and the pH level was significant ($r= 0.555$; $p=0.03$). The
330 organic acids are biologically important because they participate in various metabolic processes,
331 such as the Krebs cycle. These compounds affect the acidity and pH of potatoes, depending on
332 their concentration and their pKa.¹⁶

333 The concentrations of the six most abundant organic acids (oxalic, tartaric, malic, lactic, citric
334 and fumaric acids) in the potato tubers are listed in Table 4. Citric acid was the most abundant
335 organic acid in all the tested potato cultivars. The ranges of the oxalic, tartaric, malic and lactic
336 acids contents were similar; however, significant differences in the organic acid contents of the
337 cultivars were found ($p<0.05$). Fumaric acid was the least abundant organic acid, and there were
338 no significant differences in its content among the cultivars ($p>0.05$). Rovers and Guttman³⁵
339 reported citric acid contents ranging from 6.0 to 20 $\text{g}\cdot\text{kg}^{-1}$ DW, which is consistent with our
340 finding of a range of 7.1 to 11.3 $\text{g}\cdot\text{kg}^{-1}$ DW; the malic acid content ranged from 1.0 to 6.0 $\text{g}\cdot\text{kg}^{-1}$
341 DW, which was slightly higher than our results (with a range of 0.7 to 1.3 $\text{g}\cdot\text{kg}^{-1}$ DW). The
342 tartaric acid and oxalic acid contents found in our study are similar to those of Wichrowska et
343 al.⁵ and Rodriguez-Galdon et al,¹⁶ respectively. None of the previous studies reported the
344 presence of lactic acid.

345 The concentrations of individual organic acid differed among the potato cultivars. In general,
346 Agata potatoes presented the higher content of citric and lactic acid, and Agria potatoes had the
347 higher content of tartaric and oxalic acid. The higher content of malic and fumaric acids were
348 found in Caesar potatoes. Organic acids (lactic acid, citric acid, oxalic acid, tartaric acid and
349 fumaric acid) have been described as strong antimicrobial agents against psychrophilic,
350 mesophilic microorganisms in fresh-cut fruit and vegetables (including potatoes).³⁶ The
351 decreasing of citric acid in potato tubers, as well tendency of the boiled potatoes to darken, was
352 due to the non-enzymatic process of the antioxidants.³⁷

353 The Pearson's correlation analysis showed a positive and significant correlation between the

354 oxalic and tartaric acid contents ($r=0.768$; $p<0.05$), the citric and oxalic acid contents ($r=0.661$;
355 $p<0.05$) and the AA level and total acid content ($r=0.565$; $p<0.05$), but no significant
356 correlations ($p<0.05$) were found between the total organic acid content and the values of the
357 other parameters that were studied.

358

359 **Total phenolic content**

360 The phenolic content of potatoes depends primarily on the genotype and the growing location,
361 but certain methods of extraction and sample-preparation, such as vigorous extraction methods,
362 can lead to an apparent increase in the phenolic content.³⁸ According to Ji et al,³⁹ with the
363 exception of a few cultivars, the selected potatoes had higher concentrations of phenolic in the
364 skin than in the flesh, often twice as much. Potato skins are a rich source of phenolic
365 compounds, but much of the phenolic content of potatoes is wasted in the manufacture of some
366 potato products, such as potato chips. The effect of the cultivars on the total phenolic content
367 and the differences between the total phenolic contents of skin and flesh are shown in Table 5.

368 The total phenolic content (TPC) of the potato skins ranged from 3.8 to 6.7 g GAE·kg⁻¹ LW.
369 These results were in accordance with those of Wu et al,⁴⁰ who reported that the total phenolic
370 content of potato skins ranged from 0.76 to 7.88 g GAE·kg⁻¹ of dry weight. In general,
371 significant differences between the TPC of the cultivars were found ($p<0.05$). Apart from the
372 Caesar and Agata cultivars, the white-yellow skinned cultivars had a lower content of TPC
373 compared with that of the red-skinned cultivars, as expected. Several studies have reported that
374 red or purple skinned potatoes contained higher amounts of phenolic compounds (anthocyanins)
375 than the yellow-skinned cultivars.⁴¹ The high phenolic content of some white-yellow skinned
376 cultivars (Caesar and Agata) may be explained by the differences in their phenolic compound
377 profiles.

378 As shown in Table 5, the TPC of the potato flesh ranged from 0.89±0.02 (Kennebec) to
379 1.73±0.04 (Agata) g GAE·kg⁻¹ LW, which was lower than those reported by Ji et al (1.60 to 8.40
380 g GAE·kg⁻¹ DW).³⁹ Ah-Hen et al.⁴² analysed Shepody and Desiree cultivars and found that the

381 TPC of peeled potatoes varied from 1.91 to 18.6 g GAE·kg⁻¹ DW, which was also higher than
382 the range we observed. The TPC of different potato cultivars clearly varied widely. According to
383 Teow et al,¹¹ the purple-fleshed sweet potato clones had the highest total phenolic content,
384 followed in order by the orange-, yellow- and white-fleshed clones. Negative correlations were
385 found between the DM and TS contents and the TPC content (-0.585, p<0.05 and -0.456,
386 p<0.05, respectively).

387

388 **Antioxidant activity**

389 There are some difficulties in measuring the true antioxidant status of food products in vivo.
390 The antioxidant activities (AA) of foods or food components have been evaluated using in-vitro
391 chemical models. There are two major mechanisms to explain how antioxidants deactivate free
392 radicals (hydrogen-atom transfer and single-electron transfer).⁴² Two assays of the AA are
393 generally used, the ORAC (oxygen radical-absorption capacity) assay to determine the
394 hydrogen-atom transfer activity and the FRAP (ferric reducing/antioxidant power) assay to
395 determine the single-electron transfer activity. However, the ORAC assay is considered more
396 closely related to a human biological assay.⁴³

397 The antioxidant activities (AA) of the potato skin and flesh extracts are presented in Table 5.
398 The AA in the flesh and skin extracts of the potato cultivars ranged from 4.83 to 10.79 and from
399 26.03 to 77.04 g TE·kg⁻¹ LW, respectively, and significant differences between the values for
400 different cultivars were found (p<0.05). The flesh extracts of the yellow and light-yellow
401 cultivars exhibited significant higher AA compared with those of the white-flesh potato cultivars.
402 The highest AA was found in the Agata flesh sample (10.79 g TE·kg⁻¹ LW). Apart from the
403 potato, other foods, such as sorghum and sweet potato, also show a similar relationship between
404 the colour intensity and the AA.¹¹

405 The Pearson's correlations between the total phenolic contents (TPC) and ORAC values of the
406 skin and flesh samples were evaluated. The results showed positive correlations between the
407 ORAC values of the potato skins and their TPC (r=0.984; p<0.01) and between the ORAC

408 values of the potato flesh samples and their TPC ($r=0.659$; $p<0.01$). High correlations between
409 the TPC and AA have been reported for various food commodities, such as sorghum ($r=0.971$),
410 cactus pear ($r=0.970-0.990$) and sweet potato cultivars ($r=0.937$).^{11,44} These high correlations
411 result from the contribution that the main phenolic compounds of potato tubers make to their
412 hydrophilic antioxidant capacity. Additionally, phenolic compounds can transfer an oxygen
413 molecule to the peroxy radical, which is the basis of the peroxy radical reaction evaluated in
414 the ORAC assay, which involves a hydrogen-atom transfer mechanism.¹² Therefore, the TPC
415 can be used as an indicator in assessing the AA of fruits and vegetables, including the tubers of
416 potato cultivars.

417

418 **Principal component analysis**

419 Principal component analysis (PCA) revealed that fourteen principal components (PC1 to PC14)
420 explained the variance among the data, and the first two principal components explained 56.84%
421 of the total variance (Figure 1). The first principal component explained 38.31% of the total
422 variance, and the second principal component explained 18.53% of the total variance, which
423 indicated that the first two principal components concerned variables that differentiated the
424 studied cultivars. The TS, DM, RS and texture parameters (Shear force (She), Hardness (Har),
425 Chewiness (Che) and Cohesiveness (Coh)) had positive loadings and the AA, TPC, reducing
426 sugar, total acid and colour parameter (Hue-angle) had negative loadings for the first principal
427 component. The AA, DM and Har had negative loadings and other properties which showed in
428 Figure 1 had positive loadings for the second principal component.

429 The scores of the eight potato cultivars for the first and second principal components are shown
430 in Figure 2. The distribution of the cultivars along the first principal component reflected their
431 DM, TS, She and Har. The tubers of the Spirit, Kennebec, Cherie and Caesar cultivars had
432 positive scores for the first principal component, indicating their higher DM, TS, She and Har
433 values. In contrast, the tubers of the Agata, Agria and Monalisa cultivars which had negative
434 scores for the first principle component, had higher antioxidant activity and reducing sugar

435 content. Furthermore, the distribution of the cultivars along the second principal component
436 reflected their Hue angle, springiness (Spr), Coh and Che. Regarding the first and second
437 principal components, the Spirit cultivar had higher positive scores for both the first and second
438 principle components, which indicated that the Spirit cultivars had the higher DM, TS, She, Har,
439 Spr, Coh, Che and Hue angle values than that of other cultivars tested.

440 According to PCA, the eight commercial potato cultivars were classified into 3 groups: the
441 Cherie, Kennebec, Agria, Caesar and Red Pontiac cultivars into group 1; the Agata and
442 Monalisa cultivars into group 2 and the Spirit cultivar into group 3. The physical and chemical
443 properties of different group cultivars were distinct. The cultivars of group 1 had higher dry
444 matter, total starch content, shear force and hardness; the group 2 cultivars had higher
445 antioxidant activity, reducing sugar content, Hue angle, springiness, cohesiveness and chewiness;
446 the group 3 cultivar had higher dry matter, total starch, shear force, hardness, springiness,
447 cohesiveness, chewiness and Hue angle. Certain nutritional and functional parameters of
448 different group cultivars indicated the potential efficacy of different cultivars to satisfy the
449 nutritional needs of consumers and industrial use. The group1 and group 3 cultivars were
450 suitable for frying because of the higher dry matter and the lower reducing sugar content,
451 enabling to obtain a less colored product and low levels of acrylamide content which was
452 attributed to the Maillard reactions;³³ The group 2 cultivars were fit for boiling and baking
453 because of higher value of springiness, cohesiveness and chewiness, and the cultivars without
454 peeling during boiling and baking may increase the antioxidant properties due to health
455 benefits.¹⁰ The texture attributes are important for describing the variation of industrial use
456 between the analyzed cultivars. As one of the textural parameters, the hardness could indicant
457 the aptitude use of potato cultivars. Kreutzmann et al.²² found that the frying potato cultivars
458 with higher mealiness which significantly and positively correlated with hardness had higher
459 scores; the boiling and baking potato tubers with lower mealiness had higher scores. As Figure 2
460 showed, the group 1 and group 3 cultivars had higher hardness than the group 2 cultivars, which
461 also proved that the group1 and group 3 cultivars were fit for frying and group 2 for boiling and

462 baking. In general, our study showed that the physicochemical properties analysed enable to
463 classify the potato cultivars and predict the aptitude use of the potato cultivars.

464

CONCLUSIONS

465 A statistical study of correlations between all parameters analysed was conducted to discover
466 associations between measured pairs of these parameters. A significant ($p < 0.05$) and positive
467 correlations were found which due to the common and complex interactions. The potato skin
468 recommended for use without removing during the cooking process (boiling and baking) was
469 due to the higher of total phenolics and antioxidant activity. The organic acids content was
470 similar to the previously reported, but the acid lactic was presented in all the cultivars studied.

471

472 The PCA results classified the eight cultivars studied into three groups: (1) Cherie, Kennebec,
473 Agria, Caesar and Red Pontiac; (2) Agata and Monalisa; (3) Spirit. The physical and chemical
474 properties of three group cultivars were notable different, which determined the aptitude of the
475 potato cultivars on being processing. The group 1 and group 3 cultivars were suitable for frying
476 which was due to the higher hardness, dry matter and the lower reducing sugar content, while
477 group 2 cultivars were fit for boiling and baking because of the higher value of springiness,
478 cohesiveness and chewiness, and the cultivars without peeling increased the antioxidant
479 properties. These properties analysis and the groups may explain and reinforce the cooking type
480 of the cultivars proposed by the European Cultivated Potato database.

481

ACKNOWLEDGEMENTS

482 This work was supported in part by the China Scholarship Council (File number.
483 201206990014).

484

485

REFERENCES

486 1 Ministerio de Agricultura Alimentación y Medio Ambiente. Estudio de la cadena de
487 valor y formación de precios del sector de la patata fresca de consumo. Campaña 2010 (2012)
488 [<http://www.magrama.gob.es>]

- 489 2 Tharanathan M and Tharanathan RN, Resistant starch in wheat-based products:
490 isolation and characterization. *J Cereal Sci* **34**: 73-84 (2001).
- 491 3 Song W, Janaswamy S and Yao Y, Structure and vitro digestibility of normal corn starch:
492 Effect of acid treatment, autoclaving, and β -amylolysis. *J Agri Food Chem* **58**: 9753-9758
493 (2010).
- 494 4 Lisinska G and Leszczynski W, *Potato science and technology*. Elsevier Applied
495 Science. London and New York: 1-140 (1989).
- 496 5 Wichrowska D, Rogozinska I and Pawelzik E, Concentrations of some organic acids in
497 potato tubers depending on weed control method, cultivar and storage conditions. *Polish J Env*
498 *S* **3**: 487-491 (2009).
- 499 6 Friedman M, Chemistry, biochemistry and dietary role of potato phenolics. *J Agri Food*
500 *Chem* **45**: 1523-1540 (1997).
- 501 7 Lopez-Cobo A, Gomez-Caravaca AM, Cerretani L, Segura-Carretero A and Fernandez-
502 Gutierrez A, Distribution of phenolic compounds and other polar compounds in the tuber
503 of *Solanum tuberosum L.* by HPLC-DAD-q-TOF and study of their antioxidant activity. *J*
504 *Food Compos Anal* **36(1)**: 1-11 (2014).
- 505 8 Lachman J and Hamouz K, Red and purple colored potatoes as a significant antioxidant
506 source in human nutrition-a review. *Plant Soil Environ* **51**: 477-482 (2000).
- 507 9 Brown CR, Antioxidants in potato. *American J Potato Res* **82(2)**: 163-172 (2005).
- 508 10 Reyes LF, Miller JC and Cisneros-Zevallos L, Antioxidant capacity, anthocyanins and
509 total phenolics in purple- and red-fleshed potato (*Solanum tuberosum L.*) genotypes. *American*
510 *J Potato Res* **82**: 271-277 (2005).
- 511 11 Teow CC, Truong V, Mcfeeters RF, Thompson RL, Pecota KV and Yencho GC,
512 Antioxidant activities, phenolic and β -carotene contents of sweet potato genotypes with
513 varying flesh colors. *Food Chem* **103**: 829-838 (2007).
- 514 12 Andre CM, Oufir M, Hoffmann L, Hausman JF, Rogez H, Larondelle Y and Evers D,
515 Influence of environment and genotype on polyphenol compounds and in vitro antioxidant

516 capacity of native Andean potatoes (*Solanum tuberosum* L.). *J Food Compos Anal* **22**: 517-
517 524 (2009).

518 13 Alvarez MD and Canet W, Principal component analysis to study the effect of
519 temperature fluctuations during storage of frozen potato. *Eur Food Res and Techn* **211**: 415-
520 421 (2000).

521 14 Bourne MC, Texture profile analysis. *Food Technol* **32**: 62-66 (1978).

522 15 Singh J and Kaur L, *Advances in potato chemistry and technology*. Elseviser,
523 Amsterdam, pp. 147, 153, 155, 157 (2009).

524 16 Rodriguez-Galdon B, Rios MD, Rodriquez EM and Diaz RC, Influence of the cultivar
525 on the organic acid and sugar composition of potatoes. *J Sci Food Agr* **90**: 2301-2309 (2010).

526 17 Singleton VL, Orthofer R and Lamuela-Raventos RM, Analysis of total phenols and
527 other oxidation substrates and antioxidants by means of Folin-Ciocalteau reagent. *Methods*
528 *Enzymol* **299**: 152-178 (1999).

529 18 Gorjanovic SZ, Alvarez-Suarez JM, Novakovic MM, Pastor FT, Pezo L, Battino M and
530 Suznjevic DZ, Comparative analysis of antioxidant activity of honey of different floral sources
531 using recently developed polarographic and various spectrophotometric assays. *J Food*
532 *Compos Anal* **30**: 13-18 (2013).

533 19 Cabezas-Serrano AB, Amodiob ML, Cornacchiab R, Rinaldib R and Colelli G,
534 Suitability of five different potato cultivars (*Solanum tuberosum* L.) to be processed as fresh-
535 cut products. *Postharvest Biol and Tech* **53**: 138-144 (2009).

536 20 Bordoloi A, Kaur L and Singh J, Parenchyma cell microstructure and textural
537 characteristics of raw and cooked potatoes. *Food Chem* **133**: 1092-1100 (2012).

538 21 Casanas R, Gonzalez M, Rodriguez E, Marrero A and Diaz C, Chemometric studies of
539 chemical compounds in five cultivars of potatoes from Tenerife. *J Agr Food Chem* **50**: 2076-
540 2082 (2002).

- 541 22 Kreutzmann S, Bassompierre M, Thybo AK, Buch L and Engelsen SB, Exploratory
542 study of potato cultivar difference in sensory and hedonistic applicability tests. *Potato Res* **54**:
543 13-28 (2011).
- 544 23 Kaur L, Singh N, Sodhi NS and Gujral HS, Some properties of potatoes and their
545 starches I. Cooking, texture and rheological properties of potatoes. *Food Chem* **79(2)**: 177-181
546 (2002).
- 547 24 Liu Q, Tarn R, Lynch D and Skjoldt NM, Physicochemical properties of dry matter and
548 starch from potatoes grown in Canada. *Food Chem* **105**: 897-907 (2007).
- 549 25 Camire ME, Kubow S and Donnelly DJ, Potatoes and human health. *Crit Rev Food Sci*
550 **49**: 823-840 (2009).
- 551 26 Fuentes-Zaragoza E, Riquelme-Navarrete MJ, Sanchez-zapata E and Perez-Alvarez JA,
552 Resistant starch as functional ingredient: a review. *Food Res Int* **43**: 931-942 (2010).
- 553 27 Lante A and Zocca F, Effect of β -cyclodextrin addition on quality of precooked vacuum
554 packed potatoes. *LWT-Food Sci Tech* **43**: 409-414 (2010).
- 555 28 Garcia-Alonso A and Goni I, Effect of processing on potato starch: In vitro availability
556 and glycaemic index. *Starch-Starke* **52**: 81-84 (2000).
- 557 29 Plata-Guerrero PR, Hernandez GE and Villanova GB, Determination of reducing sugars
558 and asparagine in potatoes. *J Liquid Chrom Related Tech* **32**: 2556-2568 (2009).
- 559 30 Kumar D, Singh BP and Kumar P, An overview of the factors affecting sugar content of
560 potatoes. *A. Appl. Biol.* **145**: 247-256 (2004).
- 561 31 Wegener CB, Jansen G, Jurgens HU and Schutze W, Special quality traits of coloured
562 potato breeding clones: Anthocyanins, soluble phenols and antioxidant capacity. *J Sci Food*
563 *Agr* **89**: 206-215 (2009).
- 564 32 Endo CM, Ohara-Takada A, Chuda Y, Ono H, Yada H, Yoshida M, Kobayashi A,
565 Tsuda S, Takigawa S, Noda T, Yamauchi H and Mori M, Effects of storage temperature on the
566 contents of sugars and free amino acids in tubers from different potato cultivars and
567 acrylamide in chips. *Biosci, Biotechn and Biochem* **70(5)**: 1173-1180 (2006).

- 568 33 Muttucumaru N, Powers SJ, Elmore JS, Briddon A, Mottram DS and Halford NG,
569 Evidence for the complex relationship between free amino acid and sugar concentrations and
570 acrylamide-forming potential in potato. *Annals of Applied Biology* **164**: 286-300 (2014).
- 571 34 Noda T, Takigawa S, Matsuura-Endo C, Suzuki T, Hashimoto N, Kottarachchi NS,
572 Yamauchi H and Zaidul ISM, Factors affecting the digestibility of raw and gelatinized potato
573 starches. *Food Chem* **110**: 465-470 (2008).
- 574 35 Rovers PJW and Guttman TK, Analysis of organic acids in potato waste water. *Food*
575 *Chem* **45**: 283-287(1992).
- 576 36 Bari ML, Ukuku DO, Kawasaki T, Inatsu Y, Isshiki K and Kawamoto S, Combined
577 efficacy of bisin and pediocin with sodium lactate, citric acid, phytic acid, and potassium
578 sorbate and EDTA in reducing the listeria monocytogenes population of inoculated fresh-cut
579 produce. *J Food Protect* **68**: 1381-1387 (2005).
- 580 37 Lisinska G and Aniolowski K, Organic acids in potato tubers: Part 1- the effect of
581 storage temperatures and time on citric and malic acid content of potato tubers. *Food Chem* **38**:
582 255-261 (1990).
- 583 38 Rumbaoa R, Cornago D and Geronimo I, Phenolic content and antioxidant capacity of
584 Philippine potato (*Solanum tuberosum*) tubers. *J Food Compos Anal* **22(6)**: 546-550 (2009).
- 585 39 Ji X, Rivers L, Zielinski Z, Xu M, Macdougall E, Stephen J, Zhang S, Wang Y,
586 Chapman RG, Keddy P, Robertson GS, Kirby CW, Embleton J, Worrall K, Murphy A,
587 Koeyer DD, Tai H, Yu L, Charter E and Zhang J, Quantitative analysis of phenolic
588 components and glycoalkaloids from 20 potato clones and in vitro evaluation of antioxidant,
589 cholesterol uptake, and neuroprotective activities. *Food Chem* **133**: 1177-1187 (2012).
- 590 40 Wu T, Yan J, Liu R, Marccone MF, Aisa HA and Tsao R, Optimization of microwave-
591 assisted extraction of phenolics from potato and its downstream waste using orthogonal array
592 design. *Food Chem* **133**: 1292-1298 (2012).
- 593 41 Perla V, Holm D and Jayanty SS, Effects of cooking methods on polyphenols, pigments
594 and antioxidant activity in potato tubers. *Food Sci and Tech* **45**: 161-171 (2012).

595 42 Ah-Hen K, Fuenzalida C, Hess S, Contreras A, Vega-Calvez A and Lemus-Mondaca R,
596 Antioxidant capacity and total phenolic compounds of twelve selected potato landrace clones
597 grown in Southern Chile. *Chilean J Agr Res* **72(1)**: 2-9 (2012).

598 43 Ronald L, Prior, Xianli W and Schaich K, Standardized methods for the determination
599 of antioxidant capacity and phenolics in foods and dietary supplements. *J Agr Food Chem* **53**:
600 4290-4302 (2005).

601 44 Stintzing FC, Herbach KM, Mosshammer MR, Carle R, Yi W, Sellappan S, Akoh CC,
602 Bunch R and Felker P, Color, betalain pattern, and antioxidant properties of cactus pear
603 (*Opuntia* spp.) clones. *J Agr Food Chem* **53**: 442-451(2005).